

# Rejection of the monotypic status of *Peromyscus furvus* (Rodentia: Cricetidae), with consequences for its species group

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Previous studies using Cytochrome-*b* or ND3-ND4 mitochondrial gene have yielded intriguing evidence about the phylogenetic relationships among populations of *Peromyscus furvus*; however, those studies each based on phylogenies for a single type of genes, yielded conflicting topologies. In addition, analyses with traditional morphometrics have revealed differences in skull size among certain populations of the species. Therefore, in order to reassess the systematic and taxonomic status of *P. furvus*, we incorporated a suite of genetic and morphometric characters and employed cladistic analyses. Herein, we present results mostly derived from our genetic analyses (results from the phylogenetic examination of skull size and shape will appear later). Phylogenetic analyses were conducted using four mitochondrial genes (Cytb and ND3-ND4L-ND4) with the respective data analyzed separately or combined, followed by an analysis with genetic and morphometric data (size and shape characters). Most phylogenetic constructions were made with parsimonious methods, but probabilistic methods also were used in the analyses with the genes separated by type. Similar topologies were recovered from all analyses of the Cytb gene and from all parsimony analyses of the NADH genes; however, conflicting topologies were obtained with the probabilistic methods for the NADH genes. Additionally, to better understand the genetic variation in each type of gene, analyses for genetic divergence were conducted within and among genetic groups and haplotype networks were constructed. All the topologies obtained using genetic data questioned the monotypic status of *P. furvus*, as two additional clades were identified that seemingly correspond to unrecognized entities. The first of these, *P. latirostris*, occurs in the northern region and could be considered as either a species or subspecies. An unknown *Peromyscus species nova* that occurs to the south is considered as a valid species. Further, *P. furvus* s. s. becomes a polytypic species by recognizing at least two subspecies (*P. f. angustirostris* and *P. f. furvus*). Phylogenetic analyses also rejected membership of *P. melanocarpus* and *P. ochraventer* within the *furvus* species group. Instead, *P. melanocarpus* showed a greater affinity to *P. mexicanus totontepecus*, whereas, *P. ochraventer* either joined to the clade containing *P. melanocarpus* and *P. m. totontepecus* or to *Megadontomys cryophilus* in a sister clade. Finally, *Osgoodomys banderanus* (subgenus *Haplomylomys*) always remained basally positioned and segregated from all members of the subgenus *Peromyscus*.

Estudios previos con los genes mitocondriales citocromo-*b* y ND3-ND4, han arrojado evidencia intrigante acerca del estado monotípico de *Peromyscus furvus*; sin embargo cada uno de esos estudios basados en filogenias para un solo tipo de genes, produjeron topologías conflictivas entre ellos. Además, análisis con morfometría tradicional han revelado diferencias en el tamaño del cráneo entre ciertas poblaciones de la especie. Por ende, con el fin de reevaluar el estado sistemático y taxonómico de *P. furvus*, incorporamos un juego de caracteres genéticos y morfométricos, empleando análisis cladísticos. Aquí presentamos resultados derivados en su mayoría de nuestros análisis genéticos (los resultados del examen filogenético del tamaño y la forma del cráneo aparecerán después). Se utilizaron análisis filogenéticos en cuatro genes mitocondriales (Cytb y ND3-ND4L-ND4) con las respectivas bases de datos por separado o combinadas, seguidos de un análisis con datos genéticos y morfométricos (caracteres de tamaño y forma). La mayoría de las construcciones filogenéticas se realizaron con métodos parsimoniosos, pero también se utilizaron métodos probabilísticos en los análisis con los genes separados por tipo. Se recuperaron topologías similares en todos los análisis del gen Cytb y en todos los análisis de parsimonia con los genes NADH; sin embargo, se obtuvieron topologías en conflicto con los métodos probabilísticos para los genes de la NADH. Además, para entender mejor la variación genética en cada tipo de genes, se condujeron análisis de divergencia genética dentro y entre grupos genéticos y se construyeron redes de haplotipos. Todas las topologías obtenidas utilizando los datos genéticos, cuestionaron el estado monotípico de *Peromyscus furvus*, ya que se identificaron dos clados adicionales que al parecer, corresponden a entidades sin reconocer. La primera de estas, *P. latirostris*, ocurre en la región norteña y podría ser considerada ya sea como una especie o como una subespecie. Una *species nova* de *Peromyscus* que ocurre hacia el sur se considera como especie válida. Además, *P. furvus* s. s. se convierte en una especie politípica al reconocerse al menos dos subespecies (*P. f. angustirostris* y *P. f. furvus*). Los análisis filogenéticos también rechazaron la pertenencia de *P. melanocarpus* y *P. ochraventer* dentro del grupo de especies *furvus*. En cambio, *P. melanocarpus* mostró una mayor afinidad hacia *P. mexicanus totontepecus*, mientras que *P. ochraventer* se unió, ya sea al clado que contenía a *P. melanocarpus* y a *P. m. totontepecus*, o a *Megadontomys cryophilus* en un clado hermano. Finalmente, *Osgoodomys banderanus* (subgénero *Haplomylomys*) siempre permaneció posicionada basalmente y segregada de todos los miembros del subgénero *Peromyscus*.

**Keywords:** Cytochrome-*b* gene; *furvus* species-group; multiple-character-phylogenies; NADH genes; *Peromyscus*.

## Introduction

*Peromyscus furvus* is an endemic Cricetid species from southeastern México that inhabits the temperate cloud and mixed forests from montane highlands and currently is considered as a monotypic species (Rogers and Skoy 2011; but see Ramírez-Pulido *et al.* 2014). Its current distribution ranges from southeastern San Luis Potosí, along the Sierra Madre Oriental, through the Faja Transvolcánica Mexicana and southward to the Sierra Norte de Oaxaca (Rogers and Skoy 2011). The species includes in synonymy (Musser 1964; Hall 1968; Huckaby 1980) *Peromyscus latirostris* (Dalgarno 1950 – type locality Xilitla, San Luis Potosí) and *P. angustirostris* (Hall and Álvarez 1961 – type locality Zacualpan, Veracruz), as well as unassigned specimens from Puerto de la Soledad, Oaxaca.

Although *P. furvus* currently is considered a monotypic species (Rogers and Skoy 2011), several previous studies, under different epistemological and methodological approaches, have noted differences among its synonyms and assigned populations. Such differences include skull size (Musser 1964; Hall 1968; Martínez-Coronel *et al.* 1997; Ávila-Valle *et al.* 2012), allozymes (Harris and Rogers 1999), and mitochondrial genetic sequences (cytochrome-*b* gene, Harris *et al.* 2000, and ND3-ND4 genes, Ávila-Valle *et al.* 2012). In particular, genetic studies have added two distinct scenarios to the status of populations within *P. furvus*. One scenario (Harris *et al.* 2000) based on data from the *Cytb* gene is: a) that the southernmost populations from Puerto de la Soledad, Oaxaca genetically are distinct from others; and b) that the northernmost populations in Xilitla, San Luis Potosí are distinct from *P. furvus* proper. In the geographically intermediate zone, populations from Hidalgo, northern Veracruz and Puebla, historically attributed to *angustirostris*, appear separated from populations from central Veracruz (Xalapa, type locality of *furvus*) but are contained in the same basal clade.

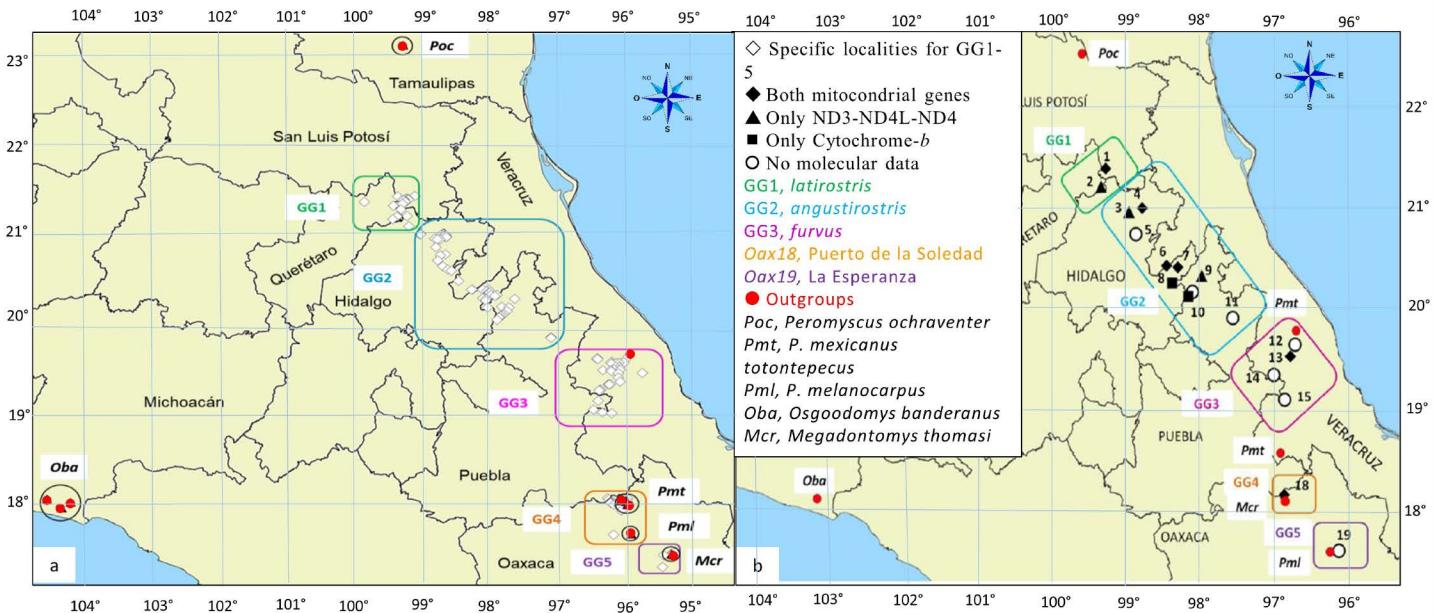
The second scenario, based on data from NAD3-ND4 genes (Ávila-Valle *et al.* 2012), suggests: a) that the most genetically distinctive populations are the northernmost ones from Xilitla, San Luis Potosí and Santa Inés, Querétaro (*latirostris*) and b) that populations from Hidalgo, Puebla, Veracruz, and Oaxaca are grouped together in a central-southern clade. This southern clade appears to contain three subclades: one from Mesa de la Yerba and Xico, Veracruz (*furvus*), a second from El Salto, Puebla (*angustirostris*), and a third containing populations from Puerto de la Soledad, Oaxaca and the remaining populations from Hidalgo, Puebla, and Veracruz, including the type locality of *angustirostris*. These scenarios lead to the following questions: how many different entities or distinctive genetic groups are included within the “monotypic” *P. furvus*? What is the level of genetic variation associated with the recognizable taxonomic units, *e. g.*, species and subspecies? Do the specimens from the northern and southern ends of the distribution deserve specific recognition? What is the systematic and taxonomic status of the intermediate populations in Hidalgo, Puebla, and Veracruz?

Herein, the degree and level of divergence in populations from throughout the geographic range of *P. furvus* was assessed in order to understand if *P. furvus* consisted of one or more evolutionary and taxonomic entities. Our analyses included genetic (mitochondrial genes), morphogeometric (ventral shape of skull), and magnitude characters (centroid size of the latter and linear measurements for skull). Results from the genetic analyses are reported herein, whereas additional morphometric analyses will be presented in a separate manuscript. Additional genetic analyses were conducted to shed light into the complex relationships among populations of the intermediate geographic zone (Harris *et al.* 2000; Ávila-Valle *et al.* 2012).

Finally, Bradley *et al.* (2007) reported some uncertainty in the composition of the *furvus* species group within the genus *Peromyscus*. For example, *P. furvus* historically has been included in different species groups (*e. g.*, *mexicanus* species group; Hooper 1968; Huckaby 1980) until it was assigned to its own group (Carleton 1989). However, the species composition within the *furvus* species group also has varied among authors (Carleton 1989; Wade *et al.* 1999; Musser and Carleton 2005). Herein, results from our analyses contribute to refine the composition of the *furvus* species group, in the light of the latest phylogenies provided for the genus (Bradley *et al.* 2007; Platt *et al.* 2015).

## Materials and methods

**Definition of groups.** Populations from the geographic range of *Peromyscus furvus* were arranged into Operational Taxonomic Units (OTUs) according to their historical or current taxonomic designation and to their geographic provenance (Appendix 1, Figure 1). Hereafter, we refer to the OTUs of the Ingroup (IG) as Genetic Groups (GG1–5). GG1–3 with their taxonomic designations and their States of origin are: GG1 (*latirostris*, *Pf*) from San Luis Potosí and Querétaro; GG2 (*angustirostris*, *Pa*) from Hidalgo, Puebla, and Veracruz; and GG3 (*furvus*, *Pf*) from central Veracruz. The specimens with no formal designation include samples from two localities in Oaxaca, Puerto de la Soledad, referred to as GG4 (*Oax18*), and La Esperanza, GG5 (*Oax19*); however, GG5 samples lacked genetic data and were included only in morphometric analyses. We also selected five OTUs as Outgroups (OGs) based on evidence from recent phylogenies for peromyscines (Bradley *et al.* 2007; Platt *et al.* 2015). The three congeneric species include *P. ochraventer* (*Poc*) and *P. melanocarpus* (*Pml*), which have been associated with the *furvus* species group by Carleton (1989) and Musser and Carleton (2005), respectively, and *P. mexicanus totontepecus* (*Pmt*) from the closely related *mexicanus* group. Also, based on the aforementioned studies (Bradley *et al.* 2007; Platt *et al.* 2015), we included two additional species from different genera; *Megadontomys cryophilus* (*Mcr*), given its relative closeness to *P. furvus*, and *Osgoodomys banderanus* (*Oba*) as the most distant and possibly more conserved species (*i. e.*, tentative root).



**Figure 1.** Geographical delimitation of the Genetic Groups (GG1–5) in the distribution of *Peromyscus furvus* used in this study as Ingroups (IGs), together with five outgroups (see legend); GG5 was included only in morphometric analyses. The map in (a) shows the location of specific localities that were pooled into group localities (numbers 1–19) in further analyses (b). See Appendix for details.

**Molecular characters.** Genetic sequences were downloaded, as complete as possible, from GenBank® (Genetic Sequence Database) for Cytochrome *b* gene (Cyt-*b*; IG  $n = 53$ , OG  $n = 13$ ) and for three genes of the Nicotinamide Adenine Dinucleotide Hydride (NADH: ND3-ND4L-ND4; IG  $n = 57$ , OG  $n = 6$ ). The sole exception was a sequence of NADH genes for *Poc* obtained from [Wade et al. \(1999\)](#). The sequences were aligned beginning at the start codon with the MEGA7 ClustalW option (Molecular Evolutionary Genetics Analysis, Ver. 7.0–7.2, [Kumar et al. 2016](#)) and truncated to similar lengths in all taxa. We analyzed 719 bases from the *Cytb* gene and 971 bases from the NADH genes (ND3, 306; ND4L, 293; ND4, 373; the tRNA-Arg interregion was eliminated, as it was unknown for *Pml*). Information about the geographic origin of sequences, GenBank accession numbers, and references appear in the Appendix.

