



## **Thermoregulation and reproductive responses of rams under heat stress.**

### **Review**



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**Abstract:**

The high temperatures recorded during the summer season in hot regions compromise the reproductive capacity of domestic animals. In rams, heat stress (HS) causes in the body a series of physiological, metabolic, endocrine and molecular adjustments in order to maintain normothermia and survive; however, several of these changes are negatively associated with their fertility, mainly endocrine ones. HS in rams causes a decrease in blood testosterone concentrations through different mechanisms, and this is negatively reflected on the process of spermatogenesis and sexual behavior. Consequently, heat-stressed rams exhibit low seminal quality and libido; at the sperm level, structural and DNA damage has been observed. Given this situation, the use of HS mitigation strategies during the summer in sheep farms in hot regions is recommended, such as the use of shades in pens, administration of antioxidants or modifications in the diet. Therefore, the objective of this document is to review the current knowledge regarding the effect of HS on the thermoregulation and reproductive capacity of rams, as well as the application of strategies for its mitigation.

**Key words:** Ram, Male sheep, Libido, Seminal quality, Sperm damage.

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## Introduction

Regions with hot climates are characterized by high ambient temperatures ( $T_a$ ) and relative humidity (RH) in summer, which generally exceeds the upper limit of the thermoneutral zone for production animals ( $\leq 30$  °C), causing them the presence of environmental conditions of heat stress (HS)<sup>(1,2)</sup>. The productive and reproductive impact generated by HS in animals varies among species, with small ruminants showing the best adaptation to these environmental conditions<sup>(3)</sup>. Some reviews have described the thermoregulation mechanisms used by sheep to avoid hyperthermia under HS<sup>(1,2,4,5)</sup>, but little attention is paid to the effect it has on ram reproduction. In hot climates, the reproductive success of the flock depends largely on the adaptation and proper reproductive functioning of the rams.

The organism of heat-stressed rams presents a series of changes to avoid hyperthermia<sup>(1,6-8)</sup>. Thus, the reproductive capacity of rams decreases while making physiological, metabolic and endocrinological efforts to stay in normothermia<sup>(3,7,9-11)</sup>. HS can negatively affect the

reproduction of the ram by different mechanisms, the main ones being: 1) decrease in testosterone concentrations, and 2) direct damage in the morphometry and content of genetic material of the sperm<sup>(12,13)</sup>. This is reflected in failures in the process of spermatogenesis, as well as in low seminal quality, reproductive behavior and fertility<sup>(7,14-16)</sup>. However, the implementation of HS mitigation strategies improves the reproductive capacity of the ram in these climatic conditions<sup>(1,8,17)</sup>.

It is worth mentioning that the results of the effects of HS on ram reproduction are not consistent among studies. Differences between breeds in the level of adaptation to HS largely explain these discrepancies<sup>(3)</sup>. Therefore, this review aims to describe the current knowledge that exists in relation to the effect of HS on the thermoregulation and reproductive capacity of rams, as well as the application of strategies for its mitigation.

## **Sheep in hot climates**

In recent decades, the excessive accumulation of greenhouse gases (GHG) in the atmosphere is causing an increase in the Ta of the earth's surface<sup>(1)</sup>, so climate change worldwide is eminent, mainly with tendencies to promote a greater presence of hot climates and, consequently, the desertification of more regions of the terrestrial globe<sup>(11)</sup>. Sheep exposed to high environmental Ta, as well as any other production animal, experience HS<sup>(2)</sup>, which represents a physiological-metabolic challenge for the organism to stay in conditions of homeothermy<sup>(6)</sup>.

In the search for strategies that help to maintain the production of food of animal origin under this adverse climate scenario, some authors propose sheep production as an alternative<sup>(1,10,18)</sup>, mainly because they are able to maintain their productive performance under HS conditions<sup>(3)</sup>. Adaptability characteristics possessed by sheep include resistance to parasites, diseases and scarcity of drinking water<sup>(15,19)</sup>; also ability to take advantage of poor quality agricultural fodder and wastes, and maintenance of the reproductive capacity of the flock and lamb growth under HS scenarios<sup>(1,3,5)</sup>. It is worth mentioning that the adaptation level of sheep varies widely among breeds, since there is a great diversity of them developed from cold to hot climatic conditions.

## Heat stress and sheep production

Stress is generated by the presence of an external event causing alterations in a biological system<sup>(20)</sup>. In production animals, there is stress when some external factor alters their health, basal metabolism and productive capacity<sup>(3)</sup>. In this sense, sheep can develop symptoms of stress from facing drastic changes in climatic conditions, and in fact, they develop HS when the combination of environmental factors cause an increase in the  $T_a$  above the upper limit of their thermoneutral zone<sup>(1)</sup>.

Climatic variables that can promote the HS environment are  $T_a$ , RH, solar radiation, wind speed and precipitation; however,  $T_a$  and RH are the main factors associated with presence of HS<sup>(5)</sup>, and consequently, both are used to construct the temperature-humidity index ( $THI = T_a - [(0.31 - 0.31 * RH)(T_a - 14.4)]$ )<sup>(4)</sup>. It should be clarified that this index was not developed for sheep, however, it is currently widely used to define the degree of HS in this species, since to date there is no specific one for them. Based on that THI, sheep are considered to begin experiencing HS at 22.2 units, being of moderate type between 22.2 and <23.3 units, severe between 23.3 and <25.6 units, and extreme severe at  $THI \geq 25.6$  units<sup>(4)</sup>.

The thermoneutral zone for most sheep breeds is between 5 and 25 °C<sup>(1)</sup>, however, there are adapted breeds that begin to experience HS above 30 °C<sup>(5,15)</sup>. This suggests that, despite being homeotherms, sheep's tolerance to HS varies widely among breeds, and specific studies for each breed should be conducted to assess their tolerance to high  $T_a$ . In the world, there are more than 1,000 sheep breeds, which vary in their ability to thermoregulate in hyperthermia environments, and this is due to their climatic origin<sup>(3)</sup>. In Mexico, there are both wool and hair breeds, but the latter are more tolerant to HS, since they originated in hot climates, while wool breeds originated in cold or temperate climates<sup>(5)</sup>. This does not mean that there are no HS-tolerant wool breeds in other countries; in Australia, the Merino breed shows great adaptability to warm regions<sup>(1)</sup>.

