

# What is the master regulator of reproductive endocrinology? A sex-based functional review of the *KISS1-KISS1R* system

Izuchukwu A. Okafor<sup>1,2\*</sup>, Nnamdi A. Nzoniwu<sup>3</sup>, Chikwesiri E. Onyema<sup>4</sup>, and Sodiq Fakorede<sup>5</sup>

<sup>1</sup>Department of Radiology, Division of Translational Anatomy, UMass Chan Medical School, Worcester, Massachusetts, United States of America;

<sup>2</sup>Faculty of Basic Medical Sciences, Department of Anatomy, College of Health Sciences, Nnamdi Azikiwe University, Nnewi, Anambra State, Nigeria;

<sup>3</sup>Department of Biology, Applied Technology Schools, Abu Dhabi Vocational Education and Training Institute, Al Ain, Abu Dhabi, United Arab Emirates;

<sup>4</sup>Faculty of Basic Medical Sciences, College of Health Sciences, Department of Human Physiology, Nnamdi Azikiwe University, Nnewi, Anambra State, Nigeria; <sup>5</sup>Department of Prosthetics and Orthotics, Federal University of Technology, Owerri, Imo State, Nigeria

## Abstract

Kisspeptin, a peptide encoded by the *KISS1* gene, and its receptor, *KISS1R*, have emerged as pivotal regulators of reproductive endocrinology. This review explores the multifaceted roles of the *KISS1-KISS1R* system in both male and female reproduction, from puberty onset to gametogenesis, ovulation, pregnancy, and lactation. In males, kisspeptin influences spermatogenesis through intratesticular signaling, modulating steroidogenesis and sperm function. In females, it regulates follicular development, oocyte maturation, and ovulation by modulating gonadotropin release and ovarian signaling pathways. Beyond reproduction, kisspeptin has been implicated in metabolic regulation, energy balance, and reproductive behavior, highlighting its broader physiological significance. Recent studies suggest potential clinical applications, including its use as a biomarker for early pregnancy detection and as a therapeutic agent for infertility, polycystic ovary syndrome, and hyperprolactinemia. In addition, kisspeptin analogs have demonstrated promise in treating hormone-dependent malignancies, such as prostate cancer, by suppressing the hypothalamic-pituitary-gonadal axis. Compared to other reproductive hormones, kisspeptin stands out as an upstream regulator, directly controlling gonadotropin-releasing hormone secretion and the entire reproductive hormonal cascade. Given its central role, further research is warranted to elucidate its precise mechanisms and optimize its clinical applications. The *KISS1-KISS1R* system is undeniably a master regulator of reproductive endocrinology, presenting exciting opportunities for therapeutic advancements in reproductive medicine.

**Keywords:** Kisspeptin. *KISS1* gene. *KISS1R* gene. Human reproduction. Reproductive physiology. Molecular mechanisms.

## ¿Cuál es el regulador principal de la endocrinología reproductiva? Una revisión funcional del sistema *KISS1-KISS1R* con perspectiva de género

### Resumen

La kisspeptina, un péptido codificado por el gen *KISS1*, y su receptor, *KISS1R*, se han identificado como reguladores clave de la endocrinología reproductiva. Esta revisión explora los diversos roles del sistema *KISS1-KISS1R* en la reproducción masculina y femenina, desde el inicio de la pubertad hasta la gametogénesis, la ovulación, el embarazo y la lactancia. En los hombres, la kisspeptina influye en la espermatogénesis mediante la señalización intratesticular, modulando la esteroidogénesis y la función espermática. En las mujeres, regula el desarrollo folicular, la maduración del ovocito y la ovulación, modulando la liberación de gonadotropinas y las vías de señalización ovárica. Más allá de la reproducción, se ha implicado

#### \*Correspondence:

Izuchukwu A. Okafor

E-mail: Izuchukwu.Okafor@umassmed.edu

Date of reception: 01-04-2025

Date of acceptance: 14-07-2025

DOI: 10.24875/PER.25000002

Available online: 26-11-2025

Perinatol Reprod Hum. 2025;39(1):39-49

www.perinatologia.mx

0187-5337/© 2025. Instituto Nacional de Perinatología Isidro Espinosa de los Reyes. Published by Permaner. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

a la kisspeptina en la regulación metabólica, el balance energético y el comportamiento reproductivo, destacando su importancia fisiológica general. Los estudios recientes sugieren potenciales aplicaciones clínicas, incluyendo su uso como biomarcador para la detección temprana del embarazo y como agente terapéutico para la infertilidad, el síndrome de ovario poliquístico y la hiperprolactinemia. Además, los análogos de la kisspeptina se han mostrado prometedores en el tratamiento de neoplasias dependientes de hormonas, como el cáncer de próstata, al suprimir el eje hipotálamo-hipófisis-gónadas. Dado su papel central, se requiere más investigación para entender sus mecanismos precisos y mejorar sus aplicaciones clínicas. El sistema *KISS1-KISS1R* es, sin duda, un regulador principal de la endocrinología reproductiva, que ofrece emocionantes oportunidades para avances terapéuticos en medicina reproductiva.