**Morphometric characters (continuous linear measurements).** In our analyses of overall skull size, we included all OTUs (GG1–5 + five OGs). Herein, 233 specimens from 129 specific localities (SL) represented the IG, and 116 specimens from 11 SL represented the OG. We georeferenced all SLs using QGIS Wien (ver. 2.080–2.93) to project them onto a map (Figure 1A) and to pool the IG samples into 19 group localities (GLs), which in turn were assigned to GG1–5. We also used geographic coordinates of GLs to estimate the geographic distances (km) between every pair of GLs. Morphometric characters for analysis of overall skull size comprised 18 cranial measurements from adult specimens following [Ávila-Valle et al. \(2012\)](#) (see figure 1:167 therein). Additional specimens were added to increase sample sizes in all OTUs ( $n \geq 15$ , except for *Mcr*,  $n = 14$ ). Appendix provides the geographical data, sample sizes, and museum designations.

Analyses of the ventral shape and size of the skull were based on only complete and undamaged skulls (listed in bold in the Appendix). We selected five structures from the ventral view of skull as morphogeometric characters. The details for the anatomical origin of the structures (and substructures contained in them), as well as the quantity and position of the landmarks and semilandmarks outlining each morphogeometric configuration, will be described elsewhere. We measured each skull with type 1 and 2 landmarks *sensu* [Bookstein \(1991\)](#), taken from the literature ([Myers et al. 1996](#); [Cordeiro-Estreia et al. 2008](#); [Grieco and Rizk 2010](#); [Cordero and Epps 2012](#); [Holmes et al. 2016](#)), using utilities in the TPS series (Thin Plate Spline, Ver 2.31, TPS, available at <https://life.bio.sunysb.edu/morph/> ...html; [Rohlf 2015](#)). In the examination of cranial contours, we followed [Sheets et al. \(2004\)](#) to adjust the semilandmarks. In the data matrix for morphogeometric characters, the columns had the consecutive coordinates of the landmarks and semilandmarks for the respective configuration ([Sheets 2002](#)). We also calculated the centroid size (CSs) of each configuration in each specimen as the square root of the sum of the squared distances, using each landmark toward the centroid ([Zelditch et al. 2004](#)). We then computed the average centroid size for the respective configuration, according to OTUs. All geometric morphometrics programs of the IMP8 series (Integrated Morphometric Package, Ver. 8.0) were downloaded from Canisius College (Buffalo, NY, <https://www3.canisius.edu/~sheets/> /morphsoft.html; [Sheets 2002](#)). Prior statistical analyses (descriptive statistics and normality tests), as well as other univariate and multivariate statistics of these data, were conducted with PAST (PAleontological STatistics, Ver. 3.15; [Hammer et al. 2001](#)) at a significance level of  $P \leq 0.05$  (unpublished results).

*Molecular phylogenies based on Cytb and NADH genes.* We constructed single gene phylogenies with all available sequences of the mitochondrial genes, using maximum parsimony (MP), Bayesian inference (BI), and maximum likelihood (ML) methods. In the MP analysis, the evolutionary model used Farris' optimization as the standard option in TNT program (Tree analysis using New Technology, ver. 1.1; [Goloboff et al. 2008](#)) with the algorithm new technologies (NTS) in the search of the most parsimonious trees. In the probabilistic methods, we performed searches for the most suitable evolutionary model by type of mitochondrial genes (Jmodeltest, ver. 2.0, <https://github.com/ddarriba/jmodeltest>; [Guindon and Gascuel 2003](#); [Felsenstein 2005](#); [Posada 2008](#)) under the Bayesian Information Criteria (BIC) and Akaike Information Criteria (AIC). To construct the Bayesian phylogenies, we used four Markov chains and the Metropolis routine of MrBayes (ver. 3.2.6, <https://mrbayes.sourceforge.net>; [Ronquist et al. 2012](#)). We searched for the most likely tree with two runs of 1,000,000 generations for the Cytb gene and twice that for the NADH genes. In each case, temperature was 0.01, 0.25 % of the data was discarded, the most plausible trees were separated in the runs, and the most probable tree was selected. We pooled the two trees from each run based on each gene in a strict consensus cladogram. The MV analyses were performed on the CIPRES web page (Cyber - Infrastructure for Phylogenetic Research, <https://www.phylo.org>), loading the respective molecular data matrices to search for the most probable tree, obtain the branch lengths, and estimate bootstrap support (1,000 replications).

*Phylogeny with genetic and morphometric characters.* In this multicharacter phylogenetic analysis, we used only the same or the closest SLs represented by sequences from both genes and that had data for both the entire cranial size and for the ventral shape and size of skull. The Appendix shows in italics the IG OTUs arranged by GLs, according to GG1–4. The character matrix combined data from molecular together with morphometric characters of the size and shape of the cranium, resulting in 29 IGs, 5 OGs as OTUs, and 1,718 characters (34r x 1,718c).

Regardless of character type, all integrated analyses used MP in the TNT program because it allowed us to examine different types and large number of characters ([Goloboff et al. 2008](#)). In all cases, we used bootstrap resampling with 1,000 repetitions to assess branch support of the resulting topologies. A bootstrap support  $\geq 90$  was considered as strong. In the molecular analyses, 29 IG sequences from 13 SLs (Cyt-b,  $n = 6$ ; ND3-ND4L-ND4,  $n = 7$ ) were selected for each gene, considering either that all genes were represented or that the SLs were  $\leq 10$  km distant from each other. We included five more sequences in each gene to represent the respective OTUs of the OGs. The geographic data of the examined genetic sequences, arranged by GG1–4, appear as italics in the Appendix. We used the sequences, saved in FASTA and ASCII formats by type of mitochondrial genes, to construct the matrix for molecular characters with the

number of bases in the columns and the OTUs (GG1–4) in the rows. The search for the most parsimonious trees was sectorial with 20 changes per sector, Wagner trees (ratchet) with 100 substitutions, drift with 100 substitutions, and merging five trees per replication. If the analysis resulted in more than one tree, a strict consensus cladogram was constructed. Likewise, to determine *a priori* the number of steps in Wagner trees, we used [Farris \(1970\)](#) optimization and an evolutionary model where the position of the four bases (A, T, C, G) was allowed to change in any direction without penalizing reversals. Each change was counted as a single step.

*Analyses of genetic variation and divergence in the IG.* To assist in the interpretation of the topologies in the molecular phylogenetic analyses, as well as to facilitate recognition of the taxonomic level of the GGs, we examined the genetic variation within and among GG1–4 in each mitochondrial gene (Appendix). Following [Ávila-Valle et al. \(2012\)](#), we used the [Tamura and Nei \(1993\)](#) genetic distances for GG1–4 with ARLEQUIN (ver. 3; [Excoffier et al. 2005](#)) to construct the respective genetic data matrices. Genetic distances between pairs of OTUs were estimated, and the probability for statistically significant differences ( $P \leq 0.05$ ) by each type of gene was analyzed using AMOVA ([Excoffier et al. 1992, 2005](#)). These analyses also provided the *Phi* genetic diversity index and the *Theta Phi* statistics, with their respective standard deviations for summarizing the degree of differentiation between the divisions of a population (variance components; [Excoffier et al. 1992](#)) in GG1–4. In addition, these analyses computed base frequency, or number of mutations, according to their type (transitions, ts; transversions, tv; substitutions, sb; indels, in), and the respective genetic distance averages and SD per GG. The AMOVA analyses were repeated each time significant differences were identified within the same GG, in order to test the significance of subgroups.

In order to visualize the relative position of sequences, GLs, and GG1–4, we used an analysis of haplotype networks. This analysis allowed identification of related groups and determine degrees of genetic divergence between GGs. Haplotypes were obtained from each population (Appendix) and DNAsp (Ver. 5.10.01; [Librado and Rozas 2009](#)) was used to build the corresponding network using the software Network (Ver. 5.0.01, <https://www.fluxus-engineering.com>). We labeled each network with a color code to identify allotted haplotypes in different GGs (GG1, green; GG2, blue; GG3 pink; and GG4, orange); when a GG contained more than one GL, we used additional tones of its assigned color. In these analyses, we interpreted the number of mutations between the haplotypes assigned to GLs and GGs as indicative of genetic divergence ([Excoffier et al. 1992](#)). We conducted additional analyses to interpret the respective distance matrices in terms of the criteria outlined by [Bradley and Baker \(2001:963\)](#) with 2-parameter model of nucleotide substitution ([Kimura 1980](#)) assuming minimum evolution in the Cytb gene. First, we estimated

the genetic distances under the [Tamura and Nei \(1993\)](#) evolutionary model with a gamma distribution using the Distance menu and the Compute Between Group Mean Distance option in MEGA7 (ver. 7.0; [Kumar \*et al.\* 2016](#)) and then transformed them into percentages. We first performed this analysis with the IG and their respective GLs used in the integrated phylogenies and then we averaged the data of the GLs for the respective GG and included the five OGs. Second, in order to determine phylogenetic and taxonomic distinctions between GG1–4, we used the criteria of [Baker and Bradley \(2006\)](#) for applying genetic distances to the genetic species concept.

## Results

*Phylogenies based on mitochondrial genes.* The most suitable evolutionary model for the *Cytb* data set was GTR I+G (BIC,  $-\ln L = 2729.48$ ; AIC =  $-\ln L 2725.67$ ). The parameters for this evolutionary model used a gamma distribution and included base frequencies of A = 0.3302, C = 0.2874, G = 0.1347, T = 0.2478. A fixed shape = 1.2190 indicated a low rate of variation with nucleotides evolving slowly; 0 invariable sites; fixed rate of transversions and transitions: AC = 5.1938; AG = 18.6338; AT = 8.3552; CG = 1.2041; CT = 92.4518; and GT = 1.0. Number of substitution types (6) indicated that all transitions and transversions were treated differently.

For the NADH dataset, the best evolutionary model was TPM 2 uf + G (BIC,  $-\ln L = 5860.0478$ ; AIC,  $-\ln L = 5860.0478$ ). This evolutionary model was used with a gamma distribution and included base frequencies of A = 0.3492, C = 0.2685, G = 0.0788, and T = 0.3035. Fixed shape = 0.3350, indicating there were different rates of change, and 0 invariable sites. Fixed rate of transversions and transitions: AC = 0.6516; AG = 5.3993; AT = 0.6516; CG = 1.0000; CT = 5.3993; and GT = 1.0, and the number of substitution types was 6.

The MP phylogeny for *Cytb* yielded one tree ( $L = 500$ ,  $IC = 0.704$ ,  $IR = 0.876$ ). The MP analysis of the NADH genes ( $L = 963$ ,  $C = 0.565$ ,  $IR = 0.876746$ ) yielded 41 trees; a strict consensus tree was used to represent the single topology. These two MP topologies (Figure 2), together with both ML and BI for *Cyt-b*, provided the same topology as the integrated phylogenies based on molecular characters (Figure 4). Most differences were due to the location of the OGs in the two probabilistic topologies for the *Cytb* gene (Figure 2a). In the ML tree, *P. ochraventer* (*Poc*), was basal to a strongly supported

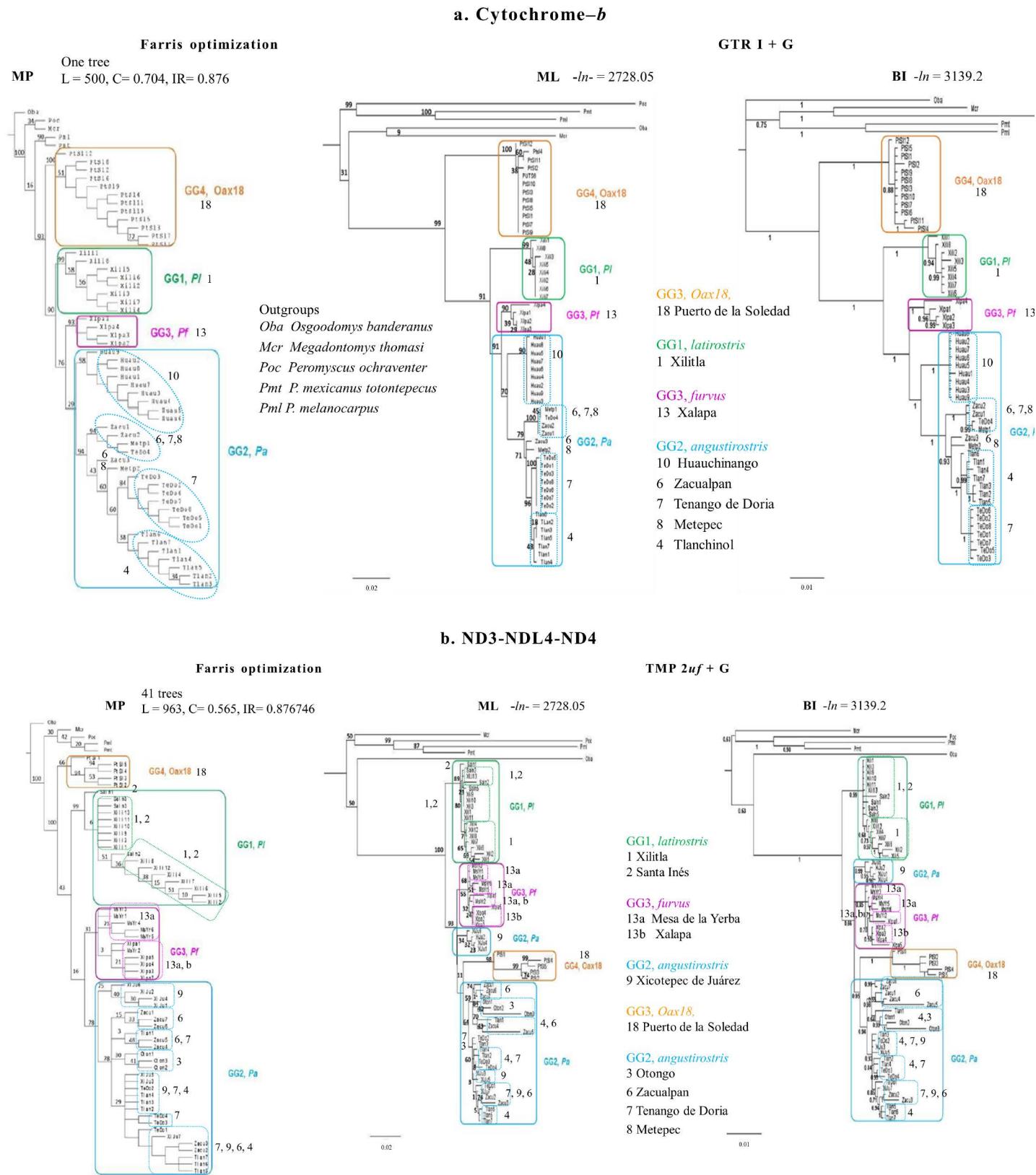
clade containing *P. melanocarpus* (*Pml*) and *P. mexicanus totontepecus* (*Pmt*). *O. banderanus* (*Oba*) and *M. cryophilus* (*Mcr*) were the sister group of the IG in a poorly supported clade (9%). In the Bayesian analysis, *Oba* formed a separate clade basal (1.0) to all the other OGs (0.75). *Mcr* and *Poc* were sister (1.0) to the other *Peromyscus* species (1.0), and all IG members formed a separate clade (1.0).

The two probabilistic topologies for the NADH genes (Figure 2b) rendered a strikingly different topology in both the location for the OGs and for the IG. *Mcr* and the three OG *Peromyscus* were located in a basal clade (50%) with internal branches well supported (87 to 99%). In both ML and BI topologies, *Oba* was the sister group to the IG in a moderately supported clade (ML 50%, BI 0.63). For the IG, GG1 (*latirostris Pl*, 89%, 0.99) was separated from the other GGs (100%, 1.0). The largest clade (93%, 0.66) was comprised of a polytomy containing three subclades. The first of these subclades possessed all samples of GG3 (*furus*, *Pf* 55%, 0.85). The second clade contained individuals with half the sequences from Xicotepic de Juárez, Puebla (GL9) of GG2 (*angustirostris Pa*), located between GG3 (*Pl*) and GG4 (Oax) in the ML (34%). The third clade contained GG4 (Oax18, 98%, 1.0) and the GLs from Hidalgo and northern Veracruz (69%, 0.94) of GG2 (*Pa*). Within the third clade, three sequences from Zacualpan (GL6 74%, 0.99) and all sequences from Otongo (GL3, 84%, 0.64) did not group with other subpopulations (11%, 0.95).