The thermoregulatory response of sheep to HS also varies with sex, and within sex with age, physiological state and reproductive activity<sup>(5,21)</sup>. While negative effects of HS are more noticeable in offspring and pregnant and lactating ewes<sup>(21,22)</sup>, rams seem to be less noticeable since their metabolic heat production is lower compared to ewes, and even more so when they are in reproductive rest<sup>(1)</sup>. The latter could be the cause that most studies are developed in ewes and consider that the topic is of little relevance for research in rams. However, testicular reproductive processes are very sensitive to changes in  $T_a$ , which is associated with low fertility in rams during the hot season. In this sense, the rest of the literature review will focus on analyzing the effects of HS on ram reproduction.

## Heat stress and thermoregulation of the ram

The thermoregulation of rams under thermoneutral conditions is essentially due to the activation of non-evaporative mechanisms, without this implying metabolic, endocrine or maintenance energy alterations<sup>(1,3)</sup>. However, under HS conditions, rams activate a series of thermoregulation mechanisms that favor homeothermy in the face of thermal challenge.

The HS in warm regions increases the average values of physiological variables, such as rectal temperature (RT), respiratory rate (RR), heart rate, and sweat rate (Table 1)<sup>(10,18,23)</sup>. Thus, rams maintain their normothermia, although it is important to note that the increase in the number of breaths is the main mechanism used by rams to lose the body heat load<sup>(21)</sup>. In fact, sheep under HS can eliminate between 60 and 90 % of the thermal load through the respiratory system<sup>(4)</sup>. Another activated physiological mechanism, which is more evident in hair-breed rams subject to HS, is the redistribution of blood flow to peripheral tissues to dissipate body heat by radiation through the skin<sup>(5,15)</sup>. As the temperature gradient between the skin and the environment decreases, RR increases until it becomes the main route of body heat dissipation<sup>(24)</sup>.

**Table 1:** Changes in the physiological variables of heat-stressed rams

Source	Breed	Treatment season	Air temperature (°C)	Findings during the season/treatment with higher temperature
(52)	Suffolk	Winter	14.5	↑ RT, ST
		Summer	28.2	
(45)	Najdi	Winter	19.8 ± 0.4	↑ RT, RR and HR
		Summer	38.4 ± 0.3	
(72)	Malpura	Winter	7.0 - 25.5	↑ RR and HR
		Summer	23.0 - 40.0	
(15)	Santa Inés	Winter	~ 12.5 - 28.0	↑ RT, HR and SR
		Summer	~ 18.0 - 32.0	
	Morada Nova	Winter	~ 12.5 - 28.0	↑ RT, HR and SR
		Summer	~ 18.0 - 32.0	
(73)	Santa Inés	Winter	~ 12.5 - 27.0	No changes
		Summer	~ 19.0 - 31.0	
	Morada Nova	Winter	~ 12.5 - 27.0	↓ RR and ↑ TMT
		Summer	~ 19.0 - 31.0	
	Texel	Winter	~ 12.5 - 27.0	No changes
		Summer	~ 19.0 - 31.0	
	Dorper	Winter	~ 12.5 - 27.0	↓ RR and ↑ TMT
		Summer	~ 19.0 - 31.0	

(18)	Small-tailed	Thermoneutral	~ 22.0 - 23.0	↑ RR
	Han	Heat stress	~ 30.0 - 35.0	
(74)	Polish	Thermoneutral	16.5 ± 1.0	↑ RT and RR
	Merino	Heat stress	50.0 ± 1.0	
(10)	Merino	Thermoneutral	20.1 - 20.9	↑ RR and HR
		Heat stress	28.6 - 30.6	
(24)	Malpura	x	Thermoneutral	33.6 ± 0.7
	Malpura	x	Heat stress	44.2 ± 0.2
	Garole			

RT= rectal temperature, RR= respiratory rate, HR= heart rate, ST= scrotal temperature, TMT= testicular mean temperature; SR= sweat rate.

In rams, the scrotum functions as a thermoregulation organ under both thermoneutral and HS conditions<sup>(1)</sup>. In hot summer environmental conditions, the scrotum is one of the body regions that most dissipates heat load due to the large vascularization (pampiniform plexus) on the testicular surface, and the large number of sweat glands<sup>(15,16)</sup>. There is a high correlation between body core and scrotal temperatures, so knowing the variability of scrotal temperature allows evaluating the thermoregulation efficiency in rams<sup>(15)</sup>.

The activation of evaporative mechanisms demands a large amount of body water, so water intake can increase between 19 and 25% in rams during the summer<sup>(25)</sup>. In consequence, feed intake is decreased by a substitution effect<sup>(26)</sup>. However, in hair sheep, it was shown that feed intake remained similar in summer and spring, regardless of the increase in water intake recorded during summer<sup>(5)</sup>. This suggests that the substitution effect of water intake for feed intake occurs mainly in rams with less tolerance to HS. Thus, the reduction in feed consumption is the result of the ram's effort to reduce endogenous heat production, by partially suppressing metabolic and rumen activity<sup>(1,4,27)</sup>.

All the physiological adjustments presented by the rams as a result of HS cause an increase in the maintenance energy requirements, while the reduction in the feed intake alters its availability<sup>(5)</sup>. Consequently, high summer Ta alter the metabolism of rams, firstly to distribute energy to thermoregulation processes, and secondly to reduce endogenous heat production, while making the use of energy substrates more efficient<sup>(28,29)</sup>. However, the results of the effect of HS on serum concentrations of metabolites and metabolic hormones are not consistent among studies (Table 2).

**Table 2:** Changes in blood metabolites of heat-stressed rams

Source	Breed	Treatment season	Air temperature (°C)	Findings during the season/treatment with higher temperature
(44)	Ossimi	Winter	24.1	↑ GLU
		Summer	33.7	↓ CHOL and LIPT
(45)	Najdi	Winter	19.8 ± 0.4	↑ GLU and PROT
		Summer	38.3 ± 0.3	
(24)	Malpura	x Thermoneutral	33.6 ± 0.7	↓ PROT and T <sub>3</sub> ↑ COR
	Malpura Garole	x Heat stress	44.2 ± 0.2	
(10)	Merino	Thermoneutral	20.1 - 20.9	↑ COR
		Heat stress	28.6 - 30.6	
(25)	Fat-tailed Iranian	Thermoneutral	21.0	↓ GLU, TRIG, T <sub>3</sub> and T <sub>4</sub> ↑ PROT and COR
		Heat stress	40.0	
(74)	Polish Merino	Thermoneutral	16.5 ± 1.0	↓ GLU ↑ COR
		Heat stress	50.0 ± 1.0	
(18)	Small-tailed Han	Thermoneutral	~ 22.0 - 23.0	↓ TRIG, PROT
		Heat stress	~ 30.0 - 35.0	

GLU= glucose, CHOL= cholesterol, TRIG= triglycerides, PROT= total protein, LIPT= total lipids, COR= cortisol, T<sub>3</sub>= triiodothyronine, T<sub>4</sub>=thyroxine.