**Palabras clave:** Kisspeptina. Gen *KISS1*. Gen *KISS1R*. Reproducción humana. Fisiología reproductiva. Mecanismos moleculares.

## Introduction

One essential quality of all living creatures required to continue life on Earth is reproduction. In addition to trying to adapt to and live in their surroundings, organisms need to create new ones to avoid going extinct<sup>1</sup>. Reproductive endocrinology involves complex signaling networks and feedback mechanisms to regulate reproductive functions in animals. Among these, the *KISS1* gene and its receptor *KISS1R* (*GPR54*) have emerged as key players<sup>2</sup>.

Kisspeptin is a peptide hormone encoded by the *KISS1* gene. Many investigations<sup>3-5</sup> have proven kisspeptin's important functions in the regulation of various elements of reproduction. Such important regulations include the regulation of the hypothalamic-pituitary-gonadal (HPG)<sup>6</sup>, upstream control over gonadotropin-releasing hormone (GnRH) neurons<sup>7-9</sup>, the regulation of reproductive events such as onset of puberty, sexual maturation, and fertility and ovulation, primarily through the tight control of GnRH neurons<sup>10,11</sup>.

Over the years, the *KISS1-KISS1R* system has been described as possessing a 'master' gene regulatory role in reproductive functions, especially through hormonal control<sup>11-14</sup>.

Therefore, this review aims to provide a comprehensive analysis of the *KISS1-KISS1R* system, focusing on its sex-based functional differences in various animal species. By examining the distinct roles of the *KISS1-KISS1R* system in male and female reproductive endocrinology, we aim to determine whether its influence is sufficiently significant to warrant the title of "master regulator". The review will highlight the system's involvement in sexual differentiation and regulation of various reproductive functions and events.

## Methods

This review adopted a traditional narrative approach with reference to the PRISMA 2020 guidelines to

enhance transparency in article selection, data organization, and synthesis, even though it does not qualify as a systematic review. A comprehensive literature search was conducted across four major academic databases – PubMed, ScienceDirect, Web of Science, and Google Scholar – covering studies published in English up to March 2025. The search strategy included the use of Boolean operators to combine relevant keywords such as "kisspeptin," "*KISS1R*," "*KISS1R*," "GnRH," "hypothalamic-pituitary-gonadal axis," "reproductive endocrinology," "spermatogenesis," "folliculogenesis," "oocyte maturation," "ovulation," "pregnancy," "lactation," "infertility," and "clinical applications of kisspeptin."

Articles were selected based on their relevance to the physiological, molecular, and clinical aspects of the kisspeptin signaling system in reproductive function. Peer-reviewed original research, reviews, and clinical studies involving mammalian or human models were included, provided they were published in English and addressed the objectives of the review. Studies were excluded if they were non-English, lacked accessible full texts, or were non-peer-reviewed commentaries or editorials that did not include primary data or substantial theoretical analysis.

After the initial search, duplicate records were removed, followed by title and abstract screening to identify studies that met the inclusion criteria. Full texts of the remaining articles were then assessed to ensure relevance and quality. The final body of literature included over 90 articles that were thematically analyzed and synthesized.

The reviewed literature was organized and discussed according to key thematic areas, including the anatomical and molecular characteristics of the *KISS1-KISS1R* system, its physiological roles in male and female reproductive function, its influence beyond reproduction – particularly in metabolic regulation and behavior – and its emerging clinical applications. The narrative approach allowed for critical integration of diverse

findings across experimental and clinical contexts, providing a comprehensive synthesis of the present state of knowledge while identifying knowledge gaps and future research directions.

### Characteristics of the *KISS1-KISS1R* system

The *KISS1-KISS1R* system, comprising the *KISS1* gene and its receptor *KISS1R* (also known as *GPR54*), plays a central role in the regulation of reproductive endocrinology. The system's significance lies in its ability to modulate the HPG axis, influencing processes such as puberty, fertility, and overall reproductive health<sup>6</sup>.

### MOLECULAR STRUCTURE OF *KISS1* GENE AND RECEPTOR

The *KISS1* gene encodes a peptide known as kisspeptin, which is a member of the RFamide family of neuropeptides. Kisspeptin exists in several forms, with the most studied being kisspeptin-54, kisspeptin-14, kisspeptin-13, and kisspeptin-10, named according to the number of amino acids they contain. The processing of the full-length kisspeptin-54 into shorter forms is a critical step in its biological activity<sup>15,16</sup>. All kisspeptins have the same affinity for their respective receptors<sup>17</sup>.

Furthermore, the *KISS1R* receptor is a G protein-coupled receptor that mediates the effects of kisspeptin. It is encoded by the *GPR54* gene and is primarily expressed in GnRH neurons in the hypothalamus, although it is also found in other tissues, including the placenta and liver<sup>16,17</sup>. Upon binding to kisspeptin, *KISS1R* activates intracellular signaling pathways that result in the release of GnRH, thereby influencing downstream reproductive functions.