*Interpopulation genetic variation in the IG: genetic distances and tests.* The respective AMOVAs for *Cytb* gene and NADH genes revealed significant differences both among the GGs and within their group localities (GLs, Table 1) as reflected in the single and combined genetic topologies (Figures 2, 4). In the *Cytb* gene, all the genetic fixation indices (FI) scored >70%, whereas in the three NADH genes, only the variation within populations (FST) was greater. FST values were higher in NADH genes than in *Cytb*, indicating a higher level of genetic differentiation within the GLs of the same GG. Similarly, in all genes, the second highest index (FSC) indicated divergence among GGs, although it was lower for the NADH genes. Low FSC values were detected for NADH among populations among GLs within a GG. The variance component that explained most percentage of variance was variation among GG1–4 (FCT) in both mitochondrial genes, followed by variation among the GLs of a GG (FSC), and then variation among the sequences in the same GL (FST).

**Table 1.** AMOVA results for mitochondrial genes showing degrees of freedom (df), sum of squares (SS), variance components (VC), percentage of variance explained (% V), and fixation indexes (FI) indicating average variation: among GG1–4 (FCT); among all populations (GLs) among GGs (FSC); and within a GLs of the same GG (FST). All *p*-values were significant (\*).

Variation source	Cytochrome b					ND3-ND4L-ND3				
	df	SS	VC	% V	FI*	df	SS	VC	% V	FI*
Among GGs (FCT)	3	603.7	16.8	78.3	0.78	3	803.4	17.7	63.24	0.63
Among populations within GGs (FSC)	7	130.0	3.6	16.9	0.77	7	158.1	3.08	10.63	0.28
Within populations in GG (FST)	42	44.1	1.0	4.9	0.95	50	366.4	7.3	26.13	0.73
Total	52	777.7	21.5			60	1328.0	28.0		



**Figure 2.** Topologies resulting from phylogenetic analyses using Maximum Parsimony (MP), Maximum Likelihood (ML), and Bayesian Inference (BI) with sequences for the Cytochrome-b gene with 719 bases (a) and sequences for the NADH genes with 971 bases (b). The numbers on the nodes indicate percentage of support for the branches per 1000 bootstrap replicas. See text for description of support values. See Appendix for details.

Overall, most paired comparisons between GLs among GG1–4 were statistically significant for the *Cytb* and NADH genes (Table 2). GGs at the ends of the distribution range (GG1 *Pl*; GG4 *Oax18*) diverged from the intermediate GGs

(GG, *Pa*; GG3 *Pf*) and from each other with greater and significant genetic distances in both genes (Table 2). For NADH genes, the northernmost GG1 (*Pf*), included all the sequences from San Luis Potosí (GL1 Xilitla) and was not

**Table 2.** Genetic divergence in mitochondrial genes Cytochrome-*b* (A) and ND3-ND4L-ND4 (B), depicted by distances of Tamura and Nei's (N-T, below diagonal) between pairs of Group Localities (GL#) in the Genetic Groups (G1–4), arranged in a NW-SE direction. Geographic distances (e. g., straight lines for km between GLs) are also shown as reference (above diagonal). Number of asterisks depict significant N-T genetic distances among and within GLs (and GGs), according to level of *p*-values: \* *p* < 0.05 and \*\* *p* < 0.01; bold distances had non-significant *p*-values (> 0.05). Colors correspond to color-code in Figures.

A. Cytochrome- <i>b</i>												
GL#	GG	Code	Xili	Tlan	Zacu	TeDo	Metp	Huau	Xalp	PtSl		
1	GG1	Xili	0	65.17	137.18	147.53	158.52	182.87	322.11	447.53		
4	GG2	Tlan	0.96*	0	124.61	136.07	95.83	166.99	256.61	380.72		
6	GG2	Zacu	0.92*	0.73*	0	14.65	<b>24.41</b>	45.24	190.24	306.96		
7	GG2	TeDo	0.93*	0.53**	0.55*	0	19.93	34.38	177.45	298.48		
8	GG2	Metp	0.91**	0.69*	<b>0.28</b>	0.48**	0	27.99	<b>173.64</b>	292.28		
10	GG2	Huau	0.97*	0.97**	0.93**	0.92**	0.92*	0	146.03	271.31		
13	GG3	Xlpa	0.89*	0.86**	0.73*	0.82**	<b>0.65</b>	0.90**	0	161.77		
18	GG4	PtSl	0.97*	0.98**	0.96**	0.96*	0.96*	0.98**	0.96**	0		

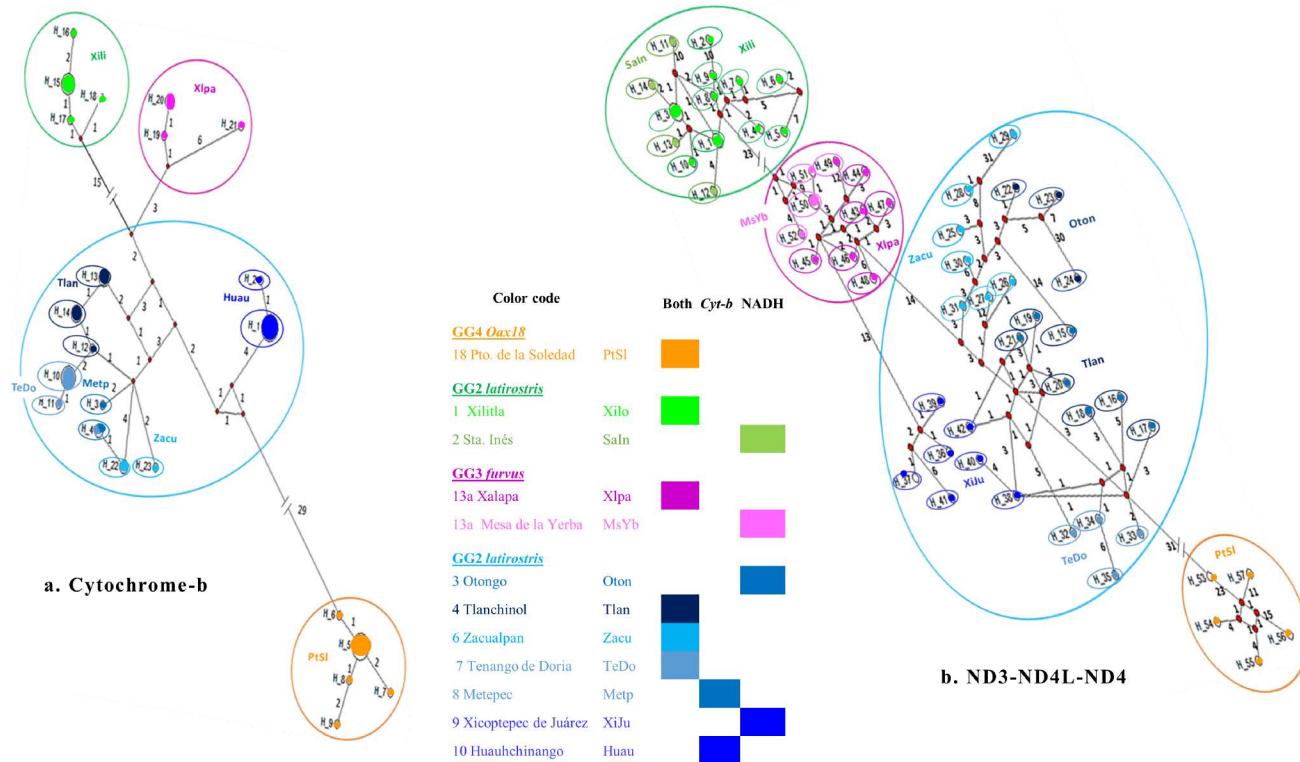
B. ND3-ND4L-ND4												
GL#	GG	Code	Xili	Saln	Oton	Tlan	Zacu	TeDo	XiJu	MsYb	Xlpa	PtSl
1	GG1	Xili	0	<b>21.86</b>	58.62	65.17	137.18	147.53	175.43	342.11	322.11	447.53
2	GG1	Saln	0.1	0	46.76	58.29	124.61	136.07	165.7	332.87	312.87	429.32
3	GG2	Oton	0.81**	0.71*	0	17.51	78.83	82.88	118.76	286.55	266.55	386.75
4	GG2	Tlan	0.80**	0.75**	0.36*	0	137.18	<b>147.53</b>	117.07	276.61	256.61	380.72
6	GG2	Zacu	0.75**	0.66**	0.20**	0.16*	0	14.65	46.46	210.24	190.24	306.96
7	GG2	TeDo	0.83**	0.80*	0.39*	<b>0.04</b>	0.22*	0	<b>32.10</b>	197.45	177.45	298.48
9	GG2	XiJu	0.75**	0.66**	0.43**	0.28*	0.3**	<b>0.29</b>	0	167.10	147.10	276.65
13	GG3	MsYb	0.84**	0.83**	0.68**	0.65**	0.53**	0.76**	0.44**	0	20.00	161.67
13	GG3	Xlpa	0.79**	0.74**	0.64*	0.65**	0.56**	0.71**	0.46**	0.26**	0	181.67
18	GG4	PtSl	0.84**	0.76**	0.72**	0.78**	0.72**	0.78*	0.73**	0.83**	0.77**	0

genetically different from samples from Querétaro (GL2 Santa Inés). Also, all sequences in GG4 (*Oax18*) from Puerto de la Soledad, Oaxaca (GL18) were genetically segregated from other GLs based on greater geographic distance.

In addition to the lack of genetic divergence between Xilitla (GL1) and Santa Inés (GL2) in GG1 (*P*) in the NADH genes, there were two additional exceptions (Table 2) for significant divergence between the GLs from the intermediate GGs (GG2 *Pa*; GG3 *Pf*). In the *Cytb* gene, Metepec, Hidalgo (GL8, GG2, *Pa*) showed the least genetic divergence from the type localities of either GG2 (GL6, Zacualpan, Veracruz) or GG3 (*Pf*, GL13, Xalapa, Veracruz), regardless of the geographic distance (24.4 and 174 km, respectively). Likewise, for the NADH genes, the lack of genetic divergence involved only GLs from GG2 (*Pa*), including Tenango de Doria (GL7), located almost 150 km from Tlanchinol (GL4) in Hidalgo, and 32 km distant from Xicotépec de Juárez (GL9) in Puebla (Table 2). All remaining GLs at each gene possessed significant distances of genetic divergence (Table 2), regardless of their geographic proximity (e. g., 20 km between Mesa de la Yerba and Xalapa, in Veracruz, NADH). These results suggest that there may be different genetic subgroups within the intermediate portion of the geographic range.

The genetic structure of the GLs in GG1–4 (Table 3) with the *Cytb* gene indicated that the two populations at the end of the geographic range (GL Xilitla, GG1 *Pl*; GL18 Puerto de la Soledad, GG4 *Oax18*) had the lowest genetic divergence. The northern populations of GG2 (*Pa*), Huauachinango in northern Puebla (GL10) and Tlanchinol in northern Hidalgo (GL4) were the most genetically conserved. Tenango de Doria, a southern location in Hidalgo (GL7) and to the southeast of the type locality of GG2 (*Pa*), Zacualpan, Veracruz (GL6), also was genetically conserved with a divergence slightly higher than GL1 and GL18. Zacualpan (GL6), which is geographically closer to populations at Tenango de Doria (GL7) and to the populations of Metepec (GL8), showed twice the amount of genetic variation than the former but 50 % less than the latter. Finally, Xalapa (GL13), in central Veracruz and the type locality for GG3 (*Pf*), showed four times more genetic divergence than Metepec (GL8) and ranked as the most variable GL for the *Cytb* gene. However, the number of indels in the Xalapan sequences (Table 3) were responsible for the higher genetic diversity. Excluding these indels, Xalapa had genetic diversity more similar to Tenango de Doria (GL7, GG2 *Pa*) for the *Cytb* data.

The three NADH genes showed a relative higher amount of genetic diversity among GG1–4 than did the *Cytb* gene



**Figure 3.** Haplotype networks for the Genetic Groups within the Ingroup in the Cytochrome-*b* gene (a) and in the NADH genes (b). Red diamonds depict unknown haplotypes; digits in the joining lines among haplotypes, indicate the number of mutations (genetic divergence); the underlined haplotype H4 is shared in two nearby group localities in Hidalgo. The tones of the colors aim to facilitate location of the group localities within the same GG. See text and Table 4.

(Table 3). The GLs with the lowest genetic divergence were Mesa de la Yerba (GL13b, GG3 *Pf*) in central Veracruz and the most northern and distant Xilitla (GL1, GG1 *Pl*). Tenango de Doria (GL7 *Pa*) and Santa Inés (GL1 *Pl*) were the next least divergent localities. Xalapa (GL13 *Pf*) and Tlanchinol (GL4 *Pa*) possessed intermediate levels of genetic diversity with Tlanchinol (GL4 *Pa*) being similar to the northernmost populations in Puebla, Xicotepec de Juárez (GL9 *Pa*). Finally, although Puerto de la Soledad in northern Oaxaca was located geographically near Zacualpan (GL6 *Pa*) and Tenango de Doria (GL7 *Pa*), it was genetically divergent based on the NADH genes.

**Haplotype networks in the Internal Group.** As in all phylogenetic topologies with molecular data (Figures 2, 4), each GG1–4 is readily recognizable in the haplotype networks (Figure 3) because all haplotypes from the GLs joined, regardless of geographic proximity between their respective GLs (Figure 1b, Table 2). Both haplotype networks reflected the overall geographical distribution of the G1–4 (Figures 1, 3), with sequences of GG1 (*latirostris*, *Pl*) and GG4 (*Oax18*) at the geographical extremes, and GG2 (*angustirostris*, *Pa*) and GG3 (*furus*, *Pf*) join in the intermediate zone. However, in both haplotype networks, GG3 (*Pf*) appears to be closer to GG1 (*Pl*) than GG2 (*Pa*). The distinctiveness of GG1–4 is supported by the large amount of the variance as seen in the AMOVA (78.30 % in *Cytb* and 63.24 % in the NADH genes, Table 1). The haplotype networks are consistent in depicting geographic representation (Table 4, Figure 1, Appendix), but differ in the intrinsic amount of

genetic variation when *Cytb* is compared to NADH (Tables 1, 2). Accordingly, each network displayed its own number of mutations (steps in each branch), including those between known haplotypes, between haplotypes and unknown haplotypes, and between unknown haplotypes. Unknown haplotypes appear as "UnkH" in Table 4 and as red diamonds on Figure 3.