The high RR observed in heat-stressed rams demands an excessive amount of glucose as an energy source for the functioning of the muscles of the respiratory system<sup>(3)</sup>. Consequently, the rams in summer increase blood glucose concentrations compared to thermoneutral seasons, which is because cortisol concentrations also increase in response to HS<sup>(1,3)</sup>. Cortisol promotes gluconeogenesis and hepatic glycolysis<sup>(28)</sup>. According to this, Ossimi<sup>(30)</sup> and Najdi<sup>(31)</sup> breed rams registered higher blood cortisol and glucose concentrations in summer than in winter. Nevertheless, there are studies where serum glucose concentrations decreased<sup>(18,32)</sup> or did not change<sup>(10,23)</sup> due to HS in rams. This could be associated with an increase in plasma insulin concentrations<sup>(28)</sup>.

In sheep exposed to chronic HS conditions, mainly those of breeds adapted to hot climates, blood insulin concentrations increase as an adaptive mechanism to maintain proper metabolic functioning, improve energy use efficiency and reduce fatty tissue catabolism<sup>(28,29)</sup>.

Particularly, high insulin levels allow heat-stressed rams to: 1) avoid apoptosis of pancreatic  $\beta$  cells by increasing the production of non-esterified fatty acids; 2) promote the circulating glucose cellular uptake for its metabolism; and 3) maintain anabolism and prevent catabolism, mainly of fatty tissue<sup>(3,28)</sup>. This last point has been associated with lower serum concentrations of triglycerides, cholesterol and total lipids in rams subjected to chronic HS<sup>(18,30)</sup>. Additionally, a reduction in blood concentrations of these lipid metabolites is partially associated with the mobilization of fatty acids to meet energy requirements when the glucose-saving system is activated<sup>(1,28)</sup>. Macías-Cruz *et al*<sup>(33)</sup> mention that, in sheep, serum concentrations of glucose, cholesterol, triglycerides, total protein and urea vary according to the type of HS. Chronic HS reduces serum metabolite concentrations associated with energy metabolism (i.e., glucose, cholesterol, and triglyceride), but increases metabolite concentrations associated with protein metabolism (i.e., total protein and urea). In the case of acute HS, variations in blood concentrations of these metabolites show an effect contrary to that observed in chronic HS, which is due to the fact that energy metabolism changes to ensure greater availability of energy substrates when making physiological adjustments<sup>(3)</sup>.

Finally, the thyroid gland also plays an important role in the thermoregulation of all species, including rams<sup>(13)</sup>. The HS causes a reduction in the release of thyroid hormones, which favors lower metabolic heat production and body heat load<sup>(23,32)</sup>. Notoriously, triiodothyronine has a shorter half-life and is more thermo-sensitive than thyroxine, as demonstrated in a study of Malpura breed rams<sup>(23)</sup>.

## Heat stress and reproductive endocrinology of the ram

Environmental factors play an important role in controlling the reproductive capacity of rams. An inadequate environment can cause stress to the ram and this triggers alterations in the neuroendocrine function of the reproductive axis<sup>(34)</sup>. In hot regions, high summer  $T_{as}$  generate a HS environment for rams, leading them to prioritize activities associated with thermoregulation processes rather than reproductive functions<sup>(13)</sup>. In fact, their reproductive capacity could be totally inhibited in breeds susceptible to HS, while such inhibition could be partial or non-existent in adapted breeds<sup>(1,15,23)</sup>.

Rams, in response to HS conditions, activate the sympatho-adrenal-medullary (SAM) system and the hypothalamic-pituitary-adrenal (HPA) axis<sup>(12)</sup>. The SAM system stimulates the release of catecholamines (adrenaline and noradrenaline) in the medulla of the adrenal glands<sup>(9)</sup>, which induce peripheral vasodilation and increase energy availability through gluconeogenesis and lipolysis<sup>(1,13)</sup>. For its part, the HPA axis begins its activation with the hypothalamic secretion of corticotropin-releasing hormones (CRH), which in turn stimulate

the secretion of adrenocorticotrophic hormone (ACTH) in the adenohipophysis<sup>(12,13,17)</sup>. Additionally, catecholamines together with CRH cause the hypothalamic release of  $\beta$ -endorphin, whose precursor is the proopiomelanocortin polypeptide, which is also a precursor of ACTH<sup>(17,34)</sup>. ACTH via endocrine stimulates the synthesis of glucocorticoids (cortisol and corticosterone) and mineralocorticoid (aldosterone) in the adrenal cortex from cholesterol<sup>(13,17,32)</sup>. The release of cortisol is the main mechanism through which the HHA axis inhibits the functioning of the hypothalamic-pituitary-gonadal (HHG) axis<sup>(9,11)</sup>, and consequently, the degree of reproductive activity in rams exposed to HS<sup>(34)</sup>.

The activity levels of HHA and HHG axes are negatively related, in such a way that lower testosterone concentrations and, consequently, reproductive activity are commonly observed in heat-stressed rams<sup>(1)</sup>. The increase in cortisol in the blood causes the levels of testosterone available in the seminiferous tubules to decrease, which in turn reduces sperm production and quality due to low activity in the process of spermatogenesis<sup>(35,36)</sup>. Also, libido and mounting ability is reduced due to low testosterone concentrations<sup>(37-39)</sup>.

Testosterone is synthesized and released by Leydig testicular cells, which respond to the stimulation of luteinizing hormone (LH) for such action<sup>(9,40)</sup>. Sertoli cells, in response to follicle-stimulating hormone (FSH) stimuli, synthesize and release the androgen-binding protein, which is responsible for binding with circulating testosterone to introduce it to the seminiferous tubules<sup>(40)</sup>. Once inside the seminiferous tubules, testosterone is responsible for synchronizing the entire process of spermatogenesis<sup>(12)</sup>. However, the activation of the HHA axis in response to HS may negatively compromise the correct functioning of this mechanism at different points. It has been widely documented that cortisol generates negative feedback on GnRH at the hypothalamus level<sup>(1,34)</sup>, a situation that in turn prevents the adenohipophysis from synthesizing and releasing gonadotropin hormones (FSH and LH)<sup>(12,13)</sup>; both essential to ensure the presence of sufficient testosterone concentrations within the seminiferous tubules, to carry out spermatogenesis. Some studies also indicate that testosterone concentrations may decrease by different mechanisms than those associated with the functioning of the hypothalamus and hypophysis in heat-stressed rams<sup>(9,12,41)</sup>.