There are notable differences in the expression and function of *KISS1-KISS1R* between males and females. For instance, in females, *KISS1* neurons are densely located in the anteroventral periventricular nucleus (AVPV), which is critical for the pre-ovulatory luteinizing hormone (LH) surge, while in males, these neurons are primarily located in the arcuate nucleus (ARC), reflecting their role in tonic GnRH secretion<sup>18</sup>.

### MECHANISM OF ACTION

Kisspeptin binds to the *GPR54/KISS1R* receptor, activating Gq/11protein, which then activates the phospholipase-C (PLC) enzyme. PLC hydrolyses

phosphatidylinositol bisphosphate to generate diacylglycerol (DAG) and inositol trisphosphate (IP<sub>3</sub>). DAG activates protein kinase C (PKC) to regulate gene expression, while (IP<sub>3</sub>) raises the concentration of Ca<sup>2+</sup> by stimulating its release from the endoplasmic reticulum. GnRH neurons get depolarized due to the rise in Ca<sup>2+</sup> causing the potassium channels to close preventing the outflow of potassium ions, and the cation transient receptor potential channels to open leading to a further influx of Ca<sup>2+</sup> into the cell (Fig. 1). As a result, hormones are secreted by GnRH neurons<sup>5</sup>. Kisspeptin stimulates the extracellular signal-regulated kinase and mitogen-activated protein kinase (MAPK) pathways through PKC, which in turn increases apoptosis and decreases cell proliferation and metastasis<sup>19</sup>.

### Kisspeptin in the hypothalamus

The hypothalamus is a crucial brain region responsible for maintaining homeostasis and regulating various endocrine functions, including reproduction. Kisspeptin neurons in the hypothalamus are integral to the control of GnRH secretion. These neurons provide direct stimulatory input to GnRH neurons, and the activation of GnRH neurons by kisspeptin triggers the release of gonadotropins (LH and FSH) from the anterior pituitary, which in turn regulate gonadal function<sup>20</sup>.

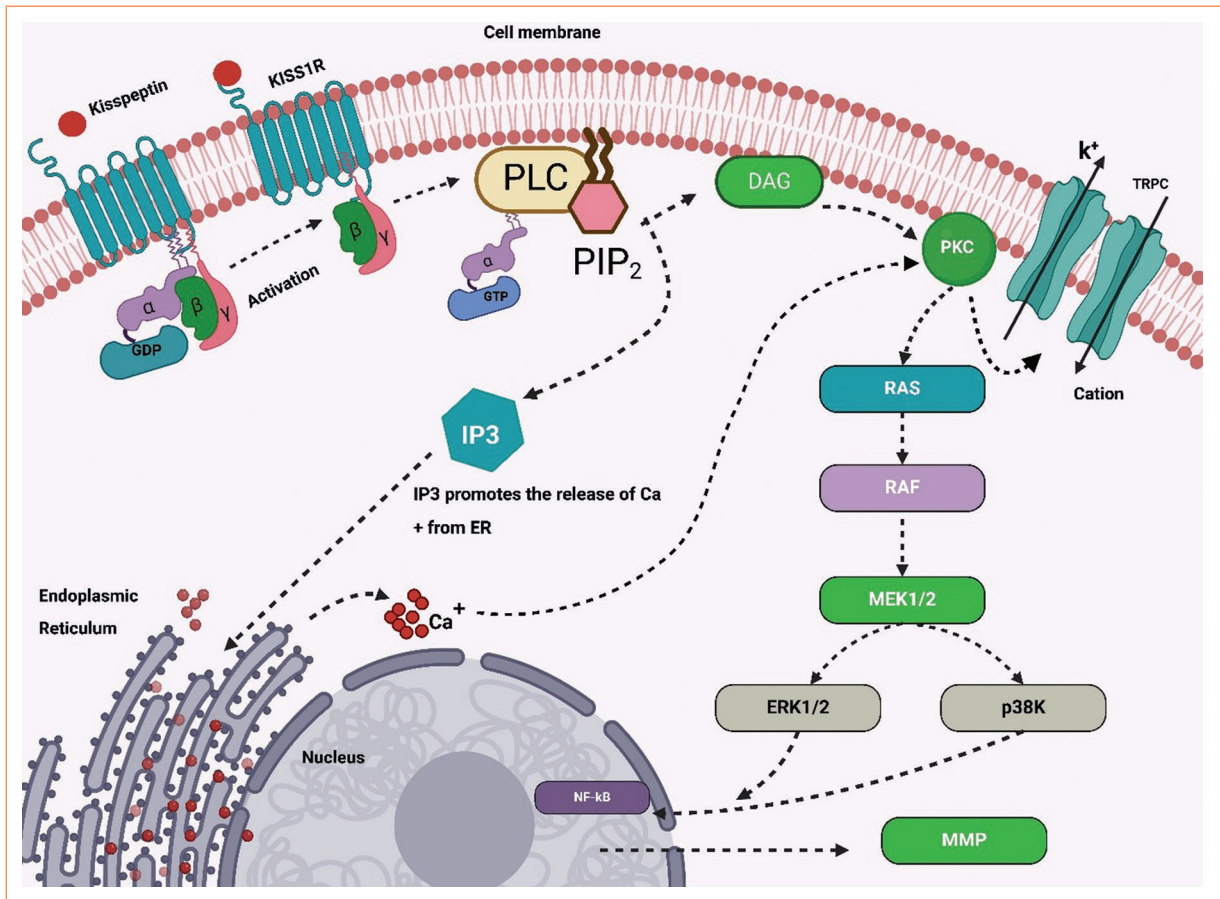
Kisspeptin is a critical neuropeptide that plays a vital role in regulating the HPG axis, thereby influencing reproductive function. Kisspeptin receptor (*KISS1R*) is highly expressed in the hypothalamus, particularly in regions such as the ARC and the AVPV<sup>21</sup>.