The 53 sequences in the *Cytb* gene coalesced into 24 distinctive haplotypes inside the eight GLs representing GG1–4 with one exception (Table 4, Figure 3a). GG2 (*Pa*) from Hidalgo, Tenango de Doria (GL7), and Metepec (GL8), shared the same haplotype (H4). For *Cytb*, all GLs had 1–2 shared haplotypes except for Metepec (GL8). The number of mutations ranged from 1 to 29 with most found within the GGs, except for each of the type localities of GG3 (*Pf*, GL13, H21, 6) and GG3 (*Pa*, GL6, H22, 4). Among GGs represented by a single GL (Figure 3a), populations from Oaxaca (GG4 *Oax18*) had no unknown haplotypes, whereas San Luis Potosí (GG1 *Pl*) and (GG3 *Pf*) had one each. GG2 (*Pa*) had five GLs and included seven unknown haplotypes (Figure 3a), most of which joined the GLs from Hidalgo (GL4, GL7, and GL8) with Zacualpan, Veracruz (GL6), the type locality of *angustirostris*. GL4 (Tlanchinol) and GL7 (Tenango de Doria), located 82.93 km apart (Table 2), joined through two mutations; whereas an unknown shared haplotype joined four geographically closer GLs (Table 2) through either 1, 2, or 4 mutations. Zacualpan (GL6) joined through to Metepec (GL8, 21.41 Km) or Tenango de Doria (14.65 km) by a single mutation (H4). Tenango de Doria (GL7) and Metepec

**Table 3.** Indexes of genetic diversity ( $Pi$ ) in the representative populations of the genetic groups (GG1–4) of the Ingroup, indicating the related statistics ( $\Theta Pi$  with one standard deviation, SD), the number (No.) of mutations by type, and the respective mean with its standard deviation (SD) for cytochrome-*b* and ND3-ND4L-ND4. Colors correspond to those in the Figures. See Appendix for details.

Cytochrome- <i>b</i>										
Group Locality#	GL1	GL4	GL6	GL7	GLG8	GL10	GL13	GL18	Mean	SD
Genetic Group GL Code	GG1 Xili	GG2 Tlan	GG2 Zacu	GG2 TeD	GG2 Metepec	GG2 Huau	GG3 Xipa	GG4 PtSl		
<i>n</i> of sequences	8	7	3	80	2	9	4	12		
Statistic	Genetic diversity									
$Pi$	1.43	0.86	5.33	3	10	0.22	61.5	1.44	10.47	19.5
$\Theta Pi$	1.43	0.86	5.33	3	10	0.22	61.5	1.44	10.47	19.5
SD $\Theta Pi$	1.1	0.78	4.4	1.99	10.49	0.33	40.56	1.06	7.59	12.8
Mutations, number of										
transitions, ts	5	2	7	11	10	0	8	6	6.13	3.5
transversions, tv	0	0	1	1	0	1	1	1	0.63	0.5
substitutions, sb	5	2	8	12	10	1	9	7	6.75	3.6
indels, in	0	0	0	0	0	0	114	0	14.25	37.7
Sum	10	4	16	24	20	2	132	14		
No. of ts sites	5	2	7	11	10	0	8	6	6.13	3.52
No. of tv sites	5	2	7	11	10	0	8	6	0.63	0.48
No. of ss sites	5	2	8	12	10	1	9	7	6.75	3.6
No. of in sites	0	0	0	0	0	0	114	0	14.25	37.7
Sum	15	6	22	34	30	1	139	19		
ND3-ND4L-ND4										
Group Locality#	GL1	GL2	GL3	GL4	GL6	GL7	GL9	GL13	GL13	GL18
Genetic Group Code	GG1 Xili	GG1 Saln	GG2 Oton	GG2 Tlan	GG2 Zacu	GG2 TeDo	GG2 XiJu	GG3 MsY	GG3 Xipa	GG4 PtSl
<i>n</i> of sequences	13	5	3	7	7	4	7	6	5	12
Statistic	Genetic diversity								Mean	SD
$Pi$	7.9	9.83	28.7	13.05	23.29	9	18.19	6.33	12	22.8
$\Theta Pi$	7.9	9.83	28.7	13.05	23.29	9	18.19	6.33	12	22.8
DE $\Theta Pi$	4.42	6.83	21.8	7.68	13.4	6.28	10.55	4.04	7.67	14.21
Mutations, number of:										
transitions, ts	9	11	30	27	39	11	28	10	25	34
transversions, tv	18	8	13	11	27	6	8	5	4	19
substitutions, sb	27	19	43	38	66	17	36	15	29	53
indels, in	1	0	0	0	0	0	0	0	0	0.1
Sum	30	38	86	76	132	34	70	30	58	106
No. of ts sites	9	11	30	27	39	11	28	10	25	34
No. of tv sites	9	11	30	27	39	11	28	10	25	34
No. of ss sites	27	19	43	38	64	17	36	15	29	53
No. of in sites	1	0	0	0	0	0	0	0	0	0.1
Sum	46	41	103	92	142	39	92	35	79	121

(GL8), which are separated by 19.93 km, also differed by one mutation. The most divergent subpopulation in GG2 (*Pa*) was Huachinango, Puebla (GL10) which is 28 km from Metepec (GL8) but that was separated from it (and from all the other GLs) by 5 to 6 unknown haplotypes and at least 11 mutations.

Although Huachinango (GL10) is the most proximate (271.31 km) subpopulation of GG2 (*Pa*) to Puerto de la

Soledad, Oaxaca (GG4 *Oax18*), the number of mutations between these two GGs scored highest in the *Cytb* network, with 29 mutations and 1 to 2 unknown haplotypes (Figure 3a). Branches joining GG2 (*Pa*) to GG3 (*Pf*) included 3 to 7 unknown haplotypes. In the network analysis (Figure 3a), Xalapa (GL13, *Pf*) was genetically divergent from Tlanchinol (GL4, 256.6 km) and to the group formed by Zacualpan (GL6, 190.24 km), Tenango de Doria (GL7, 177.45 km),

**Table 4.** Haplotypes (H#) analyzed in gene networks for Cytochrome-*b* and three NADH genes, according to OTU (genetic group, GG1–4, taxonomic entity and key). Geographic data include the group localities (GL#) in a NW-SE direction (see Figure 1b); the asterisk (\*) indicates type localities or first known localities. The total number of genetic sequences (#Sequences) in each OTU appears in the next column with the GL code, whereas in the last column, the number inside parenthesis indicates how many were shared in a common haplotype. See Appendix 1. Colors for OTUs are as in Figure 3.

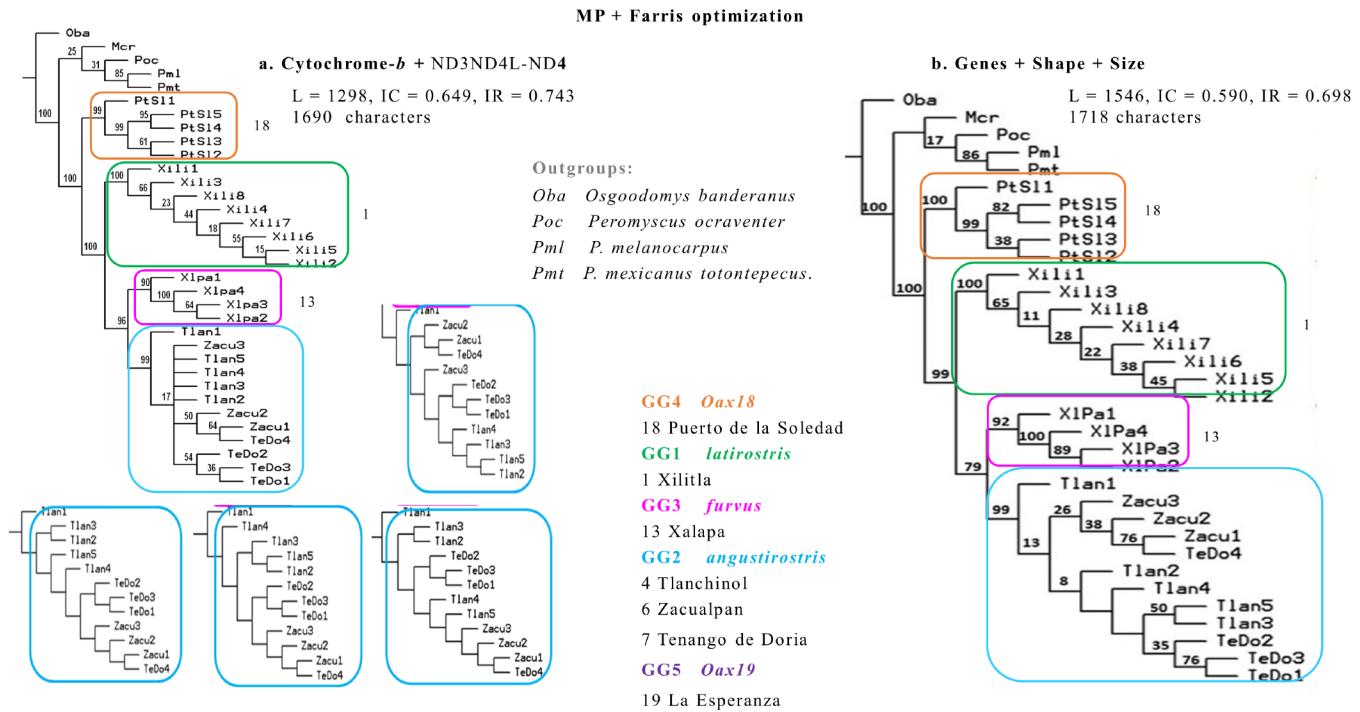
OTU	GL#. Name, State	GL Code	Haplotypes	Common H#s
		# Sequences	H#	(#Sequences)
<b>Cytochrome-<i>b</i> gene</b>				
<b>GG1 <i>latirostris Pa</i></b>	1. Xilitla, San Luis Potosí*	Xili 1–8	H15–H18	H15(5)
<b>GG2 <i>angustirostris Pa</i></b>	4. Tlanchinol, Hidalgo	Tlan 1–7	H12–H14	H13 (3), H14(2)
	6. Zacualpan, Veracruz*	Zacu 1–3	H22, H23	H22(2)
	7. Tenango de Doria, Hidalgo	TeDo 1–9	H4, H10–H11	H10(7), H4(Metpc)
	8. Metepec, Hidalgo	Metp 1–2	H3, H4	H4(TeDo)
	10. Huauchinango, Puebla	Huau 1–9	H1–H2	H1(8)
	13. Xalapa, Veracruz*	Xlpa 1–4	H19–H21, H20	H20(2)
<b>GG3 <i>furvus Pf</i></b>	18. Puerto de la Soledad, Oaxaca*	PtSl 1–12	H5–H9	H5(8)
<b>ND3-ND4L-ND4 genes</b>				
<b>GG1 <i>latirostris Pa</i></b>	1. Xilitla San Luis Potosí*	Xili 1–13	H1–H10	H3(3), H1(2)
	2. Santa Inés, Qro (Saln1–4)	Saln 1–4	H11–H14	none
<b>GG2 <i>angustirostris Pa</i></b>	3. Otongo, Hgo (Oton1–4)	Oton 1–4	H22–H24	none
	4. Tlanchinol, Hidalgo	Tlan 1–4	H15–H21	none
	6. Zacualpan, Veracruz*	Zacu 1–7	H25–H31	none
	7. Tenango de Doria, Hidalgo	TeDo 1–4	H32–H35	none
	9. Xicotepec de Juárez, Pue (XiJu1–7)	XiJu 1–7	H36–H42	none
	13a. Xalapa, Veracruz*	MsYb 1–6	H43–H48	H50(2)
<b>GG3 <i>furvus Pf</i></b>	13b. Mesa de la Yerba, Veracruz*	Xlpa 1–6	H49–H52	none
	18. Puerto de la Soledad, Oaxaca*	PtSl 1–5	H53–H57	none

and Metepec (GL8, 173.64) by 10 mutations. Xalapa (GL13 *Pf*) compared to that of Huauchinango (GL10), to which it is geographically closer (146.04 km), differed by 11 mutations. Finally, the branches between GG1 (*Pi*, Xilitla, San Luis Potosí, GL1) and GG3 (*Pa*, Xalapa, Veracruz, GL13) showed a genetic divergence of 19 mutations (Figure 3a).

In the NADH genes network (Figure 3b), the 58 sequences resulted in 57 distinctive haplotypes, the sole exception being H50 from Mesa de la Yerba in central Veracruz (GL13, GG3 *Pf*), which was presented by two sequences. With the exception of Puerto de la Soledad, Oaxaca (GL18, GG4 *Oax18*), all other GGs had more than one GLs representing them: two in either GG1 (*Pi*) and GG3 (*Pf*) and five in GG2 (*Pa*). Overall, the number of mutations in the network branches ranged from 1 to 31. In the NADH network, the total number of unknown haplotypes was slightly more than triple that found in *Cytb* (Figure 3b). Although most haplotypes from the same GL were spatially closer, there was more intermingling among haplotypes from different GLs, thus allocating haplotypes among some GLs. Furthermore, the higher and more fluctuating number of mutations among the GLs, the greater intermingling of haplotypes from different GLs in the same genetic group.

The lower number of shared haplotypes rendered a lower percentage of variance explained by the variation among GG1–4 in the AMOVA (Table 1). For the NADH genes, variation within subpopulations (GLs) accounted for a higher amount of variance than in the *Cytb* gene.

In GG4 (*Oax18*, GL1), no sequence joined another except through unknown haplotypes (number of mutations was 4, 11, 15, and 23; Figure 3b). H53, the most divergent haplotype, diverged by 31 mutations from an unknown haplotype that join a GL of GG2 (*Pa*) from Puebla (GL9, Xicotepec de Juárez, 276 km) and two from Hidalgo (GL7, Tenango de Doria, 298 km; GL4 Tlanchinol, 380 km). The haplotypes of three GLs formed their own subpopulation and shared haplotypes from others through unknown haplotypes. These three GLs contained 1 to 6 mutations and only Tenango de Doria (GL7; H34, H35) and Xicotepec de Juárez (GL9; H38, H20) had haplotypes that joined to the same subpopulation. Xicotepec de Juárez, Puebla (GL9) is 32 km from Tenango de Doria (GL7) and 117 km from Tlanchinol (GL4) and their haplotypes diverge from unknown haplotypes by 1, 2, or 6 mutations. Only H15 from Tlanchinol (GL4) joins Otongo (GL3), 17 km distant, through a genetic divergence of at least 17 unknown haplotypes. The level of divergence



**Figure 4.** Topologies from selected populations of *Peromyscus furvus*, integrating two types of mitochondrial genes in a strict consensus cladogram (a; note the four outcomes for GG2), and the former with 28 morphometric characters for skull shape and size in a completely resolved single phylogenetic tree (b). The numbers on the nodes indicate percentage of support for the branches per 1000 bootstrap replicas. See text for description of support values.

in the sequences from Otongo (GL3) includes 1, 7, and 30 mutations from unknown haplotypes. The sequences from the type locality Zacualpan, Veracruz (GL6) also formed a separate group but joined by 7 or 11 mutations to Otongo (GL3), 78 km away. Most genetic divergence among subpopulations contained 1 or 3 mutations, however, haplotypes had 6, 8, 12, and 31 mutations to an unknown haplotype. Therefore, the overall number of mutations in GG2 (*Pa*) was high. Consistent with its intermingling pattern, GG2 (*Pa*) joins to GG3 (*Pf*) through clusters of two different subpopulations separated by 46 km (Zacualpan GL6, Xicotepec de Juárez GL9). The route from Zacualpan (GL6 *Pa*) involved 14 to 17 mutations between haplotypes connecting it to Xalapa (GL13 *Pf*). The alternative route from Xicotepec de Juárez (GL9 *Pa*) to Xalapa (GL13 *Pa*) encompasses 13 mutations between unknown haplotypes (GL9 is 147 km from GL13).