Testosterone concentrations may decrease because glucocorticoids reduce the expression of receptors for LH into Leydig cells<sup>(41-43)</sup>. It has also been reported that Leydig cells require certain cytokinins such as IL-1 and IL-6 for the testosterone release, however, an increase in glucocorticoid synthesis showed to decrease the immune response and, therefore, the production of these cytokinins<sup>(12,44)</sup>. Other evidence indicates that the production of androgen-binding protein in Sertoli cells may decrease due to a low production of thyroid hormones<sup>(22,45)</sup>. Similarly, germ cell damages and low expression of the protein Connexin-43 (responsible for the union among Sertoli cells) are attributed to the direct effect of testicular hyperthermia<sup>(46)</sup>. These alterations at the level of Sertoli cells could lead to a low availability

of testosterone within the seminiferous tubules<sup>(12)</sup>. Note that some of these studies were not done in rams, so they may be the reason for future lines of research.

## Heat stress and reproductive capacity of the ram

### Effects on seminal quality

Seminal quality in rams decreases under HS conditions due to the activation of neuroendocrine, physiological and metabolic mechanisms, as well as the increase in maintenance energy expenditure to preserve normothermia conditions<sup>(1)</sup>. Generally, the damage caused to the sperm by HS becomes visible between 14 and 21 d after the start of exposure of the rams to high Tas<sup>(47)</sup>, therefore, a decrease in seminal quality is detected until then.

The most affected seminal characteristics are progressive motility, sperm abnormalities, plasma membrane integrity, sperm concentration, and ejaculate volume (Table 3)<sup>(1,48)</sup>. Progressive and mass motility decrease between 5 and 25 %<sup>(49,50)</sup>, which is associated with an increase in the percentage of abnormal sperm<sup>(16)</sup>. The sperm abnormalities predominating due to HS are head and acrosomal defects<sup>(51)</sup>. It is worth mentioning that these abnormalities are less frequent in native breed rams of warm regions, in such a way that these breeds adapted to HS present between 1 and 5 % of abnormal sperm<sup>(49,52)</sup>.

**Table 3:** Changes in semen characteristics of heat-stressed rams

Source	Breed	Treatment season	Air temperature (°C)	Findings during the season/treatment with higher temperature
(49)	Chios	Autumn	9.7 - 18.3	↓ MOT and CON
		Summer	19.1 - 30.6	↑ SA
	Friesian	Autumn	9.7 - 18.3	↓ MOT and CON
		Summer	19.1 - 30.6	↑ MP and SA
(54)	Persian	Winter	5.8 ± 3.8	↓ VOL
	Karakul	Summer	26.0 ± 4.8	↑ VIT and TES
(52)	Suffolk	Winter	14.5	↓ SC, MM, VIT and CON
		Summer	28.2	↑ seminal pH, SA and acrosomal damage

(55)	Hamari (not sheared)	Winter	14.1 - 32.4	↓ VOL, VIT, MM and MP ↑ SA
		Summer	22.9 - 43.3	
(50)	Hamari (sheared)	Winter	14.1 - 32.4	↓ VOL, MM and VIT ↑ SA
		Summer	22.9 - 43.3	
(27)	Dorper	Winter	18.0 - 26.0	↓ SC, CON, MM and MP ↑ SA
		Summer	26.0 - 32.0	
(15)	Zulu	Winter	23.3	↓ VOL, CON and PMI ↑ SC
		Summer	28.3	
(14)	Morada Nova	Winter	~ 12.5 - 28.0	↑ CON and SSA
		Summer	~ 18.0 - 32.0	
(56)	Santa Inés	Winter	~ 12.5 - 28.0	↓ PMI ↑ CON
		Summer	~ 18.0 - 32.0	
(53)	Pelibuey	Winter	--	↓ SC and CON ↑ SA
		Autumn	26.0 - 27.8	
(24)	Ouled Djellal	Spring	--	↓ SC, VIT and TES
		Summer	33.0 - 40.0	
(53)	Malpura	Thermoneutral	--	↓ SC, VOL, MM, CON and TES
		Heat stress	42.0	
(24)	Malpura x Garole	Thermoneutral	33.6 ± 0.7	↓ MOT
		Heat stress	44.2 ± 0.2	

SC= scrotal circumference, MOT= sperm motility, MM= mass motility, PM= progressive motility, CON= sperm concentration, VOL= ejaculate volume, VIT= sperm vitality, PMI= plasma membrane integrity, SA= sperm abnormalities, SSA= secondary sperm abnormalities, TES= serum testosterone.

The scrotal perimeter and sperm concentration have also shown to decrease due to HS<sup>(16)</sup>, which is possibly related to less sperm cell proliferation and greater apoptosis of cells of testicular parenchyma<sup>(47)</sup>. Some studies indicate a decrease of 2 to 7 cm in the scrotal perimeter and 3,000 million sperm per milliliter of ejaculate, after subjecting the rams to HS conditions<sup>(52,53)</sup>.

On the other hand, the secretory activity of the accessory glands decreases in heat-stressed rams, which is directly reflected in lower ejaculate volume<sup>(36,53-55)</sup>. The lower secretion of seminal plasma in the accessory glands is associated with serum testosterone concentrations<sup>(12,56)</sup>. Additionally, the composition of seminal plasma is modified by HS conditions, mainly at the level of electrolyte and protein concentrations, compounds that maintain the seminal pH between neutral and slightly alkaline (7.0 to 7.3)<sup>(11)</sup>. In general, HS

increases the seminal pH of rams<sup>(52,57)</sup>, which reduces the number of sperm per ejaculate and increases the percentage of abnormalities<sup>(36)</sup>.

In summary, elevated environmental  $T_a$ s negatively affects ram fertility, essentially because they decrease sperm production, as well as the seminal plasma quantity and quality. This ends up having a negative impact on the microscopic characteristics of the semen. Note that heat-stressed rams do not immediately regain their optimal fertility when switching to a thermoneutral environment; in fact, they require staying between 9 and 11 wk in this environment to ejaculate a semen of normal quality<sup>(47)</sup>.

### **Effects on sexual behavior**

The sexual behavior of rams has been little evaluated under HS conditions, and the results are contradictory so far. Considering that the service of females is mostly given by natural mounting in the different production systems, it is imperative to elucidate in future research the impact of HS on the mounting capacity of rams.