### REGULATION AND FEEDBACK MECHANISMS

The *KISS1-KISS1R* system is involved in both positive and negative feedback loops that regulate GnRH secretion. In females, the system mediates the estrogen-induced LH surge (positive feedback) that triggers ovulation, as well as the suppression of GnRH during the luteal phase (negative feedback) to prevent pre-mature ovulation. In males, *KISS1* neurons regulate the negative feedback of testosterone on GnRH secretion, ensuring the proper balance of reproductive hormones<sup>22</sup>.

### Kisspeptin and puberty

Puberty begins with the strengthening of excitatory signals and the weakening of inhibitory indications over GnRH neurons, resulting in a continual rise in pulsatile GnRH production from the hypothalamus. Increased



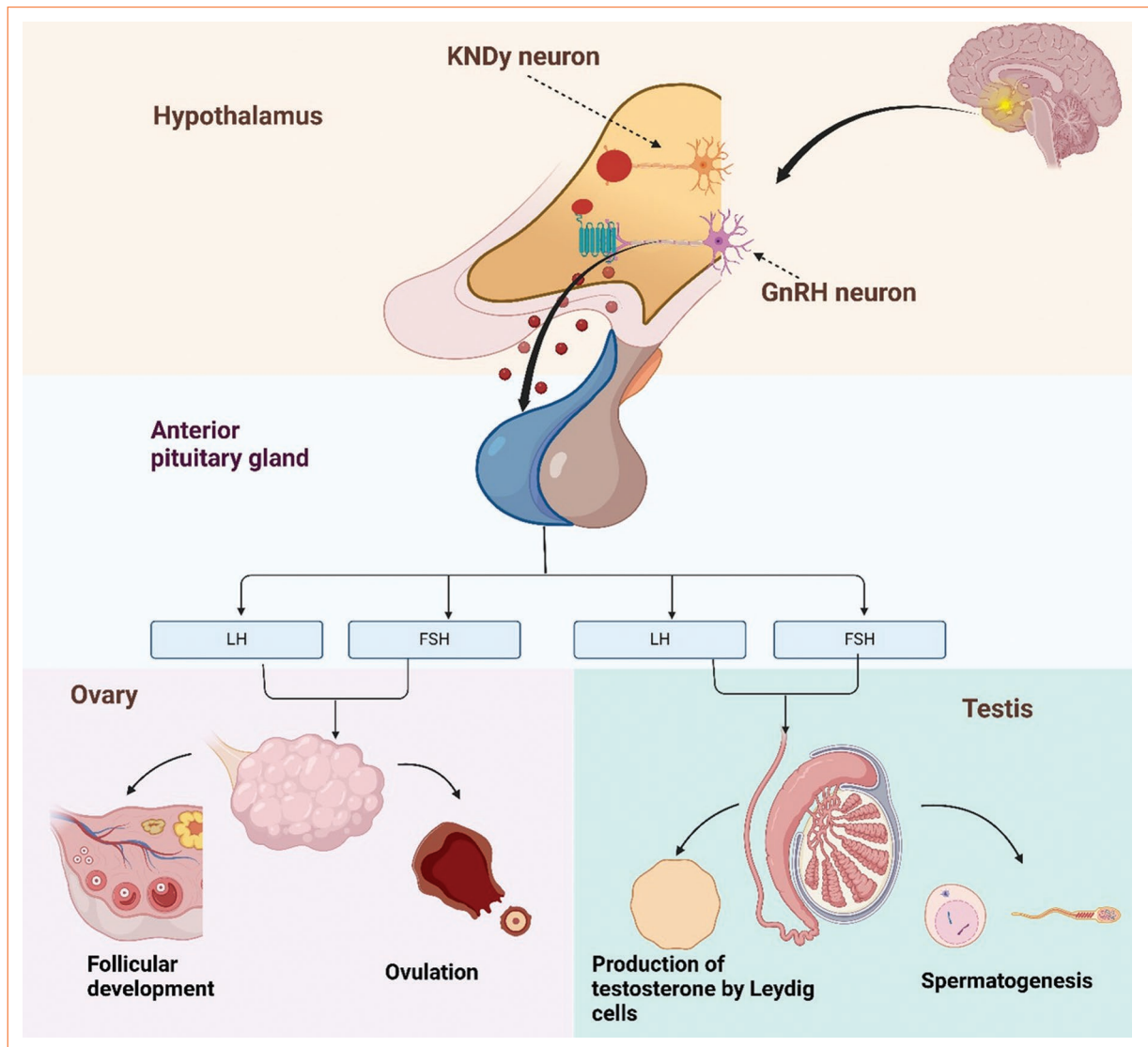
**Figure 1.** Intracellular signaling cascade activated by Kisspeptin Binding to the *KISS1R* receptor. This figure depicts the signaling cascade initiated by the binding of kisspeptin to the *KISS1R* (*GPR54*) receptor. Upon kisspeptin activation, the *KISS1R* receptor activates G-proteins, leading to the activation of phospholipase C (PLC). PLC catalyzes the breakdown of PIP<sub>2</sub> into inositol trisphosphate (IP<sub>3</sub>) and diacylglycerol (DAG), with IP<sub>3</sub> facilitating the release of calcium ions (Ca<sup>2+</sup>) from the endoplasmic reticulum (ER) into the cytosol. This increase in intracellular calcium activates protein kinase C (PKC) and the transient receptor potential canonical (TRPC) channels, which contribute to further calcium influx. The signaling cascade also involves the RAS-RAF-MEK1/2-ERK1/2 pathway, which regulates downstream gene expression through NF-κB and matrix metalloproteinase (MMP) activation. These signaling events play a crucial role in regulating gonadotropin-releasing hormone secretion, reproductive function, and cellular processes, as highlighted in the review.