The two GLs in GG3 (*Pf*) are 20 km distant and joined by either 5 or 15 mutations (Table 2). In Xalapa (GL13a), the number of mutations from an unknown haplotype ranged from 1 to 3, and 6, but in Mesa de la Yerba (GL13b) they were higher with 1, 4, 9, or 12 mutations. The only joining of two haplotypes from the same subpopulation (Figure 3b) occurred in Mesa de la Yerba, which had a single mutation between haplotype H50 and H51. H50 was the only shared haplotype in the entire NDH network. Between GG3 (*Pf*) and GG1 (*Pf*), there are 23 mutations segregating the two unknown haplotypes to which 5 or 10 mutations join Mesa de la Yerba (GL13a) and another one joined Xilitla, San

Luis Potosí (GL1). These two GLs are 322 km distant. In Xilitla (GL1), the number of mutations from a haplotype to an unknown haplotype was 1, but there also were 2, 5, 7, and 10 mutations. The number of mutations between GL1 and Santa Inés, Querétaro (GL2), was 2 or 4, and the latter joined to unknown haplotypes by 1, 2, or 10 mutations.

**Combined datasets.** In the MP analyses for combined datasets (Figure 4), all OGs were distinct from the members of the IG and each GG1–4 was readily recognizable with support values of > 90 to 100 %. *Oba* was always at the root, whereas the sister group to all IG was (*Mcr*(*Poc*(*Pml* *Pmt*))). In this sister group, only the clade of *Pml* and *Pmt* was well supported (85%, 86%). In the IG, the sequences from Puerto de la Soledad, Oaxaca (GG4, *Oax18*) were located basally and separated from other GGs. In the most derived clade, sequences from Xilitla, San Luis Potosí (GG1 *Pf*) showed full support and separated from a sister subclade with sequences from the two remaining GGs. Clade support was high in ML but lowered in the IB topology for the sequences from Xalapa, Veracruz (GG3 *Pf*) and in GG2 (*Pa*) with all the remaining sequences from Hidalgo and Veracruz (GG2 *Pa*). However, the clades of these two GGs were distinctive from each other and with strong support.

The MP strict consensus cladogram combining all molecular sequences (Figure 4a) resulted from four most parsimonious trees ( $L = 1298$ ,  $IC = 0.649$ ,  $IR = 0.743$ ). The differences among these original trees (Figure 4a) were due to the relative position of the specimens of GG2 (*Pa*) from the two group localities in Hidalgo, Tlanchinol (GL4) and

Tenango de Doria (GL7) with respect to the specimens from Zacualpan, Veracruz (GL6). The addition of morphometric data in the multicharacter phylogeny (Fig 4b) resolved the aforementioned discrepancies in a single, though longer tree ( $L = 1546$ ,  $IC = 0.590$ ,  $IR = 0.698$ ). In agreement to its geographic location, a sequence from Tlanchinol (GL4) preceded a group with all the sequences from Zacualpan (GL6) and one sequence from Tenango de Doria (GL7). This sister clade included sequences from GL4 and GL7 that are geographically more distant (Table 2) and poorly supported (8%). However, some clades ranked from moderate (Tlanchinol, 50%) to acceptable (Zacualpan and Tenago de Doria, or the latter alone, with 76% each). In both combined datasets with genetic data (Figure 4), branches defining subclades associated with the separation of every GG were fully (100%) or very highly supported (90 to 99%).

*Percentage genetic distances within the IG and with all OTUs for the Cytb gene.* In the analysis of the IG (Table 5a), the larger genetic distances occurred between GG4 (Oax18) from Puerto de la Soledad, Oaxaca (GL18) and the other GGs (6.6 to 7.8%), with the exception of Metepec, Hidalgo (GL8) of GG2 (*angustirostris*) which was <2%. Likewise, although of smaller magnitude, the genetic distances associated with GG1 (*latirostris*) from Xilitla, San Luis Potosí (GL1) showed segregation from all the other GGs (3.8 to 7.8%). GG3 (*furvus*) from Xalapa, Veracruz (GL13) differentiated from GG1 (*latirostris*, 3.8%) and especially from GG4 (Oax18, 7.6%). However, the genetic distances of GG3 (*furvus*) were lower (2.4

to 3.0%) with respect to the remaining localities in Hidalgo, Puebla, and Veracruz (corresponding to GG2, *angustirostris*). The variation among the localities within GG2 (*angustirostris*) ranged 0.7 to 2.7%. Noticeably, these analyses also showed minimal genetic distances even among individuals from the same locality (Table 5a) both in Metepec, Hidalgo (GG2 *angustirostris*) and in Xalapa, Veracruz (GG3 *furvus*).

In the genetic distances including all OTUs (Table 5b), all comparisons among the OGs and IGs, averaged >11% with an average genetic divergence among the ten OTUs of 16.2% (range values, 11.8 to 22.9%); among the OGs from the IG, the values were 15.6% (11.8 to 19.8%). *Peromyscus ochraventer* (Poc, 18.4%) had the greatest divergence from the IG, followed by *O. banderanus* (Oba, 16.4%) and *P. melanocarpus* (Pml, 16.1%). *P. mexicanus totontepecus* (Pmt, 14.2%) and *M. cryophilus* (Mcr, 13.0%) showed the lower genetic distances from the IG. The overall average genetic divergence of the IG from all the OGs was 15.6%. GG3 (Pf, 16%) was the most divergent to all OGs, followed by both GG2 (Pa, 15.7%) and GG4 (Oax18, 15.7%), whereas GG1 (Pl, 15.1%) was slightly less divergent. When pooled together, GG3 (Pl) and GG2 (Pa) were slightly more divergent from the OGs (15.9%) than GG4 (Oax18). When the IG was individually compared with all three *Peromyscus* in the OG (Poc, Pml, and Pmt), they diverged by an average genetic distance of 16.1% if the GG3 (*furvus*) and GG2 (*angustirostris*) were pooled together, but with an average distance of 14.2% if they were separated.

**Table 5.** Percentage genetic divergence (%) of the Tamura and Nei genetic distances, *sensu* Bradley and Baker (2001), for the Cytochrome-b gene in the genetic groups (GG1–4) of the ingroup (a) and in all the species examined herein (b). Genetic distances between pairs of entities appear below the diagonal the numbers on diagonal in the genetic groups are distances within populations (GLs).

a. Genetic Groups	GL#. Name, State	GL1	GL4	GL6	GL7	GL8	GL10	GL13	GL18
<b>GG1 <i>latirostris</i> Pl</b>	1. Xilitla, San Luis Potosí	0.0							
	4. Tlanchinol, Hidalgo	4.4	0.0						
	6. Zacualpan, Veracruz	4.5	1.1	0.0					
<b>GG2 <i>angustirostris</i> Pa</b>	7. Tenango de Doria, Hidalgo	5.0	0.7	1.2	0.0				
	8. Metepec, Hidalgo	4.1	1.2	0.7	1.2	0.01			
	10. Huachinango, Puebla	4.9	2.4	2.5	2.7	2.6	0.0		
<b>GG3 <i>furvus</i> Pf</b>	13. Xalapa, Veracruz	3.8	2.4	2.8	3.0	2.7	2.5	0.01	
<b>GG4 unknown Oax18</b>	18. Puerto de la Soledad	7.8	7.2	7.3	6.7	1.2	6.6	7.6	0

b. All OTUs	Oba	Mcr	Poc	Pml	Pmt	Pl	Pff	Pfa	Oax18
<i>Osgoodomys banderanus</i> , Oba									
<i>Megadontomys cryophilus</i> , Mcr	16.7								
<i>Peromyscus ochraventer</i> , Poc	20.3	15.9							
<i>P. melanocarpus</i> , Pml	17.9	14.1	22.9						
<i>P. mexicanus totontepecus</i> , Pmt	18.9	13.5	21.9	12.5					
<i>P. latirostris</i> / <i>P. f. latirostris</i> , Pl	15.8	13.7	16.9	15.0	14.0				
<i>P. f. furvus</i> , Pff	16.9	13.3	19.1	16.8	13.8	5.0			
<i>P. f. angustirostris</i> Pfa	16.1	13.1	19.8	15.6	14.1	4.0	2.8		
<b>Unknown <i>Peromyscus</i> from Oax18</b>	16.8	11.8	17.8	17.1	14.9	8.4	7.3	8.1	

GG1–4 differed from each other with lower average genetic distances (5.9 %, 2.8 to 8.4 %; Table 5b). Puerto de la Soledad (GG4 *Oax18*) had an average genetic distance of 7.9 % from GG1–3. GG1 (*latirostris*) differed from GG2–4 by 5.8 %, and GG3 (*furvus*) and GG2 (*angustirostris*) diverged from the other three GGs by 5 %. Either pooled together or alone, the two latter GGs had an average divergence of 6.0 to 6.1 % from the two geographically extreme GG1 (*latirostris*) and GG4 (*Oax18*), but were 7.7 % from GG4 and 4.5 % from GG1. The least average genetic distance (2.8 %) was between GG2 (*furvus*) and GG3 (*angustirostris*).

Finally, the average genetic distance among the OGs was 17.5 % (12.5 to 22.9 %). *P. ochraventer* (*Poc*, 20.3 %) had the most divergent average genetic distance, followed by *O. banderanus* (*Oba*, 18.5 %). *P. melanocarpus* (*Pml*, 16.9 %) and *P. mexicanus totontepecus* (*Pmt*, 16.7 %) and *M. cryophilus* (*Mcr*, 15.1 %) showed lower genetic divergence. The least divergent OGs were *Pml* and *Pmt* (12.5 %), and the most divergent were *Poc* and *Pml* (22.9 %).

## Discussion

The results presented herein were obtained from several analyses, including single-character phylogenies and all methods combined (MP, ML, and BI). Regardless of method, GG1 (*latirostris*, *Pl*), GG3 (*furvus*, *Pf*), and GG4 (*Oax18*) were always distinct because their sequences were reciprocally monophyletic. All analyses of the *Cytb* gene and the MP analyses for the NADH genes yielded a topology nearly identical to that reported by [Harris et al. \(2000:2131–2\)](#). However, the probabilistic topologies of the NADH genes concurred with the findings of [Ávila-Valle et al. \(2012:171\)](#). In [Harris et al. \(2000\)](#), the most distinctive entity was GG4 (*Oax18*), followed by GG1 (*Pl*), whereas in [Ávila-Valle et al. \(2012\)](#) the latter was the most distinctive entity and GG4 (*Oax18*) was associated with GG2 (*Pa*). Although these authors pointed out the distinctiveness of these two GGs, they did not propose their formal recognition. It is clear that the populations in the intermediate zone of the distribution range posed a challenge to interpreting the phylogenetic relationships among them, especially because their subpopulations (GLs) were not monophyletic in several of their topologies ([Harris et al. 2000; Ávila-Valle et al. 2012](#)). Aside from Huauchinango, Puebla (GL10), Otongo, some sequences of Tlanchinol, Tenango de Doria from Hidalgo, and some sequences of Mesa de la Yerba and Xalapa from Veracruz, combinations of sequences from different GLs (i.e., Zacualpan-Metepec-Tenango de Doria) had greater support than did all sequences representative of GG2 (*Pa*) in topologies of single types of genes. Similar results were obtained from the variance components in the AMOVA and in the haplotype networks herein.

Differences in the outcomes of these topologies were based on the method (non-parametric MP, parametric ML, BI; e.g., [Steel and Penny 2001; Sober 2004](#)), especially in the methods used to calculate branch length; probably due to different ontological and epistemological assump-

tions ([De Luna 1995](#)). Another possible source for discordance among topologies could have been from the evolutionary model used in the probabilistic methods and its relationship with the evolutionary assumptions in the MP analysis. The GTR I + G (GTR) used in the *Cytb* is more realistic, though more complex and thus with more probability of error because it considers six substitution parameters; whereas, the more simple model TMP 2 uf + G (TMP2) only has three parameters ([Posada 2008](#)). The GTR model, by assigning the same importance to the changes from one base to another and vice versa, is more like the Farris optimization model in MP analysis ([Farris 1970; Lipscomb 1998](#)). However, GTR considers different probabilities for the different types of mutations ([Tavare 1986](#)) and assigns a specific value to particular types of mutations. Conversely, the TPM2 model accepts and evaluates only three forms of base substitutions ([Kimura 1981](#)), thus ignoring possible informative mutations for phylogenetic analyses, unlike the [Farris \(1970\)](#) optimization. Use of GTR as an evolutionary model for the *Cytb*, with or without invariant sites, is quite frequent in studies on the systematics of *Peromyscus* (e.g., [Harris et al. 2000; Bradley et al. 2007; Bradley et al. 2014; Platt et al. 2015](#); and other references therein and herein). However, exploration of TPM2 in *Peromyscus* ([Walker and Greenbaum 2006; Ávila-Valle et al. 2012; Castañeda-Rico et al. 2014](#)) and other species ([León-Paniagua et al. 2007](#)) is just beginning.

[Ávila-Valle et al. \(2012\)](#) also used the GTR model and obtained the same topology as presented herein for the NADH genes, even when they examined only whole ND3-ND4 sequences (1,043 base pairs). Therefore, the differences between the topologies may be due to a different evolutionary rate for nucleotide substitution (synonymous and non-synonymous) between the mitochondrial genes. *Cytb* evolves faster than do any of ND3, ND4L and ND4 genes in other mammalian orders (primates, carnivores, perissodactyls, and cetaceans; [Pesole et al. 1999](#)) and may have caused the discordance among topologies. For non-synonymous mutations, the rate of change for *Cytb* is at least two times faster than in any of the NDAH genes. For synonymous mutations, the rate of change in ND3 is 50 % less than that of the ND4 and *Cytb*, and the rate of change in ND4L is 25 % less than the *Cytb* and NADH genes. Therefore, GG4 may have grouped with GG2 and GG3 because of the influence of the ND3 and ND4L genes, whereas sequences from *Cytb* clearly separated it from the other GGs. The rate of change in *Cytb* has proven to be adequate to discern both intraspecific (intrapopulation, intrasubspecific, intra-specific) and intrageneric (sister species, interspecific) variation *sensu* [Bradley and Baker \(2001\)](#). It also is likely that our adjustment in the number of bases in the GL (*Pl*) and the OG in the ND3-ND4L-ND4 genes allowed us to avoid possible inconsistencies between the sequences recovered from the literature and GenBank due to indels.

In our integrated phylogeny, based on all characters, we used the same epistemological approach to perform our

analyses under the same evolutionary model (Farris optimization) and method (MP) in the TNT package ([Goloboff et al. 2006](#)). The decision to overcome technical restrictions for analyzing a big character matrix with different type characters allowed us to test the behavior of NADH genes under a more open evolutionary scenario. Moreover, it allowed us to include continuous morphometric characters for the shape and size of skull ([Goloboff et al. 2006; Ramírez-Sánchez et al. 2016](#)). As with the molecular data, we conducted single-character phylogenies for each size (linear measurements) and shape characters (morphogeometric configurations) with a much larger amount of individuals (in prep.), especially for size (*i. e.*, those used in [Ávila-Valle et al. 2012](#)). Herein, morphometric characters solved the discrepancies among GLs in GG2 (*Pa*, Figure 2) in spite of reluctance of including them within combined molecular and morphometric phylogenies ([Scotland et al. 2003](#)). In other studies (*e. g.*, [Lee and Camens 2009; Ledevin and Millien 2013](#)), incorporation of morphometric characters into integrated phylogenies with molecular data has proven to be useful to understand and solve conflicting molecular outcomes.