In Malpura breed rams (adapted to hot climates), the HS induced in thermo-environmental chamber (42 °C) reduced libido and mounting capacity, which was deduced because heat-stressed rams took more time to perform a mount with ejaculation, as well as a higher number of mounting attempts to the first and second ejaculate<sup>(53)</sup>. Likewise, Rembi breed rams exhibited lower libido during the summer season in an arid region<sup>(38)</sup>. The reduction in sexual behavior shown by rams exposed to HS was associated with a lower ability to secrete testosterone. However, there are other studies conducted on purebred<sup>(23)</sup> or crossed<sup>(7)</sup> rams from Malpura genotype, where the effects of HS on sexual behavior were minimal without any difference in serum testosterone concentrations. In hair breed rams used in Mexico, one study reported only an increase in the reaction time of mounting by the effect of the dry and hot season compared to the cool-humid season of a tropical climate<sup>(39)</sup>.

Discrepancies between results could be due to the fact that in those studies where there were no effects<sup>(7,23)</sup>, the differences in  $T_a$  were not so marked. Other important factors to consider are body condition (CC) and reproductive seasonality. Rams with optimal CC (3.0 on a 1-5 scale) have better sexual behavior than rams with low ( $\leq 2$  points) or high ( $\geq 4$  points) CC under HS conditions<sup>(37)</sup>. On the other hand, the summer season represents a transition period between the end of the anoestrus period and the beginning of the natural reproductive period<sup>(58)</sup>. Therefore, rams of breeds with greater sensitivity to reproductive seasonality could present a reduction in sexual behavior during the summer in hot regions, not only because of

high temperatures, but also because of their natural reproductive circannual rhythm. In the case of Mexican hair sheep breeds, which are characterized by low reproductive seasonality but high adaptation to hot climates<sup>(5)</sup>, the expected negative effects of HS on their sexual behavior could be minimal, as demonstrated in tropical conditions<sup>(39)</sup>. However, little research has been done on this topic in hair breed rams and existing studies are still superficial. Hair breeds in Mexico have great relevance for meat production in warm climates, so it is necessary to investigate in depth the impact that HS has on the behavior of these rams.

### **Effects on sperm damage**

Sperm damage due to HS begins to be generated from sperm cells that are in differentiation within the seminiferous tubules until sperm that are in transit in the epididymis. Ram sperm last between 13 and 15 d in epididymal maturation, so they are the first to show damage from hyperthermia<sup>(59)</sup>. Previous studies report that rams exposed to Ta greater than 35 °C can cause 17.5 % of pyriform heads<sup>(60)</sup>, 18.5 % of abnormalities in acrosome<sup>(61)</sup> and about 30 % of tailless sperm<sup>(35)</sup>. Overall, chronic HS (> 60 d) is estimated to cause 43.4 % of minor abnormalities in sperm (e.g., presence of distal cytoplasmic droplet, coiled tip or fully coiled tail, and free normal heads) and 3.6 % of major abnormalities (e.g., proximal cytoplasmic droplet and microcephalic sperm)<sup>(57)</sup>. Another study indicated that head ellipticity appears in the ejaculates of rams from d 42 after testicular hyperthermia, which is associated with direct damage from HS to sperm in the spermiogenesis phase, although the mechanism that leads to this head malformation is unclear<sup>(62)</sup>. It is worth mentioning that the sperm damage generated by HS becomes constant while such environmental conditions remain, and is usually projected for several more weeks after the thermal challenge ends<sup>(1,50)</sup>.

The testicles must remain between 2 and 8 °C below body temperature for proper functioning, otherwise testicular hyperthermia causes damage to testicular somatic and germ cells<sup>(63)</sup>. Spermatocytes and spermatids are considered more susceptible to apoptosis due to the effect of HS because of their high meiotic rate<sup>(1,47)</sup>, although degeneration can also occur in spermatogonia, and Leydig and Sertoli cells<sup>(23,64)</sup>. Chronic HS, but not acute, appears to affect sperm that have already completed their formation and are in the epididymis<sup>(63,64)</sup>. Sperm located in the epididymis increase their level of oxidative stress and decrease their antioxidant capacity in response to continuous and prolonged exposure to HS<sup>(63)</sup>. The latter has been widely demonstrated in mice, so research is required in rams.

On the other hand, blood flow in the testicles of heat-stressed rams is insufficient, which causes testicular hypoxia and, together with direct hyperthermia, it promotes oxidative stress

conditions due to an increase in reactive oxygen species (ROS)<sup>(63)</sup>. Excessive testicular production of ROS leads to peroxidation of sperm membrane phospholipids, triggering direct damage at the level of membrane integrity (20 % degradation) and DNA<sup>(23)</sup>. These damages can decrease the expression of the PH-20 protein in the membrane, which is associated with the activity of binding of the sperm with zona pellucida<sup>(65)</sup>. In addition, they make sperm from rams more susceptible to damage to chromatin conformation<sup>(11)</sup>, and to the presence of DNA fragmentation<sup>(47,65)</sup>. This damage to sperm DNA can cause subfertility or infertility in rams<sup>(11)</sup>, as well as a decrease in sperm resistance when used in artificial insemination and *in vitro* fertilization programs<sup>(51)</sup>.

## Mitigation of heat stress in rams

The use of HS mitigation strategies is a necessity to improve the reproductive capacity of rams in warm climates. There is a wide variety of strategies that can be implemented; however, they are not equally efficient in all climates and production systems. For example, in wool breeds, shearing in the summer months is a widely used strategy to improve thermoregulation capacity in rams, however, in Argentina, it was reported that the incidence of elliptical sperm heads increased (76 %) in Australian Merino rams for shearing them completely in an HS environment<sup>(62)</sup>. Similarly, shearing in Desert Hamari hair breed rams was shown to be effective in improving thermoregulation, but counterproductive to seminal quality during the summer season<sup>(60)</sup>. For his part, Rathore<sup>(61)</sup> found 16 % for sperm abnormalities when testicular wool was sheared in rams. These findings suggest the need for further studies that determine the effectiveness of this HS mitigation strategy in improving ram fertility.

In Morada Nova and Santa Inés rams, thermoregulation capacity and scrotal circumference and testicular firmness improved, in addition, sperm abnormalities decreased below 4% due to the shade installation<sup>(66)</sup>. Similarly, the implementation of asbesto shade improved the maintenance of normothermia in Barki breed rams<sup>(67)</sup>. On the other hand, an increase in airflow under high Ta provided advantages in physiological variables of rams<sup>(10)</sup>. In addition, the use of straw beds in rams' housing pens improved body heat loss by conduction<sup>(1)</sup>; however, the effect of cooling systems or the use of different bedding materials on ram reproductive activity is not known.