GnRH pulse activates downstream components, resulting in increased gonadotropins and sex hormones, gametogenesis, secondary sex characteristics, and fast development, all of which contribute to fertility<sup>23</sup>. Although kisspeptin is vital for pubertal development in both males and females, there are notable differences in its function and regulation between the sexes.

In females, kisspeptin is integral to the initiation of puberty and the regulation of the menstrual cycle. The hormone exerts its effects by acting on GnRH neurons to initiate the pre-ovulatory surge of GnRH, which is critical for the onset of ovulation<sup>24</sup>. Kisspeptin neurons in the AVPV and the ARC of the hypothalamus are

particularly important in females. The AVPV kisspeptin neurons are implicated in the positive feedback mechanism of estrogen, which induces the LH surge necessary for ovulation<sup>25</sup>.

In males, kisspeptin is essential for the activation of the HPG axis during puberty, leading to increased testosterone production and spermatogenesis (Fig. 2). Unlike in females, the regulation of kisspeptin in males does not involve a pre-ovulatory surge but is crucial for maintaining steady-state levels of GnRH and LH secretion necessary for testicular function<sup>10</sup>. Kisspeptin neurons in the ARC play a more prominent role in males,



**Figure 2.** Regulation of the hypothalamic-pituitary-gonadal axis by the *KISS1-KISS1R* system. This figure illustrates the regulatory cascade of the hypothalamic-pituitary-gonadal (HPG) axis mediated by the *KISS1-KISS1R* system. In females, kisspeptin is primarily produced by neurons in the anteroventral periventricular nucleus (AVPV) of the hypothalamus. Here, kisspeptin stimulates gonadotropin-releasing hormone (GnRH) neurons, promoting the release of gonadotropins (LH and FSH) from the anterior pituitary, which subsequently trigger follicular development and ovulation. In males, kisspeptin is mainly synthesized by neurons in the arcuate nucleus (ARC). Kisspeptin in this region regulates the tonic secretion of GnRH, leading to continuous gonadotropin release that stimulates testosterone production by Leydig cells in the testes and supports spermatogenesis. Both in females and males, the interaction of kisspeptin with its receptor, *KISS1R* (*GPR54*), in the hypothalamus leads to downstream signaling events, resulting in the activation of the HPG axis and gonadal function.

where they are involved in the negative feedback regulation of gonadotropin secretion by testosterone<sup>26</sup>.

The sexually dimorphic nature of kisspeptin's role in puberty is influenced by sex steroids, such as estrogen and testosterone, which differentially regulate kisspeptin expression and action in the hypothalamus<sup>27</sup>.

The administration of kisspeptin has been investigated as a potential strategy to enhance pubertal development, particularly in cases of delayed or disrupted puberty. Studies in both male and female animal models have shown that exogenous kisspeptin can stimulate the HPG axis, leading to the onset of puberty.

In female models, kisspeptin administration has been shown to advance the timing of puberty by inducing the early release of GnRH and subsequent increases in LH and FSH levels, which promote ovarian development and estrogen production. This acceleration of puberty is associated with earlier onset of estrous cycles and ovulation<sup>28</sup>. In addition, kisspeptin treatment has been observed to restore reproductive function in female models with impaired kisspeptin signaling, indicating its therapeutic potential<sup>29</sup>.

In male models, kisspeptin administration has similarly been shown to advance the onset of puberty by stimulating GnRH release and increasing LH and FSH secretion. This leads to enhanced testosterone production and spermatogenesis<sup>12</sup>. The continuous or pulsatile administration of kisspeptin can effectively mimic the natural activation of the HPG axis, resulting in the advancement of pubertal milestones, such as testicular enlargement and increased secondary sexual characteristics<sup>30</sup>.