After reduction of the number of genetic sequences in order to include only the GLs with both sequences, followed by an even more reduced number of GLs with all genetic and morphometric characters, GG1–4 appeared as well recognizable geographic and evolutionary entities with better supporting values compared to single-characters topologies. Herein, specimens from Puerto de la Soledad (GG4, *Oax18*), in the farther southern range, show the greatest genetic distances from the other GGs, and its geographic remoteness. Likewise, specimens (GG1, *latirostris*) from Xilitla, San Luis Potosí, with the second greatest geographic distances, showed twice the genetic divergence from specimens in the intermediate range. Finally, sequences from Xalapa, Veracruz and its surrounding locations (GG3, *furus*), as well as those from Tlanchinol and Tenango de Doria, Hidalgo, and Zacualpan, Veracruz (GG2, *angustirostris*), were distinctly separated from each other as the subunits of a third genetic clade. Therefore, the monotypic status currently assigned to the populations in the overall distribution of *P. furvus* ([Rogers and Skoy 2011](#)) does not coincide with our genetic nor the morphometric data. Rather, the data presented herein confirms the morphometric ([Musser 1964; Hall 1968; Martínez-Coronel et al. 1997; Ávila-Valle et al. 2012](#)) and biochemical ([Harris and Rogers 1999; Harris et al. 2000; Ávila-Valle et al. 2012](#)) discrepancies reported previously.

**Systematic and taxonomic remarks.** Application of the genetic species concept ([Wiley and Mayden 2000](#)) *sensu* [Baker and Bradley \(2006\)](#), allows us to suggest the taxonomic level (clades and haplotypes clearly forming monophyletic groups) and the subspecific level (clades and shared haplotypes) of the distinctive genetic entities in the *Cytb* dataset. All GG1–4, with GG2–3 either separated (mean 14.2 %, 13.8 to 14.9 %) or pooled (16.1 %, 15.3 to 16.6 %), were similar in levels of genetic divergence as other congeneric species in

the OG. When *M. cryophilus* and *O. banderanus* are included, the divergence level increased (15.6 %, 15.1 to 16 %) for the two genes. These levels of genetic distances among GG1–4 indicate evidence supporting the recognition of genetic entities *sensu* [Bradley and Baker \(2001\)](#). The population of Puerto de la Soledad, Oaxaca (GG4) possessed the greatest genetic diversity of the four populations (GG1–4). GG4 had an average genetic distance of 7.5 % (or of 7.9 %, when the 10 OTUs were included), which is sufficient to distinguish it as a distinctive species from the other populations (GG1, *latirostris*; GG2, *angustirostris*, and GG3, *furus*) *sensu* [Bradley and Baker \(2001\)](#) and [Baker and Bradley \(2006\)](#).

The northern population, (GG1, *latirostris*), possessed an average genetic distance of 4.2 % among GG1–3 (and of 4.9 % when the OGs are included in the analysis). [Baker and Bradley \(2006\)](#) mention a K2P distance interval of 2.8 to 10.8 % to validate species within *Peromyscus*, with several authors using a lesser distance than 7.0 %, although it is necessary to support genetic distance values with other kinds of evidence ([Bradley and Baker 2001; Bradley et al. 2014](#)). In the *boyliei* species group, in addition to the genetic data of *Cytb*, the fundamental number of chromosomes and/or morphometric data have been used to describe new species in which the average lower limit of (*i. e.*, the percentage genetic distance to its closest relative) is 4.3 %  $\pm$  1.1 SD (3.3 to 5.6 %). Following this logic, *P. schmidlyi* was described with a mean distance of 3.3 % from *P. levipes* ([Bradley et al. 2004a](#)), *P. carletoni* from *P. levipes* (3.4 %, [Bradley et al. 2014](#)), and *P. kilpatricki* from *P. levipes* (5.2 %, [Bradley et al. 2016](#)). Similarly, in the *mexicanus* species group, [Lorenzo et al. \(2016\)](#) described *P. gardneri* based on a K2P distance of 3.6 to 4.2 % from other species and reinforced the decision with morphometric data. In our analysis of linear (size) and morphogeometric (shape) characters, GG1 (*latirostris*) behaved as a separate entity, similar to that found in the morphometric data of [Bradley et al. \(2004a\)](#) and [Lorenzo et al. \(2016\)](#). In addition, GG1 (*latirostris*) has a larger skull size compared to GG2–4 ([Martínez-Coronel et al. 1997; Ávila-Valle et al. 2012](#)) and some distinctive qualitative cranial traits have been reported ([Hall and Alvarez 1961; Musser 1964; Hall 1968](#)).

GG2 (*angustirostris*) and GG3 (*furus*) also possessed N-T genetic distances that averaged greater or similar to those K2P values reported for *Peromyscus* ([Tiemann-Boege et al. 2000; Durish et al. 2004; Ordóñez-Garza et al. 2010](#)). The average genetic distance between GG3 and GG2 was 2.7 % and 2.8 %, compared to other OTUs. Based on the logic presented in [Bradley and Baker \(2001\)](#), our results revealed that GG3 (*furus* from the type locality Xalapa, Veracruz and its surroundings) and GG2 (*angustirostris* from Hidalgo, Puebla and locations further NW in Veracruz) would appear to be subspecies. The genetic distances between GG2 and GG3 are similar to that reported by [Bradley et al. \(2015\)](#) for two subspecies (ranging from 1.77 % in *P. p. laceianus* to 2.21 % in *P. p. pectoralis*). Similarly, [Lorenzo et al. \(2016\)](#) use a K2P genetic distance of 2.08–3.65% to define two subspe-

cies of *P. zarhynchus* in Chiapas (*P. z. zarhynchus* and *P. z. cristobalensis*). Although our analyses were based on N-T distances, our overall conclusion concurs with the framework of [Bradley and Baker \(2001\)](#).

Finally, we suggest the possibility of additional subspecies within *P. furvus*. For example, ambiguities in the phylogenetic topologies were found by [Harris et al. \(2000\)](#) and [Avila-Valle et al. \(2012\)](#), as well as in haplotype networks presented herein. This is especially true for populations assigned herein to *P. f. angustirostris* to the north of Puebla (Huauchinango) and to the SE of the type locality (Zacualpan, Veracruz) in Hidalgo (i. e., Tlanchinol, Metepec, and Tenango de Doria). Sequences from the intermediate group localities are needed to resolve this issue. Further, sequences are needed from geographic areas such as Molango, Hidalgo, and from vegetational transition zones between Puebla and Veracruz (e. g., [Rogers and Skoy 2011](#); [Peralta-Moctezuma 2011](#)).

*Remarks for the furvus species group.* *Peromyscus furvus* as currently conceived (*sensu lato*), contains multiple species and subspecies. The population of Puerto de la Soledad Oaxaca (GG4, Oax18) is a *species nova*, distinctive from *P. furvus* by its evolutionary divergence. This interpretation agrees with the genetic distances reported by [Harris and Rogers \(1999\)](#) for the allozyme PGDH, as well as with the possible biogeographic scenario proposed by [Harris et al. \(2000\)](#). *Peromyscus latirostris* from Xilitla, San Luis Potosí, and Santa Inés, Querétaro, is either a sister species to *P. furvus*, as suggested initially by [Dalquest \(1950\)](#), or is valid subspecies, *P. f. latirostris*. Our morphometric analyses (in prep.) strongly support the recognition of a separate species. Finally, populations along the intermediate zone (Hidalgo, Puebla, and Veracruz) represent at least two different subspecies, *P. furvus angustirostris* (as described by [Hall and Álvarez 1961](#)) and *P. f. furvus* (as described by J. A. [Allen and Chapman 1897](#)). Based on these findings, the *furvus* species group now comprises three species (*P. latirostris*, *P. species nova*, and *P. furvus*) and at least two subspecies (*P. f. angustirostris* and *P. f. furvus*).

The pairing of *P. m. totontepecus* and *P. melanocarpus* agrees with the observation by [Bradley et al. \(2007\)](#) that *P. melanocarpus* did not seem to belong to the *furvus* species group, as it was more closely associated with species of the *mexicanus* species group (see [Hooper 1968](#), [Huckaby 1980](#)). *Peromyscus ochraventer*, which also was considered a member of the *furvus* species group ([Carleton 1989](#)), behaved as an even more distant taxon from the *furvus* species group, as defined here and by others such as [Wade et al. \(1999\)](#) and [Musser and Carleton \(2005\)](#). Herein, *P. ochraventer* was more basal in all phylogenetic analyses, suggesting that this species is closer to the more derived *difficilis* species group ([Durish et al. 2004](#); [Bradley et al. 2007](#)). The grouping of *M. cryophilus* with *P. ochraventer* in the OG is consistent with recent *Cytb* gene phylogenies, where this species and *M. thomasi* were genetically closer to *P. furvus* and the Oaxacan *species nova* ([Bradley et al. 2007:1115](#)) and to other

subgenera such as *Podomys* and *Neotomodon* ([Platt et al. 2015:712](#)). Finally, except for the single molecular phylogenies with probabilistic methods in the NADH genes, *O. banderanus* was always basal and remained segregated from the other taxa. Its position at the base of the topology suggests an affiliation to the subgenus *Haplomylomys*, the basal most subgenus of *Peromyscus* in recent phylogenies of the genus ([Bradley et al. 2004b, 2007](#); [Platt et al. 2015](#)).

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## Literature Cited

ALLEN, J. A., AND F. M. CHAPMAN. 1897. On a collection of mammals from Jalapa and Las Vigas, State of Veracruz, Mexico. Bulletin of the American Museum of Natural History 9:201–203.

ÁVILA-VALLE, Z. A., A. CASTRO-CAMPILLO, AND L. LEÓN PANIAGUA. 2005. Revisión del Estado Sistemático de *Peromyscus furvus* (Rodentia: Muridae), México, a Partir de los Genes Mitocondriales ND3- ND4. División de Ciencias Biológicas y de la Salud, Universidad Autónoma Metropolitana, Unidad Iztapalapa. Tesis de Maestría. Ciudad de México, México.

ÁVILA-VALLE, Z. A., A. CASTRO-CAMPILLO, L. LEÓN-PANIAGUA, I. H. SALGADO-UGARTE, A. G. NAVARRO-SIGÜENZA, E. B. HERNÁNDEZ-BÁÑOS, AND J. RAMÍREZ-PULIDO. 2012. Geographic variation and molecular evidence of the Blackish Deer Mouse complex (*Peromyscus furvus*, Rodentia: Muridae). Mammalian Biology 77:166–177.

**BAKER, R. J., AND R. D. BRADLEY.** 2006. Speciation in mammals and the Genetic Species Concept. *Journal of Mammalogy* 87:643–662.

**BOOKSTEIN, F. L.** 1991. *Morphometric Tools for Landmark Data. Geometry and Biology*. Cambridge University Press. New York, U.S.A.

**BRADLEY, R. D., AND R. J. BAKER.** 2001. A test of the genetic species concept: Cytochrome-*b* sequences and mammals. *Journal of Mammalogy* 82:960–973.

**BRADLEY, R. D., D. S. CARROLL, M. L. HAYNIE, R. M. MARTÍNEZ, M. J. HAMILTON, AND C. W. KILPATRICK.** 2004a. A New Species of *Peromyscus* from Western Mexico. *Journal of Mammalogy* 85:1184–1193.

**BRADLEY, R. D., N. D. DURISH, D. S. ROGERS, J. R. MILLER, M. D. ENGSTROM, AND C. W. KILPATRICK.** 2007. Toward a molecular phylogeny for *Peromyscus*: Evidence from mitochondrial Cytochrome-*b* sequences. *Journal of Mammalogy* 88:1146–1159.

**BRADLEY, R. D., C. W. EDWARDS, D. S. CARROLL, AND C. W. KILPATRICK.** 2004b. Phylogenetic relationships of Neotomine–Peromyscine rodents: Based on DNA sequences from the mitochondrial Cytochrome-*b* gene. *Journal of Mammalogy* 85:389–395.

**BRADLEY, R. D., N. ORDÓÑEZ-GARZA, G. CEBALLOS, D. S. ROGERS, AND D. J. SCHMIDLY.** 2016. A new species in the *Peromyscus boylii* species group (Cricetidae: Neotominae) from Michoacán, México. *Journal of Mammalogy* 98:154–165.

**BRADLEY, R. D., N. ORDÓÑEZ-GARZA, C. G. SOTERO-CAIO, H. M. HUYNH, C. W. KILPATRICK, L. I. IÑIGUEZ-DÁVALOS, AND D. J. SCHMIDLY.** 2014. Morphometric, karyotypic, and molecular evidence for a new species of *Peromyscus* (Cricetidae: Neotominae) from Nayarit, Mexico. *Journal of Mammalogy* 95:176–186.

**BRADLEY, R. D., D. J. SCHMIDLY, B. R. AMMAN, R. N. PLATT, K. M. NEUMANN, H. M. HUYNH, R. MUÑIZ-MARTÍNEZ, C. LÓPEZ-GONZÁLEZ, AND C. ORDÓÑEZ-GARZA.** 2015. Molecular and morphologic data reveal multiple Species in *Peromyscus pectoralis*. *Journal of Mammalogy* 96:446–459.

**CARLETON, M. D.** 1989. Systematics and evolution. Pp. 7–141, in *Advances in the Study of *Peromyscus* (Rodentia)* (Kirkland, G. L., and J. N. Layne, eds.). Texas Tech University Press. Lubbock, U.S.A.

**CASTAÑEDA-RICO, S. S., L. LEÓN-PANIAGUA, E. VÁZQUEZ-DOMÍNGUEZ, AND A. NAVARRO-SIGÜENZA.** 2014. Evolutionary diversification and speciation in rodents of the Mexican lowlands: The *Peromyscus melanophrys* species group. *Molecular Phylogenetics and Evolution* 70:454–463.

**CORDEIRO-ESTREIA P., M. BAYLAC, C. DENYS, AND J. POLOP.** 2008. Combining geometric morphometrics and pattern recognition to identify interspecific patterns of skull variation: Case study in sympatric Argentinian Species of the Genus *Calomys* (Rodentia: Cricetidae: Sigmodontinae). *Biological Journal of the Linnean Society* 94:365–378.

**CORDERO, G. A., AND C. W. EPPS.** 2012. From desert to rainforest: phenotypic variation in functionally important traits of Bushy-tailed Woodrats (*Neotomacinerea*) across two climatic extremes. *Journal of Mammalian Evolution* 19:135–153.

**CRUZ-GÓMEZ A., A. CASTRO-CAMPILLO, AND L. LEÓN-PANIAGUA.** 2018. Filogenia integrada de *Peromyscus furvus* con caracteres morfométricos y moleculares. División de Ciencias Biológicas y de la Salud, Universidad Autónoma Metropolitana, Universidad Iztapalapa. Tesis de Maestría. Ciudad de México, México.

**DALQUEST, W. W.** 1950. Records of mammals from the Mexican State of San Luis Potosí. *Occasional Papers of the Museum of Zoology, Louisiana State University* 23:1–15.

**DE LUNA, E.** 1995. Philosophical bases of cladistic analyses for taxonomic research. *Acta Botánica Mexicana* 33:63–79.

**DURISH, N. D., K. E. HALCOMB, C. W. KILPATRICK, AND R. D. BRADLEY.** 2004. Molecular systematics of the *Peromyscus truei* Species Group. *Journal of Mammalogy* 85:1160–1169.

**EXCOFFIER, L., G. LAVAL, AND S. SCHNEIDER.** 2005. Arlequin v3.0. An integrated software package for population genetics data analysis. *Evolutionary Bioinformatics (online)* 1:47–50.

**EXCOFFIER, L., P. E. SMOUSE, AND J. M. QUATTRO.** 1992. Analysis of molecular variance inferred from metric distances among DNA haplotypes: application to human mitochondrial DNA restriction data. *Genetics* 131:479–491.

**FARRIS, J. S.** 1970. Methods for computing Wagner trees. *Systematic Biology* 19:83–92.

**FELSENSTEIN, J.** 2005. *PHYLIP (Phylogeny Inference Package)*, Version 3.6. Distributed by the Author. Department of Genome Sciences, University of Washington, Seattle, WA, USA. <http://evolution.genetics.washington.edu/phylip.html>

**GOLOBOFF, P. A., J. S. FARRIS, AND K. C. NIXON.** 2008. TNT, Free Program for Phylogenetic Analysis. *Cladistics* 24:774–786.