Dietary supplementation with proteins, lipids, antioxidants and minerals has been shown to improve the ability of adult sheep and lambs to cope with HS<sup>(1,68,69)</sup>. Nevertheless, in rams, there is only information on the use of antioxidants as a strategy to mitigate the effects of HS. Dietary supplementation of the antioxidant  $\gamma$ -oryzanol in rams decreased ROS production by

26 % and increased the percentage of sperm with intact membrane after testicular hyperthermia; however, there was also an increase in sperm abnormalities<sup>(59)</sup>. Parenteral administration of vitamin E or vitamin E plus selenium improved the seminal quality and libido of Awassi rams subjected to Ta from 43 to 54 °C<sup>(70)</sup>. Research needs to be developed on some nutritional strategies that can help minimize the negative effects of HS on the reproduction of rams.

On the other hand, with the intention of improving the thermoregulation capacity in the offspring, it has been chosen to select progenitors of autochthonous breeds that show thermoresistance capacity and adaptation to the environment in which they have developed<sup>(27)</sup>. In this way, interest in the identification of genetic markers, such as the Booroola fertility gene (FecB), is growing, which apart from increasing the prolificacy of ewes, also positively influences the ability to produce semen of desirable quality under conditions of hot semi-arid climate in purebred Garole rams or crosses with Malpura<sup>(23,71)</sup>.

## Conclusions

Despite the characteristics of resistance and natural rusticity having sheep, HS causes a series of physiological and metabolic changes in the ram that modify energy and reproductive hormonal balance, which finally has a negative impact on blood testosterone concentrations and, therefore, on seminal quality and sexual behavior. In addition, hyperthermia causes direct damage at the level of the membrane and DNA of the sperm, decreasing its fertilizing capacity. Therefore, the use of HS mitigation strategies in rams is a necessity to maintain fertility in the flock, particularly in the hot season of the year of hot climates. The HS mitigation strategy to be used will depend on HS type (acute or chronic) and intensity (moderate or severe) to which the ram is exposed, as well as its degree of adaptation, so it could be used from a simple shaded area with or without fans, to the supplementation of additives such as antioxidants.

### Literature cited:

1. Sejian V, Bhatta R, Gaughan J, Malik PK, Naqvi S, Lal R. Sheep production adapting to climate change. Singapore: Springer Nature Singapore Pte Ltd; 2017.
2. Bernabucci U, Lacetera N, Baumgard LH, Rhoads RP, Ronchi B, Nardone A. Metabolic and hormonal acclimation to heat stress in domesticated ruminants. *Animal* 2010;4(7):1167-1183.

3. Al-Dawood A. Towards heat stress management in small ruminants—a review. *Ann Anim Sci* 2017;17(1):59-88.
4. Marai IFM, El-Darawany AA, Fadiel A, Abdel-Hafez MAM. Physiological traits as affected by heat stress in sheep — A review. *Small Ruminant Res* 2007;71(1):1-12.
5. Vicente-Pérez R, Macías-Cruz U, Avendaño-Reyes L, Correa-Calderón A, López-Baca MA, Lara-Rivera AL. Impacto del estrés por calor en la producción de ovinos de pelo. *Rev Mex Cienc Pecu* 2020;11(1):205-222.
6. Bett B, Kiunga P, Gachohi J, Sindato C, Mbotha D, Robinson T, *et al.* Effects of climate change on the occurrence and distribution of livestock diseases. *Prev Vet Med* 2017;137:119-129.
7. Kumar D, Sejian V, Gaughan JB, Naqvi SMK. Biological functions as affected by summer season-related multiple environmental stressors (heat, nutritional and walking stress) in Malpura rams under semi-arid tropical environment. *Biol Rhythm Res* 2017;48(4):593-606.
8. Belhadj SI, Mohamed C, Najjar T, Ghram A. Meta-analysis of some physiologic, metabolic and oxidative responses of sheep exposed to environmental heat stress. *Livest Sci* 2019;229:179–187.
9. Tort L, Teles M. The endocrine response to stress—a comparative view. In: Akin F editor. *Basic and clinical endocrinology up-to-date*. InTech; 2011:263-286.
10. Wojtas K, Cwynar P, Kołacz R. Effect of thermal stress on physiological and blood parameters in merino sheep. *Bull Vet Inst Pulawy* 2014;58(2):283-288.
11. Rahman MB, Schellander K, Luceno NL, Van Soom A. Heat stress responses in spermatozoa: Mechanisms and consequences for cattle fertility. *Theriogenology* 2018;113:102-112.
12. Damián JP, Bausero M, Bielli A. Acute stress, hypothalamic-hypophyseal-gonadal axis and testicular function—A review. *Ann Anim Sci* 2015;15(1):31-50.
13. Binsiya T, Sejian V, Bagath M, Krishnan G, Hyder I, Manimaran A, *et al.* Significance of hypothalamic-pituitary-adrenal axis to adapt to climate change in livestock. *Int Res J Agri Food Sci* 2017;2(1):1-20.
14. Aké-López J, Aké-Villanueva N, Segura-Correa J, Aké-Villanueva J, Montes-Pérez R. Effect of age and season on semen traits and serving capacity of Pelibuey rams under tropical conditions. *Livest Res Rural Dev* 2016;28(9):166. <http://www.lrrd.org/lrrd28/9/akel28166.htm>. Accessed Nov 25, 2019.