Puberty cannot occur without normal Kisspeptin-receptor contact, as demonstrated by inactivating mutations of the *GPR54* gene in hypogonadotropic hypogonadism (HH) patients<sup>31,32</sup>. Endogenous kisspeptin rhythmicity and sensitivity increase with puberty; in primates and rats, both the number of KISS1 neurons and the amount of KISS1 mRNA have been found to rise during the juvenile-pubertal transition<sup>33-35</sup>.

## Kisspeptin in male reproduction

### KISS1 AND KISS1R IN TESTIS AND SPERMATOZOA

Spermatogenesis is a complex process modulated step-by-step by the autocrine, paracrine, and endocrine routes. It needs the coordination between germ cell proliferation and death, meiotic division and differentiation events, and the key contribution of Leydig cells in the interstitium to produce sex-steroids, and Sertoli cells in the seminiferous tubules to provide structural and nourishment support to developing germ cells<sup>36</sup>. The kisspeptin system has been characterized in the testis of mammalian and non-mammalian vertebrates, revealing possible roles in the autocrine and paracrine intra-testicular communications, steroid biosynthesis, spermatogenesis progression, and sperm functions, but also species-specific differences in localization and possible functions<sup>37-39</sup>. Kisspeptin, but not GnRH, has been detected in human plasma and measured in different health conditions<sup>40</sup>. In males, circulating kisspeptin levels change in fertility status, being significantly higher in fertile than in infertile men<sup>41</sup>. Some

hypogonadotropic HH patients have high levels of kisspeptin in the plasma, but, after GnRH replacement therapy, circulating kisspeptin levels decrease as a consequence of the restored sex-steroid feedback mechanisms at hypothalamic levels<sup>42</sup>. However, gonadotropin stimulation is not always able to rescue testosterone biosynthesis and spermatogenesis in clinical cases of *KISS1R* inactivating mutations<sup>43,44</sup>, suggesting the need for testicular *KISS1R* signaling for steroidogenesis. Similarly, the specific reactivation of the *KISS1R* gene in the GnRH-secreting neuron of *KISS1R* knockout mice does not restore spermatogenesis, further confirming the need for the intratesticular kisspeptin signal for successful spermatogenesis<sup>45</sup>. By contrast, testosterone replacement in KISS knockout mice that exhibit HH restores plasma and intratesticular testosterone levels and sustains spermatogenesis until the production of spermatozoa capable of fertilizing eggs *in vitro*, but treated mice failed to impregnate females<sup>46</sup>. The administration of kisspeptin usually promotes spermatogenesis in intact animal models<sup>47-49</sup>. Post-natal testis development and Leydig cell maturation require kisspeptin signaling, and synergistic effects involving both the hypothalamic and LH-dependent intratesticular production of kisspeptin have been suggested in rodents<sup>50</sup>.

## Kisspeptin in female reproduction

### THE ROLE OF KISSPEPTIN IN FOLLICULAR DEVELOPMENT

Kisspeptin levels increase from the early follicular to the pre-ovulatory phase<sup>40</sup> in females. In the process of follicular development, kisspeptin affects primary and secondary follicle recruitment by reducing the FSH receptor (FSHR) expression. In both 6 and 10-month-old rats, local administration of kisspeptin into the ovary reduced the number of total antral follicles (including atretic follicles) and the use of kisspeptin receptor antagonist p234 played the opposite role<sup>51</sup>. In an *in vitro* experiment, kisspeptin acted as a functional antagonist by preventing the increase in FSHR expression produced by Isoproterenol, a  $\beta$ -adrenergic agonist. Besides, kisspeptin can upregulate the level of serum anti-Müllerian hormone (AMH), which is a vital dimeric glycoprotein in the regulation of follicle development. Produced by pre-antral and small antral follicles, AMH exerts its regulatory role by attenuating primordial follicle recruitment and changing the sensitivity of follicles to FSH<sup>52,53</sup>. The study found that serum AMH levels increased after local administration of kisspeptin and

decreased after the use of p234 in 6 and 10-month-old rats. To sum up, kisspeptin may negatively affect the development of pre-antral follicles by upregulating AMH and downregulating the expression of FSHR in the ovary.

### THE ROLE OF KISSPEPTIN IN OOCYTE MATURATION

It is well known that the pre-ovulatory LH surge triggers the resumption of meiosis and the progression to metaphase II during each reproductive cycle<sup>54</sup>. Besides, the direct effect of kisspeptin on oocyte maturation has been studied in porcine cumulus-oocyte complexes (COCs). Adding kisspeptin to porcine COCs *in vitro* promotes oocyte maturation, suggesting kisspeptin acts on oocytes directly<sup>55</sup>. The mechanisms may include upregulating the expression of C-MOS, growth differentiation factor 9 (GDF 9), and bone morphogenetic protein 15 (BMP 15)<sup>56</sup>. C-MOS plays a stimulating role in various processes during oocyte maturation, including the meiosis process, normal spindle and chromosome formation, and reactivation of purified maturation-promoting factor after first meiosis. Furthermore, GDF 9 and BMP 15 take part in regulating follicle development, oocyte maturation, ovulation, luteinization, and other physiological processes<sup>56,57</sup>. It has been found that cumulus granulosa cells (GCs) play a vital role in regulating oocyte maturation. Several researchers have observed a remarkable expression of kisspeptin in gonadotropin-treated GCs, while *KISS1R* in oocytes, suggesting that GC-derived Kisspeptin may have a direct function on oocytes *KISS1R* to modulate oocyte maturation through a MAPK signaling pathway<sup>58-60</sup>. The kisspeptin expressed in GCs is estrogen receptor beta (ER $\beta$ ) dependent since the expression of kisspeptin in GCs is absent in ER $\beta$  knockout rat ovaries. Consistent with the findings above, the administration of kisspeptin can increase the maturity of oocytes without cumulus cells in both wild-type and ER $\beta$  knockout rats<sup>58</sup>. Therefore, kisspeptin may have a persistent and direct effect on oocytes in an autocrine and paracrine manner.