**GOLOBOFF, P. A., C. I. MATTONIY, AND A. S. QUINTEROS.** 2006. Continuous characters analyzed as such. *Cladistics* 22:589–601.

**GRIECO, T. M., AND O. T RIZK.** 2010. Cranial shape varies along an elevation gradient in Gambel's White-footed Mouse (*Peromyscus maniculatus gambelii*) in the Grinnell Resurvey Yosemite transect. *Journal of Morphology* 271:897–909.

**GUINDON, S., AND O. GASCUEL.** 2003. A simple, fast, and accurate algorithm to estimate large phylogenies by maximum likelihood. *Systematic Biology* 52:696–704.

**HALL, E. R.** 1968. Variation in the Blackish Deer Mouse, *Peromyscus furvus*. *Anales del Instituto de Biología, Universidad Nacional Autónoma de México* 1:149–154.

**HALL, E. R., AND T. ÁLVAREZ.** 1961. A new species of mouse (*Peromyscus*) from northwestern Veracruz, México. *Proceedings of the Biological Society of Washington* 74:203–206.

**HAMMER, Ø., D. A. T. HARPER, AND P. D. RYAN.** 2001. PAST: Paleontological statistics software package for education and data analysis. *Palaeontologia Electronica* 4:9.

**HARRIS, D. J., AND D. S. ROGERS.** 1999. Species limits and phylogenetic relationships among populations of *Peromyscus furvus*. *Journal of Mammalogy* 80:530–544.

**HARRIS, D. J., D. S. ROGERS, AND J. SULLIVAN.** 2000. Phylogeography of *Peromyscus furvus* (Rodentia: Muridae) based on Cytochrome-*b* Sequence data. *Molecular Ecology* 9:2129–2135.

**HOLMES, M. W., G. K. BOYKINS, R. C. BOWIE, AND E. A. LACEY.** 2016. Cranial morphological variation in *Peromyscus maniculatus* over nearly a century of environmental change in three areas of California. *Journal of Morphology* 277:96–106.

**HOOPER, E. T.** 1968. Classification. Pp. 27–74, in *Biology of *Peromyscus* (Rodentia)* (King, J. A, ed.). Special Publication no 2, American Society of Mammalogists. Lawrence, U.S.A.

**HUCKABY, D. G.** 1980. Species limits in the *Peromyscus mexicanus* group (Mammalia: Rodentia: Muroidea). *Contributions in Science, Natural History Museum of Los Angeles County* 326:1–24.

**KIMURA, M.** 1980. A simple method for estimating evolutionary rate of base substitutions through comparative studies of nucleotide sequences. *Journal of Molecular Evolution* 16:111–120.

**KIMURA M.** 1981. Estimation of evolutionary distances between homologous nucleotide sequences. *Proceedings of the National Academy of Sciences* 78:454–458.

**KUMAR, S., G. STECHER, AND K. TAMURA.** 2016. MEGA7: Molecular Evolutionary Genetics Analysis Version 7.0 for bigger datasets. *Molecular Biological Evolution* 33:1870–4.

**LEDEVIN, R., AND V. MILLIEN.** 2013. Congruent morphological and genetic differentiation as a signature of range expansion in a fragmented landscape. *Ecology and Evolution* 3:4172–4182.

**LEE, M. S. Y., AND A. B. CAMENS.** 2009. Strong morphological support for the molecular evolutionary tree of placental mammals. *Journal of Evolutionary Biology* 22:2243–2257.

**LEÓN-PANIAGUA L., A. G. NAVARRO-SIGÜENZA, B. HERNÁNDEZ-BAÑOS B, AND J. C. MORALES.** 2007. Diversification of the arboreal mice of the Genus *Habromys* (Rodentia: Cricetidae: Neotominae) in the Mesoamerican highlands. *Molecular Phylogenetics and Evolution* 42:653–664.

**LIBRADO, P., AND J. ROZAS.** 2009. DnaSP v5: A software for comprehensive analysis of DNA polymorphism data. *Bioinformatics* 25:1451–1452.

**LIGHT, J. E., M. O. OSTROFF, AND D. J. HAFNER.** 2016. Phylogeographic assessment of the Northern Pygmy Mouse, *Baiomys taylori*. *Journal of Mammalogy* 97:1081–1094.

**LIPSCOMB D.** 1998. Basics of Cladistic Analysis. George Washington University. District of Columbia, U.S.A. Available from: <http://citeseerx.ist.psu.edu/viewdoc/summary?doi=10.1.1.329.9770>.

**LORENZO, C., S. T. ÁLVAREZ-CASTAÑEDA, S. G. PÉREZ-CONSUEGRA, AND J. L. PATTON.** 2016. Revision of the Chiapan Deer Mouse, *Peromyscus zarhynchus*, with the description of a new Species. *Journal of Mammalogy* 97:910–918.

**MARTÍNEZ-CORONEL, M., A. CASTRO-CAMPILLO, AND J. RAMÍREZ-PULIDO.** 1997. Variación no geográfica de *Peromyscus furvus* (Rodentia: Muridae). Pp. 183–203, in Homenaje al Profesor Ticul Álvarez (Arroyo Cabrales, J., and O. J. Polaco, eds.). Colección Científica, Instituto Nacional de Antropología e Historia. Ciudad de México, México.

**MUSSER, G. G.** 1964. Notes on geographic distribution, habitat, and taxonomy of some Mexican mammals. *Occasional Papers of the Museum of Zoology*, University of Michigan 636:1–22.

**MUSSER, G. G., AND M. D. CARLETON.** 2005. Superfamily Murioidea. Pp. 894–1531, in *Mammal Species of the World: A Taxonomic and Geographic Reference* (Wilson, D. E. and D. M. Reed, eds.) third edition. John Hopkins University Press. Baltimore, U.S.A.

**MYERS, P., B. L. LUNDIGAN, B. W. GILLESPIE, AND M. L. ZELDITCH.** 1996. Phenotypic plasticity in skull and dental morphology in the Prairie Deer Mouse (*Peromyscus maniculatus bairdii*). *Journal of Morphology* 229:229–237.

**ORDÓÑEZ-GARZA, N., J. O. MATSON, R. E. STRAUSS, R. D. BRADLEY, AND J. SALAZAR-BRAVO.** 2010. Patterns of phenotypic and genetic variation in three species of endemic Mesoamerican *Peromyscus* (Rodentia: Cricetidae). *Journal of Mammalogy* 91:848–859.

**PERALTA-MOCTEZUMA, J. V.** 2011. Estado actual de los mamíferos silvestres de la Sierra Norte de Puebla. Escuela de Biología, Benemérita Universidad Autónoma de Puebla. Tesis de Licenciatura. Puebla de los Ángeles, México.

**PÉREZ-CONSUEGRA, S. E., AND E. VÁZQUEZ-DOMÍNGUEZ.** 2015. Mitochondrial diversification of the *Peromyscus mexicanus* Species Group in nuclear Central America: Biogeographic and taxonomic implications. *Journal of Zoological Systematics and Evolutionary Research* 53:300–311.

**PESOLE, G., C. GISSI, A. DE CHIRICO, AND C. SACCONI.** 1999. Nucleotide substitution rate of mammalian mitochondrial genomes. *Journal of Molecular Evolution* 48:427–434.

**PLATT, R. N., B. R. AMMAN, M. S. KEITH, C. W. THOMPSON, AND R. D. BRADLEY.** 2015. What is *Peromyscus*? Evidence from nuclear and mitochondrial DNA sequences suggests the need for a new classification. *Journal of Mammalogy* 96:708–719.

**POSADA, D.** 2008. jModelTest: Phylogenetic model averaging. *Molecular Biology and Evolution*, 25:1253–1256.

**RAMÍREZ-PULIDO, J., N. GONZÁLEZ-RUIZ, A. L. GARDNER, AND J. ARROYO-CABRALES.** 2014. List of recent land mammals of Mexico, 2014. Special Publications, Museum of Texas Tech University 63:1–69.

**RAMÍREZ-SÁNCHEZ, M., E. DE LUNA E, AND C. CRAMER.** 2016. Geometric and traditional morphometrics for the assessment of character state identity: Multivariate statistical analyses of character variation in the Genus *Arrenurus* (Acari, Hydrachnidia, Arrenuridae). *Zoological Journal of the Linnean Society* 177:720–749.

**ROGERS, D. S., AND J. A. SKOY.** 2011. *Peromyscus furvus* (Rodentia: Cricetidae). *Mammalian Species* 43:209–215.

**ROHLF, F. J.** 2015. The tps series of software. *Hystrix, the Italian Journal of Mammalogy* 26:9–12.

**RONQUIST, F., M. TESLENKO, P. VAN DER MARK, D. L. AYRES, A. DARLING, S. HÖHNA, AND J. P. HUELSENBECK JP.** 2012. MrBayes 3.2: Efficient Bayesian phylogenetic inference and model choice across a large model space. *Systematic Biology* 61:539–542.

**SCOTLAND, R. W., R. G. OLSTEAD, AND J. R. BENNETT.** 2003. Phylogeny reconstruction: the role of morphology. *Systematic Biology* 52:539–548.

**SHEETS, H. D.** 2002. Morphometric Software IMP-Integrated Morphometrics Package. Department of Geology. Buffalo, U.S.A.

**SHEETS, H. D., K. KIM, AND C. E. MITCHELL.** 2004. A combined landmark and outline-based approach to ontogenetic shape change in the Ordovician trilobite *Triarthrusbecki*. Pp. 67–82, in *Morphometrics—Applications in Biology and Paleontology* (Elewa, A. M. T., ed.). Springer-Verlag. Heidelberg, Germany.

**SOBER, E.** 2004. The contest between parsimony and likelihood. *Systematic Biology*, 53:644–653.

**STEEL, M., AND D. PENNY.** 2000. Parsimony, likelihood, and the role of models in molecular phylogenetics. *Molecular Biology and Evolution* 17:839–850.

**TAMURA, K., AND M. NEI.** 1993. Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees. *Molecular Biology and Evolution* 10:512–526.

**TAVARE, S.** 1986. Some probabilistic and statistical problems in the analysis of DNA sequences. *Lectures on mathematics in the life sciences* 17:57–86.

**TIEMANN-BOEGE, I., C. W. KILPATRICK, D. J. SCHMIDLY, AND R. D. BRADLEY.** 2000. Molecular phylogenetics of the *Peromyscus boylii* Species Group. *Molecular Phylogenetics and Evolution* 16:366–378.

**WADE, N. L., M. ENGSTROM, AND A. BAKER.** 1999. Molecular Systematics of Neotropical Deer Mice of the *Peromyscus mexicanus* Species Group. Graduate Department of Zoology, University of Toronto. Master Thesis. Toronto, Canada.

**WALKER, M. L., AND I. F. GREENBAUM.** 2006. Mitochondrial-DNA variation and the evolutionary affinities of the *Peromyscus maniculatus* complex from western North America. PhD Thesis. Texas A&M University. College Station, U.S.A.

**WILEY, E. O., AND R. L. MAYDEN.** 2000. The evolutionary species concept. Pp. 70- 89, in *Species Concepts and Phylogenetic Theory: A Debate* (Wheeler, Q. D., and R.: Meier, R., eds.). Columbia University Press. New York, USA.

**ZELDITCH, M. L., D. L SWIDERSKI, H. D. SHEETS, AND W. L. FINK.** 2004. *Geometric Morphometrics for Biologists: A Primer*. Academic Press. Elsevier. Amsterdam, Neaderlands.

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## Appendix 1

Sequences of four mitochondrial genes examined in the analyses. Mitochondrial genes included 719 bases of the Cytochrome-*b* gene (*Cytb* gene), and 971 bases comprising three genes (ND3-ND4L-ND3) of the coenzyme 1, Nicotinamide Adenine Dinucleotide Hydride (NADH). Data are arranged by operational taxonomic units (OTUs), comprise in parenthesis: Genetic Groups (GG1–4) and Ingroup (IG), assigned according to its geographic distribution (Figure 1), and former taxonomic designations with its respective key in italics. The other five OTUs are Outgroups (OGs). After each type of mitochondrial gene, with its total sample size inside parenthesis, follows the Mexican State, number and name of group locality (GL#; see map in Figure 1b), and numbered specific localities (SLs) *sensu* Cruz-Gómez *et al.* (2018). An asterisk (\*) indicates close GLs to topotypic populations (GG1–3) or the first sampled Oaxacan populations (GG4). Following the SLs the list continues with the codes for the sequences inside parenthesis (abbreviations of GLs names + specimens consecutive numbers: 1–n, where n = total number of sequences) and their GenBank accession numbers (GnBnk#). In the list, note that:

- a. All 130 sequences were used in phylogenies and genetic analyses of variation and divergence, according to type of mitochondrial genes.
- b. Sequences and codes inside brackets were used in the integrated molecular phylogenies and in haplotype networks (*Cytb*, GG1–4 n = 31, OGs n = 6; NADH genes, GG1–4 n = 29, OGs n = 5, Cruz-Gómez *et al.* 2018).
- c. Sequences and codes in italics, were used in the molecular integrated phylogeny with 29 sequences in each type of mitochondrial genes and one sequence for each one of the five OGs.

The total 53 *Cytb* sequences in GG1–4 previously were analyzed by Harris *et al.* (2000), while the 13 sequences in the OGs were analyzed elsewhere (a, Bradley *et al.* 2007; b, Light *et al.* 2016; c, Pérez-Consuegra and Vázquez-Domínguez 2015). Likewise, the total 57 sequences for the three NADH genes in the IG were analyzed by Ávila-Valle *et al.* (2012), along with seven more sequences for the OGs, examined there and in other studies (d, Ávila-Valle *et al.* 2012; e, Wade 1999; f, Ávila-Valle *et al.* 2005). Colors in GG1–4 correspond to the visual code used in figures.

OTUs currently assigned to *Peromyscus furvus*:

OTU1 (GG1, *latirostris*, *Pl*). *Cytb* gene (n = 8). San Luis Potosí: GL1 Xilitla\*. 3) Ejido Aguayo, 6.2 km N Xilitla\*, [Xili1–8]: [AF270981, AF270987, AF271001, AF271004, AF271006, AF271015, AF271016, AF271020]. NADH genes (n = 17, 11). San Luis Potosí: GL1 Xilitla\*. 3) Ejido Aguayo, 6.2 km N Xilitla\*, Xili1–13: [Xili1–8, JN885476, JN885477, JN885478, JN885479, JN885480, JN885481, JN885482, JN885483], JN885484, JN885485, JN885486, JN885487, JN885488. Querétaro: GL2 Santa Inés. 15) El Pemoche [Saln1, JN885489]; 22) 2.5 Km NW Santa Inés [Saln2–3, JN885490, JN885491]; 23) 2 Km W Santa Inés [Saln4–5, JN88549, JN885493].