15. Kahwage PR, Esteves SN, Jacinto MAC, Junior WB, Machado R, Romanello N, *et al.* Assessment of body and scrotal thermoregulation and semen quality of hair sheep rams throughout the year in a tropical environment. *Small Ruminant Res* 2018;160:72-80.
16. Moura ABB, Brandao FZ, Esteves SN, de Souza GN, da Fonseca JF, Pantoja MHA, *et al.* Differences in the thermal sensitivity and seminal quality of distinct ovine genotypes raised in tropical conditions. *Theriogenology* 2019;123:123-131.
17. Inbaraj S, Sejian V, Bagath M, Bhatta R. Impact of heat stress on immune responses of livestock: a review. *Pertanika J Trop Agric Sci* 2016;39(4):459-482.
18. Li FK, Yang Y, Jenna K, Xia CH, Lv SJ, Wei WH. Effect of heat stress on the behavioral and physiological patterns of Small-tail Han sheep housed indoors. *Trop Anim Health Prod* 2018;50(8):1893-1901.
19. Leite PG, Marques JI, Furtado DA, Lopes Neto JP, de Souza BB, do Nascimento JWB. Ethology, physiological, and ingestive responses of sheep subjected to different temperatures and salinity levels of water. *Int J Biometeorol* 2019;63(8):1091-1098.
20. Collier RJ, Baumgard LH, Zimbelman RB, Xiao Y. Heat-stress: physiology of acclimation and adaptation. *Anim Frontier* 2019;9(1):12-19.
21. Macías-Cruz U, Correa-Calderón A, Mellado M, Meza-Herrera CA, Aréchiga CF, Avendaño-Reyes L. Thermoregulatory response to outdoor heat stress of hair sheep females at different physiological state. *Int J Biometeorol* 2018;62(12):2151-2160.
22. Macías-Cruz U, Álvarez-Valenzuela FD, Correa-Calderón A, Díaz-Molina R, Mellado M, Meza-Herrera CA, *et al.* Thermoregulation of nutrient-restricted hair ewes subjected to heat stress during late pregnancy. *J Therm Biol* 2013;38(1):1-9.
23. De K, Kumar D, Balaganur K, Kumar Saxena V, Thirumurugan P, Khursheed Naqvi SM. Effect of thermal exposure on physiological adaptability and seminal attributes of rams under semi-arid environment. *J Therm Biol* 2017;65:113-118.
24. Fonsêca VFC, Maia ASC, Saraiva EP, de Melo Costa CC, da Silva RG, Abdoun KA, *et al.* Bio-thermal responses and heat balance of a hair coat sheep breed raised under an equatorial semi-arid environment. *J Therm Biol* 2019;84:83-91.
25. Cain IJW, Krausman PR, Rosenstock S, Turner JC. Mechanisms of thermoregulation and water balance in desert ungulates. *Wildl Soc Bull* 2006;34(3):570-581.
26. NRC. National Research Council. Nutrient requirements of sheep. 6th ed. Washington, DC, USA: National Academy Press; 1985.

27. Berihulay H, Abied A, He X, Jiang L, Ma Y. Adaptation mechanisms of small ruminants to environmental heat stress. *Animals* 2019;9(3):75.
28. Baumgard LH, Rhoads RP, Jr. Effects of heat stress on postabsorptive metabolism and energetics. *Annu Rev Anim Biosci* 2013;1:311-337.
29. Mahjoubi E, Yazdi MH, Aghaziarati N, Noori GR, Afsarian O, Baumgard LH. The effect of cyclical and severe heat stress on growth performance and metabolism in Afshari lambs. *J Anim Sci* 2015;93(4):1632-1640.
30. Khalek TMMA. Thermoregulatory responses of sheep to starvation and heat stress conditions. *Egyptian J Anim Prod* 2007;44(2):137-150.
31. Al-Haidary A, Aljumaah R, Alshaikh M, Abdoun K, Samara E, Okab A, *et al.* Thermoregulatory and physiological responses of Najdi sheep exposed to environmental heat load prevailing in Saudi Arabia. *Pak Vet J* 2012;32(4):515-519.
32. Nazifi S, Saeb M, Rowghani E, Kaveh K. The influences of thermal stress on serum biochemical parameters of Iranian fat-tailed sheep and their correlation with triiodothyronine (T 3), thyroxine (T 4) and cortisol concentrations. *Comp Clin Path* 2003;12(3):135-139.
33. Macías-Cruz U, López-Baca MA, Vicente R, Mejía A, Álvarez FD, Correa-Calderón A, *et al.* Effects of seasonal ambient heat stress (spring vs. summer) on physiological and metabolic variables in hair sheep located in an arid region. *Int J Biometeorol* 2016;60(8):1279-1286.
34. Charmandari E, Tsigos C, Chrousos G. Endocrinology of the stress response. *Annu Rev Physiol* 2005;67:259-284.
35. Braden A, Mattner P. The effects of scrotal heating in the ram on semen characteristics, fecundity, and embryonic mortality. *Aust J Agric Res* 1970;21(3):509-518.
36. Chella L, Kunene N, Lehloenya K. A comparative study on the quality of semen from Zulu rams at various ages and during different seasons in KwaZulu-Natal, South Africa. *Small Ruminant Res* 2017;151:104-109.
37. Maurya VP, Sejian V, Kumar D, Naqvi SM. Effect of induced body condition score differences on sexual behavior, scrotal measurements, semen attributes and endocrine responses in Malpura rams under hot semi-arid environment. *J Anim Physiol Anim Nutr (Berl)* 2010;94(6):e308-e317.
38. Benia A, Taibi K, Ait-Amrane A, Belhamiti T, Hammoudi S, Kaidi R. Study of seasonal sexual activity variations in Algerian rams: Sexual behaviour, testosterone concentration control and environmental factors. *Afr J Biotechnol* 2013;12(41):6042-6048.

39. Cárdenas-Gallegos M, Aké-López J, Magaña-Monforte J, Centurión-Castro F. Libido and serving capacity of mature hair rams under tropical environmental conditions. Arch Med Vet 2015;47(1):39-44.
40. Senger PL. Pathways to pregnancy and parturition. 3rd ed. Pullman, Washington: Current conceptions, Inc; 2012.
41. Byers SW, Glover TD. Effect of scrotal insulation on the pituitary-testicular axis of the ram. J Reprod Fertil 1984;71(1):23-31.
42. Huanca W, Coronado L, Galloway DB. Efecto de la manipulación de la temperatura escrotal sobre las características clínicas, seminales y endocrinas en carneros. Rev Inv Vet Perú 2015;26(4):604-613.
43. Narayan E, Parisella S. Influences of the stress endocrine system on the reproductive endocrine axis in sheep (*Ovis aries*). Ital J Anim Sci 2017;16(4):640-651.
44. Tsigos C, Papanicolaou DA, Kyrou I, Raptis SA, Chrousos GP. Dose-dependent effects of recombinant human interleukin-6 on the pituitary-testicular axis. J Interferon Cytokine Res 1999;19(11):1271-1276.
45. Patel N, Kashanian JA. Thyroid dysfunction and male reproductive physiology. Semin Reprod Med 2016;34(6):356-360.
46. Hassanpour H, Kadivar A, Mirshokraei P, Nazari H, Afzali A, Badisanaye M. Connexin-43: A possible mediator of heat stress effects on ram Sertoli cells. Vet Res Forum 2015;6(2):125-130.
47. Alves MB, Andrade AF, Arruda RP, Batissaco L, Florez-Rodriguez SA, Oliveira BM, *et al.* Recovery of normal testicular temperature after scrotal heat stress in rams assessed by infrared thermography and its effects on seminal characteristics and testosterone blood serum concentration. Theriogenology 2016;86(3):795-805.e2.
48. Saab SA, Sleiman FT, Kallassy N, Darweesh WY, Aad PY. Effect of adaptation and heat stress on reproductive performances of fat-tail Awassi rams in eastern mediterranean. Leban Sci J 2011;12(1):31-44.
49. Karagiannidis A, Varsakeli S, Alexopoulos C, Amarantidis II. Seasonal variation in semen characteristics of Chios and Friesian rams in Greece. Small Ruminant Res 2000;37(1):125-130.
50. Panyaboriban S, Suwimonteerabutr J, Swangchan-Uthai T, Tharasanit T, Phutikanit N, Techakumphu M. Effect of heat stress on reproductive performance of an imported dorper ram: a case study in Thailand. Thai J Vet Med 2016;46(4):671-677.