### THE ROLE OF KISSPEPTIN IN OVULATION

Ovulation is a complicated process described as the follicle rupture and oocyte release, which is mediated by the LH surge and is regulated by a series of specific genes<sup>61</sup>. At the end of the follicular phase, high levels of estrogen act on AVPV kisspeptin neurons, promoting the release of kisspeptin, which then causes the

cascade of GnRH surge, LH peak, and ovulation<sup>62</sup>. The functions of the LH peak are achieved by upregulating the expression of COX-2 and producing prostaglandin, which are essential for follicular rupture and ovulation<sup>63</sup>. It has been confirmed that peripheral kisspeptin administration induces ovulation in many species, such as rats and ewes<sup>64</sup>. The effect of kisspeptin on ovulation is mainly achieved by increasing the levels of LH and FSH. Subcutaneous administration of kisspeptin markedly elevated plasma FSH and LH levels in 25-day-old female rats<sup>65</sup>. In humans, the LH pulses increased immediately after an administration of kisspeptin-10<sup>40</sup>. Kisspeptin-54 induced ovulation in mice by stimulating precisely timed endogenous LH release of consistent amplitude and duration<sup>66</sup>. Both the expression of ovarian *KISS1* mRNA and the ovulation efficiency in rats could be reduced by the administration of a COX-2 inhibitor or a COX non-selective inhibitor, indicating that the upregulation of COX-2 may act on the expression levels of kisspeptin to induce the LH peak<sup>67</sup>. The role of ovarian kisspeptin in ovulation may not be indispensable because, in *KISS1R* knockout mice, standard gonadotropin priming could induce ovulation<sup>12</sup>, indicating that the ovarian kisspeptin signaling is not necessary for ovulation. However, although the oocyte quality between neuron-specific *KISS1* and *KISS1R* knockout mice and wild-type mice shows little difference, the knockout mice presented fewer ovulated oocytes and corpora lutea. This suggests the GnRH plus gonadotropin stimulation is not sufficient to reverse the loss of function due to *KISS1R* knockout<sup>68</sup>.

### THE ROLE OF KISSPEPTIN IN PREGNANCY AND LACTATION

Kisspeptin concentrations in human plasma increased dramatically throughout pregnancy, with the placenta producing the majority of it. Histochemical investigation revealed that *KISS1* mRNA is localized in syncytiotrophoblast; these findings imply that kisspeptin may play a function in regulating trophoblast invasion. The greatest levels of *KISS1* and *KISS1R* mRNAs in trophoblast cells correspond to maximal trophoblast invasion, which should be appropriately controlled.

Kisspeptin was reported to suppress the metastasis in cancer cells, and *KISS1* expression levels were revealed to be lower in metastatic compared with non-metastatic cancer tissue<sup>16,69</sup>. Trophoblasts are cells that are important for the growth and attachment of the placenta to the uterus. Trophoblast invasion resembles tumor metastasis<sup>70</sup> as the invading

trophoblasts follow a similar molecular mechanism for migration and invasion as tumor cells. The discovery of high levels of endogenous kisspeptin and *KISS1R* in placental trophoblast<sup>71,72</sup>, suggests its direct involvement in inhibiting cellular invasion, migration, and angiogenesis, thereby preventing excessive invasion of the endometrium by trophoblast cell<sup>73</sup>.

Recent research has demonstrated that central Kisspeptin 10 injection stimulates oxytocin neurons during the end of pregnancy and throughout lactation, indicating that Kisspeptin-induced oxytocin release is essential for parturition and lactation. Increased plasma Kisspeptin during pregnancy may thereby enhance oxytocin release; yet oxytocin receptor expression and sensitivity remain low before childbirth<sup>74</sup>.

However, studies in mice have revealed that prolactin treatment significantly decreased Kisspeptin expression in the hypothalamus, reducing GnRH release. Again, utilizing bromocriptine as a prolactin suppressor resulted in significantly enhanced *KISS1* mRNA expression in the rostral periventricular region of the third ventricle (RP3V) in mice. Furthermore, during breastfeeding, rats revealed lower expression of *KISS1* mRNA in the hypothalamus and LH secretion, resulting in the shutdown of the estrous cycle. Indeed, in virtually all animals, breastfeeding generates a period of infertility that allows for healthy offspring development and survival, and proper regulation of Kisspeptin expression contributes to lactational anovulation<sup>75,76</sup>.