OTU2 (GG2, *angustirostris*, *Pa*). *Cytb* gene (n = 29 [n = 14]). Hidalgo: GL4 Tlanchinol. 35) 3 km E Tlanchinol, Tlan1–7: [Tlan1–5, AF270989, AF270998, AF270999, AF2701002, AF2701008], AF2701023, AF2701028. Veracruz: GL6. Zacualpan\*. 65) La Colonia, 6.5 km W Zacualpan\*, [Zacu1–3, AF270985, AF270993, AF2701021]. GL7 Tenango de Doria. 80) El Potrero, 10 km SW Tenango de Doria, TeDo1–8: [TeDo 1–4, AF270991, AF271003, AF271005, AF271009], AF271010, AF271019, AF271024, AF271028. GL8 Metepec. 74) 21.8 km NE Metepec, [Metp1–2, AF271014, AF271026]. Puebla: GL10 Huauchinango. 82) Rancho El Paraíso, 6 km SW Huauchinango, Huau1–9: [Huau1–5, AF270982, AF270983, AF270984, AF270986, AF270988], AF270990, AF2701018, AF2701022, AF2701025. NADH genes (n = 24 [n = 11]). Hidalgo: GL3 Otongo, [Oton1–3, JN885501, JN885502, JN885503]. GL4. Tlanchinol. 35) 3 km E Tlanchinol, Tlan1–7: [Tlan1, JN885494; Tlan2–4, JN885495, JN885496, JN885497, JN885498], JN885499, JN885500. Veracruz: GL6 Zacualpan\*. 65) La Colonia, 6.5 km W Zacualpan\*, Zacu1–7: [Zacu1–3 JN885504, JN885505, JN885506], JN885507, JN885508, JN885509, JN885510. Hidalgo: GL7. Tenango de Doria. 80) El Potrero, 10 km SW Tenango de Doria, [TeDo1–4: JN885511, JN885512, JN885513, JN885514]. Puebla: GL9. Xicotepec de Juárez. 83) El Salto, [XiJu1–7, JN885515, JN885516, JN885517, JN885518, JN885519, JN885510, JN885521].

OTU3 (GG3, *furvus*, *Pf*). *Cytb* gene (n = 4). Veracruz: GL13 Xalapa\*. 115) Banderillas, 6 km NW Xalapa\* [Xalp1–4: AF270980, AF271030, AF271031, AF27103]. NADH genes (n = 11 [n = 10]). Veracruz: GL13b Mesa de la Yerba\*. 120) Mesa de la Yerba\*, MsYb1–6: [MsYb1–4, JN885522, JN885523, JN885524, JN885525], JN885526, JN885527. GL13a Xalapa\*. 116) 1.5 Km SE Banderillas\*, Xalp1–5: [Xalp1–4, JN885528, JN885529, JN885530, JN885531], JN885532.

OTU4 (GG4, unknown *Peromyscus* from Oaxaca, Oax18). *Cytb* gene (n = 12 [n = 4]). Oaxaca: GL18 Puerto de la Soledad\*. 157) 1.5 km S Puerto de la Soledad\*, PtSl1–12: [PtSl1–5, AF270992, AF270994, AF270995, AF270996, AF270997], AF271000, AF271007, AF271011, AF271012, AF271013, AF271017, AF271027. NADH genes (n = 4). Oaxaca. GL18 Puerto de la Soledad\*. 150) Puerto de la Soledad\*, [PtSl–5: JN885533, JN885534, JN885535, JN885536, JN885537].

Outgroups

Congeneric species formerly assigned to the *furvus* species group (sister Outgroups):

OTU6 (*Peromyscus melanocarpus*, *Pml*). *Cytb* gene (n = 2 [n = 1]). Oaxaca: La Esperanza, *Pml*1–2: [*Pml*, EF028173<sup>a</sup>], CMC29192<sup>a</sup>. NADH genes (n = 2 [n = 1]). No data, [*Pml*1–2: [*Pml*: JN885472<sup>d</sup>], JN885473<sup>d</sup>].

OTU7 (*P. ochraventer*, *Poc*). *Cytb* gene ( $n = 2$  [ $n = 1$ ]). San Luis Potosí: Las Abritas, *Poc*1–2: [*Poc*, JX910119<sup>b</sup>], DQ973106<sup>a</sup>. NADH genes ( $n = 1$ ). San Luis Potosí: Santa Isabel, [*Poc*: ---<sup>c</sup>]

Congeneric species of a close species group (close Outgroup)

OTU8 (*P. mexicanus totontepecus*, *Pmt*). *Cytb* gene ( $n = 4$  [ $n = 2$ ]). Veracruz: Zongolica; Misantla, *Pmt*1–4: [*Pmt*1–2, *Pmt*: AY376425<sup>a</sup>, EF028174<sup>a</sup>], TTU82759<sup>c</sup>, CNMA34309<sup>c</sup>. NADH genes ( $n = 2$  [ $n = 1$ ]). No data, *Pmt*1–2: [*Pmt*: U83862<sup>f</sup>], JN885471<sup>f</sup>.

Non-congeneric species (farther Outgroups)

OTU9 (*Megadontomys cryophilus*, *Mcr*). *Cyt-b* gene ( $n = 2$  [ $n = 1$ ]). Oaxaca: Puerto de la Soledad, *Mcr*1–2: [*Mcr*: DQ861373<sup>a</sup>], BYU16076<sup>a</sup>. NADH genes ( $n = 1$ ). Oaxaca: Puerto de la Soledad, [*Mcr*, DQ793119<sup>d</sup>].

OTU10 (*Osgoodomys banderanus*, *Oba*) – root. *Cytb* gene ( $n = 3$  [ $n = 1$ ]). Michoacán: Coalcomán, *Oba*1–2, *Oba*3: [*Oba*: DQ000473<sup>a</sup>], TK45952<sup>a</sup>, EF98985<sup>a</sup>. NADH genes ( $n = 1$ ). No data, *Oba*1: [*Oba*, U83860<sup>d</sup>].

*Geographic representation and sample sizes, according to scientific collection, in the morphometric analyses.* Due to the restrictions imposed by the COVID-19 pandemic, the catalog numbers of the specimens examined are not shown in this list and only the number of specimens in each locality is referred to. Authors can send catalog numbers later upon request, when the respective scientific collections can be accessed. This list follows the same overall arrangement as the previous list for sequences, beginning with OTUs (with the genetic group, GG1–5 in the Ingroup), taxonomic or geographic designation, and key inside parenthesis. The geographic data, organized in a NW-SE direction, begin with the GLs' number and name (GL1–18; see Figure 1b), with sample sizes inside parenthesis, followed by the State and specific localities (SLs) with the number of examined specimens and collection acronyms inside parenthesis. Numbers for GLs and SLs are from Cruz Gómez *et al.* (2018). SLs with an asterisk (\*) refer to closer records to the type localities of GG1–3, or to the first records in Oaxaca for GG4–5. The specimens are hosted at Universidad Autónoma Metropolitana, Iztapalapa Unit (UAMI), and at the Museum of Zoology "Alfonso L. Herrera", Facultad de Ciencias, Universidad Nacional Autónoma de México (MZFC-M). Total sample sizes, according to analyses, and character coding in the list (Brackets, italics) is as follows:

a) A total of 233 skull in SLs were examined in the traditional morphometric analyses for overall skull size, using 18 linear measurements; their sample sizes follow the geographic data inside parenthesis.

b) From the former, a total of 217 skulls in perfect condition (complete, not damaged, and clean), were also used in morphogeometric analyzes, using five morphogeometric configurations of ventral shape of skull; their sample sizes are mentioned inside brackets. If the same skulls were used in both morphometric analyses, the sample sizes appears only once after the SLs, inside brackets.

c) Finally, a selected set of 24 skulls were sequenced for all mitochondrial genes. The obtained sequences were used in the integrated phylogeny with all kind of characters; their sample sizes appear in italics inside brackets. These specimens are linked with the previous list by its GLs' code (abbreviation of GL name and number of sequences, 1– $n$ , where the  $n$  is total sample size).

OTUs currently assigned to *Peromyscus furvus* (Ingroup):

OTU1 (GG1, *latirostris*, *Pl*,  $n = 36$  [ $n =$ ; *Xili*1–8]). GL1 Xilitla\* ( $n = 20$  [ $n = 16$ ]). San Luis Potosí: 8) Apetzco, 0.5 Km N, 2 Km W Xilitla, San Luis Potosí\* ( $n = 9$  [ $n = 3$ ] UAMI). 16) Km 241 carretera Xilitla, El Rizal ( $n = 7$  [ $n = 3$ ] MZFC); 19) 11 Km S, 8 Km W Xilitla ( $n = 4$  [ $n = 2$ ] UAMI). GL2 Santa Inés [ $n = 16$ ]. Querétaro [ $n = 14$ ]: 15) El Pemache [ $n = 4$  MZF]; 21) 2.8 Km 324 NW Santa Inés [ $n = 3$  MZFC]; 22) 2.5 Km NW Santa Inés [ $n = 5$  MZF]; 23) 2 Km W Santa Inés [ $n = 2$  MZFC]. Hidalgo [ $n = 2$ ]: 20) 13.5 Km SE Pisaflores [ $n = 1$  MZFC]; 25) 3 km S Santa Ana de Allende [1 UAMI].

OTU2 (GG2, *angustirostris*, *Pa*,  $n = 100$  [ $n = 96$ ,  $n = 14$ ]). GL3 Otongo ( $n = 20$  [ $n = 19$ ]): Hidalgo: 29) 1.5 Km N Chilijapa [ $n = 5$  UAMI]; 33) 4 km N Tepehuacán de Guerrero [ $n = 1$  UAMI]; 34) 1 Km N Chilijapa, ( $n = 1$  UAMI); 46) 1 Km S, 3.5 Km W Otongo [ $n = 2$  UAMI]. 48) Km S, 6 Km W Otongo [ $n = 3$  UAMI]. GL4 Tlachinol ( $n = 18$  [ $n = 17$ ; *Tlan*1–5]). Hidalgo: 28) 5 Km N, 1.5 km E Tlachinol ( $n = 1$  UAMI); 31) 4 Km N, 2 km E Tlachinol [ $n = 5$  UAMI]; 32) 4 Km N, 1.5 Km E Tlachinol [ $n = 1$  UAMI]; 35) 3 Km N 1 Km E Tlachinol [ $n = 3$  UAMI]; 36) 2.5 Km N, 1.5 O Tlachinol [ $n = 3$  UAMI]; 43) 10 km N Carr. Tehuacan-Huazalingo [ $n = 4$  MZFC]; 44) 10 Km NW Tehuacan, Car. Tehuacan-Huazalingo [ $n = 1$  MZFC]; 45) 1.5 Km S, 3.8 Km W Tlachinol [ $n = 4$ ,  $n = 2$  UAMI]; 47) 2 Km S, 3 Km W Tlachinol [ $n = 6$ ,  $n = 5$  UAMI]. GL5 Molango [ $n = 1$ ]. Hidalgo: 52) Tianguistengo [ $n = 1$  UAMI]. GL6 Zacualpan\* [ $n = 10$ , *Zacu*1–3]. Veracruz: 61) 9 Km W Zacualpan [ $n = 6$  MZFC]; 62) 1 Km E Zacualpan\* [ $n = 4$ ,  $n = 3$  MZFC]. GL7 Tenango de Doria ( $n = 34$  [ $n = 33$ , *TeDo*1–4]). Hidalgo: 69) San Bartolo, Tutotepec [ $n = 1$  MZFC]; 70) San Bartolo, Cueva El Cirio [ $n = 6$  MZFC]; 76) El Texmé [ $n = 2$  MZFC]; 78) Tenango de Doria ( $n = 20$  [ $n = 19$ ,  $n = 4$  MZFC]; 80) Tenango de Doria, el Potrero [ $n = 5$  MZFC]. GL8. Xicotepec de Juárez [ $n = 5$ , 5]. Puebla: 77) Xicotepec de Juárez, El Salto [ $n = 5$  MZFC]. GL9. Huauchinango [ $n = 6$ ]. Puebla: 82) 8 Km N Huauchinango [ $n = 1$  UAMI]; 84) 0.2 Km N Honey [ $n = 2$  UAMI]; 93) 5.6 KM SW Huauchinango [ $n = 4$  MZFC]. GL10 Zacapoaxtla [ $n = 6$ ]. Puebla: 95) 5.5 Km N Zacapoaxtla [ $n = 4$  UAMI]; 96) 5 Km N Zacapoaxtla [ $n = 2$  UAMI].

OTU3 (GG3, *furvus*, *Pf*,  $n = 47$  [ $n = 45$ ]). GL11. Las Minas ( $n = 0$ ). Veracruz (with no morphometric specimens in either UAMI or MZFC). GL12 Naolinco ( $n = 22$  [ $n = 21$ ]). Veracruz: 103) 4 Km N Naolinco [ $n = 2$  UAMI]; 104) 1 Km W Tlacolulan [ $n = 6$  UAMI];

106) 1 Km S Tlacolulan [ $n = 5$  UAMI]; 112) 8 Km SW Naolinco [ $n = 9$  UAMI]. GL13 Xalapa\* ( $n = 4$  [ $n = 3$ ])\*. Veracruz: 116. 1.5 Km SE *Banderillas*\* [ $n = 4$  UAMI]. GL14 Ixhuacán de los Reyes [ $n = 12$ ]. Veracruz: 129) 2 Km NO Ixhuacán de los Reyes ( $n = 5$  UAMI); 130) 1 Km W Ixhuacán de los Reyes [ $n = 7$  UAMI]. GL15 Coscomatepec [ $n = 9$ ]. Veracruz: 132. 1.5 Km SE Quimixtlán [ $n = 5$  UAMI]; 138) 5.5 km N, 6 km E Coscomatepec [ $n = 4$  UAMI].

OTU4 (GG4, undescribed *Peromyscus*, *Oax18*, [ $n = 25$ , *PtS1-5*]). GL18 Puerto de la Soledad [ $n = 25$ ]. Oaxaca\*: 148) 5 Km N, 1 Km W Huautla [ $n = 1$  UAMI]; 152) 3 km N, 1 Km Huautla [ $n = 1$  UAMI]; 151) Teotitlán, Puerto de la Soledad, Huehuetlán [ $n = 22$ ,  $n = 5$  MZFC]; 166) San Juan Bautista, Cuicatlán. El Venado [ $n = 1$  MZFC].

OTU5 (GG5, undescribed *Peromyscus*, *Oax19*, [ $n = 19$ ]). GL19. La Esperanza, Oaxaca\* [ $n = 19$ ]; 169) 5 Km S, 3 Km W La Esperanza [ $n = 9$  UAMI]; 170) 2.5 Km N, 1 Km E La Esperanza [ $n = 10$  UAMI].

OTUs in the Outgroups:

a. Congeneric species formerly assigned to the *furus* species group

*Peromyscus ochraventer*, *Poc* [ $n = 26$ ]. Tamaulipas: 174) 8 KM NW Gómez Farías [ $n = 26$  UAMI].

*Peromyscus melanocarpus*, *Pml* [ $n = 22$ ]. Oaxaca: 182) 3.5 Km Santa María Pápal, 2500 m, [ $n = 22$  UAMI].

b. Congeneric species of a close species group:

*Peromyscus mexicanus totontepecus*, *Pmt* [ $n = 36$ ]. Oaxaca: 177) 8 Km NW Huatla, 1150 m, [ $n = 6$  UAMI]; 178) 3 km N, 1 Km W Huautla, 1140 m [ $n = 16$  UAMI]; 180) Huatla, 1130 m [ $n = 6$  UAMI]; 183) 5 Km N, 1 Km W Huautla, 1120 m [ $n = 8$  UAMI].

c. Non-congeneric species

*Megadontomys criophylus*, *Mcr* [ $n = 14$ ]. Oaxaca: 176) 5 Km S, 3 Km W La Esperanza, 1950 m [ $n = 4$  UAMI]; 184) 2.5 Km N, 1 Km E La Esperanza, 1850 [ $n = 9$  UAMI].

*Osgoodomys banderanus*, *Oba* [ $n = 18$ ]. Michoacán: 175) Arteaga "Charco del Toro" [ $n = 8$  MZFC]; 179) Lázaro Cárdenas «El Habillo» [ $n = 5$  MZFC]; 181) Lázaro Cárdenas «La Bonetera» [ $n = 5$  MZFC].