51. Francis JR, Javvaji PK, Dhali A, Kolte AP, Roy SC, Giridhar K, *et al.* Seasonal variations in quality, preservability and fertilizing ability of ovine spermatozoa. *Biol Rhythm Res* 2019;1-12.
52. Marai IFM, El-Darawany AHA, Ismail ESAF, Abdel-Hafez MAM. Tunica dartos index as a parameter for measurement of adaptability of rams to subtropical conditions of Egypt. *Anim Sci J* 2006;77(5):487-494.
53. Maurya VP, Sejian V, Kumar D, Naqvi SMK. Impact of heat stress, nutritional restriction and combined stresses (heat and nutritional) on growth and reproductive performance of Malpura rams under semi-arid tropical environment. *J Anim Physiol Anim Nutr (Berl)* 2016;100(5):938-946.
54. Kafi M, Safdarian M, Hashemi M. Seasonal variation in semen characteristics, scrotal circumference and libido of Persian Karakul rams. *Small Ruminant Res* 2004;53(1):133-139.
55. Suhair SM, Abdalla MA. Effects of seasonal changes and shearing on thermoregulation, blood constituents and semen characteristics of desert rams (*Ovis aries*). *Pak J Biol Sci* 2013;16(24):1884-1893.
56. Belkadi S, Safsaf B, Heleili N, Tlidjane M, Belkacem L, Oucheriah Y. Seasonal influence on sperm parameters, scrotal measurements, and serum testosterone in Ouled Djellal breed rams in Algeria. *Vet World* 2017;10(12):1486-1492.
57. Moreira EP, Moura AdAA, Araújo AAd. Efeitos da insulação escrotal sobre a biometria testicular e parâmetros seminais em carneiros da raça Santa Inês criados no estado do Ceará. *R Bras Zootec* 2001;30(6):1704-1711.
58. Gastelum-Delgado MA, Avendaño-Reyes L, Álvarez-Valenzuela FD, Correa-Calderón A, Meza-Herrera CA, Mellado M, *et al.* Conducta estral circanual en ovejas Pelibuey bajo condiciones áridas del noroeste de México. *Rev Mex Cienc Pecu* 2015;6(1):109-118.
59. Escobar E, Lopes S, Malavolta C, Ramalho JB, Missio D, Pinto HF, *et al.* Effect of gamma-oryzanol on testicular degeneration induced by scrotal insulation in rams. *Theriogenology* 2019;128:167-175.
60. Rathore AK. A note on the effect of scrotal wool cover on morphological changes in ram spermatozoa due to heat stress. *Anim Prod* 1969;11(4):561-563.
61. Rathore AK. Acrosomal abnormality in ram spermatozoa due to heat stress. *Br Vet J* 1970;126(8):440-443.

62. Armengol MF, Sabino GA, Forquera JC, de la Casa A, Aisen EG. Sperm head ellipticity as a heat stress indicator in Australian Merino rams (*Ovis aries*) in Northern Patagonia, Argentina. *Theriogenology* 2015;83(4):553-559.e2.
63. Hamilton TR, Mendes CM, de Castro LS, de Assis PM, Siqueira AF, Delgado JdC, *et al.* Evaluation of lasting effects of heat stress on sperm profile and oxidative status of ram semen and epididymal sperm. *Oxid Med Cell Longev* 2016:12.
64. Silva LKX, Sousa JS, Silva AOA, Lourenco Junior JB, Faturi C, Martorano LG, *et al.* Testicular thermoregulation, scrotal surface temperature patterns and semen quality of water buffalo bulls reared in a tropical climate. *Andrologia* 2018;50(2):e12836.
65. Fleming JS, Yu F, McDonald RM, Meyers SA, Montgomery GW, Smith JF, *et al.* Effects of scrotal heating on sperm surface protein PH-20 expression in sheep. *Mol Reprod Dev* 2004;68(1):103-114.
66. Kahwage PR, Esteves SN, Jacinto MAC, Junior WB, Pezzopane JRM, de Andrade-Pantoja MH, *et al.* High systemic and testicular thermolytic efficiency during heat tolerance test reflects better semen quality in rams of tropical breeds. *Int J Biometeorol* 2017;61(10):1819-1829.
67. Hassanin S, Abdalla E, Kotby E, Abd-Elaziz A, El-Fouly M. Efficiency of asbestos shading for growth of Barki rams during hot summer. *Small Ruminant Res* 1996;20(3):199-203.
68. Can A, Denek N, Yazgan K. Effect of replacing urea with fish meal in finishing diet on performance of Awassi lamb under heat stress. *Small Ruminant Res* 2005;59(1):1-5.
69. Sejian V, Singh AK, Sahoo A, Naqvi SM. Effect of mineral mixture and antioxidant supplementation on growth, reproductive performance and adaptive capability of Malpura ewes subjected to heat stress. *J Anim Physiol Anim Nutr (Berl)* 2014;98(1):72-83.
70. Talib AAB, Bomboi G, Floris B. Does vitamin E or vitamin E plus selenium improve reproductive performance of rams during hot weather? *Ital J Anim Sci* 2009;8(4):743-754.
71. Kumar D, Naqvi SMK, Kumar S. Sperm motion characteristics of FecBBB and FecBB+ Garole x Malpura rams during the non-breeding season under hot semi-arid environment. *Livest Sci* 2012;150(1):337-341.
72. De K, Kumar D, Saxena VK, Naqvi SM. Study of circadian rhythmicity of physiological response and skin temperature of sheep during summer and winter in semi-arid tropical environment. *Physiol Behav* 2017;169:16-21.

73. Pantoja MHA, Esteves SN, Jacinto MAC, Pezzopane JRM, Paz CCP, Silva J, *et al.* Thermoregulation of male sheep of indigenous or exotic breeds in a tropical environment. *J Therm Biol* 2017;69:302-310.
74. Cwynar P, Kolacz R, Czerski A. Effect of heat stress on physiological parameters and blood composition in Polish Merino rams. *Berl Munch Tierarztl Wochenschr* 2014;127(5/6):177-182.