### **The role of kisspeptin beyond reproduction**

Recent studies have shown that kisspeptin may also play a role in the regulation of energy balance and metabolism<sup>77</sup>. Report an altered metabolic phenotype in *KISS1R* knockout adult female mice. These mice showed significantly increased body weights and adiposity, impaired glucose regulation, and reduced energy expenditure from 10 weeks of age. Studies have shown that chronic intracerebroventricular kisspeptin treatment from post-natal day decreased body weight at day 60 in female rats<sup>78</sup>. This link suggests that kisspeptin acts as a mediator between metabolic status and reproductive health.

Kisspeptin has been implicated in the regulation of reproductive behaviors. It influences sexual motivation and mating behavior in animals, with evidence suggesting that it may modulate the neural circuits involved in these behaviours<sup>79</sup>. This role extends the influence of

the *KISS1-KISS1R* system beyond purely physiological aspects to encompass behavioral regulation as well.

The involvement of the *KISS1-KISS1R* system in memory and learning processes was suggested by the wide expression of *KISS1R* in the learning and memory-associated brain regions, including the dentate gyrus of the hippocampus and the cortical and medial nucleus of the amygdala<sup>80</sup>. It has been reported that intracerebroventricular administration of kisspeptin-13, during a passive avoidance paradigm, enhances learning and stabilizes memory<sup>81</sup>.

### **The potential role kisspeptin in clinical application**

In recent years, many attempts have been made to explore the possibility of kisspeptin as a new diagnostic marker or therapeutic option. In humans, the plasma level of kisspeptin (kisspeptin-54) increases dramatically throughout pregnancy, making it possible to detect early pregnancy by measuring plasma kisspeptin concentrations<sup>82</sup>. Furthermore, because kisspeptin is produced by trophoblasts and trophoblast invasion is underway 5 days after blastocyst transplantation, plasma kisspeptin concentrations during the peri-implantation period may reflect the early developmental events associated with pregnancy outcome<sup>82</sup>. In a comparative study<sup>83</sup>, measured serum kisspeptin in 20 women with 6-10 weeks of intrauterine pregnancy (IUP) and 20 women who suffered spontaneous abortion (SAB) at a similar time. They found the median serum kisspeptin levels were significantly higher in IUP women (1.50 ng/mL) than in SAB women (0.20 ng/mL), indicating that kisspeptin is detectable in serum in early pregnancy and can discriminate SAB from IUP<sup>83</sup>. Girls with central precocious puberty (CPP) had higher serum kisspeptin levels compared with healthy girls<sup>84,85</sup>. However, serum kisspeptin levels are not able to become a single diagnostic tool because the evident overlap limits its use, while it may still be useful as an adjunctive tool in the diagnosis of CPP. As for therapeutic options, kisspeptin is found to have the potential to stimulate oocyte maturation and induce ovulation in infertile women. In 2014, a clinical study found that a single administration of kisspeptin-54 induced female egg maturation in women who accepted *in vitro* fertilization<sup>86</sup> suggesting its potential application in treating women with infertility. Kisspeptin and its agonist are also regarded as potential therapeutic options for some reproductive diseases. In a clinical study, repeated administration of kp-54 successfully induced ovulation

in two out of seven women with polycystic ovary syndrome (PCOS)<sup>87</sup>. Abbara and colleagues<sup>88</sup> compared the therapeutic effect of nanopeptide *KISS1R* agonist MT-602 and kisspeptin 54 in PCOS women and found both MVT-602 and kp54 induced an LH peak with similar amplitude. Recent studies also reported the application of kisspeptin as a future therapeutic option in the treatment of hyperprolactinemia, a condition characterized by elevated levels of prolactin, which inhibits GnRH, LH, and FSH secretion, and could cause irregular menstruation and infertility<sup>89</sup>. The administration of kisspeptin successfully caused LH pulses through stimulating GnRH in women with hyperprolactinemia<sup>90</sup>. In another study, the use of kisspeptin induced recovery of gonadotropin secretion<sup>89</sup>. In addition, based on the fact that a high dose of kisspeptin leads to desensitization of the HPG axis, kisspeptin may be applied in the treatment of sex hormone-dependent malignancies. For example, prostate cancer is a kind of androgen-dependent malignancy, and the present primary treatment is androgen deprivation therapy<sup>91</sup>. Two animal studies have confirmed that chronic administration of the kisspeptin analog, TAK-448 caused a stronger inhibiting effect on the HPG axis than GnRH analog and suppressed testosterone and LH release, indicating its great anti-tumor growth potential<sup>92,93</sup>.

## Conclusion

The *KISS1-KISS1R* system is a pivotal regulator of reproductive endocrinology, with profound implications for both male and female reproductive health. Its role in the activation of the HPG axis, particularly in the initiation of puberty, regulation of the menstrual cycle, and maintenance of fertility, underscores its central position in the reproductive hormonal hierarchy. The sex-specific expression and function of *KISS1-KISS1R* highlight the nuanced and essential roles it plays in coordinating reproductive processes in both sexes. Furthermore, the system's involvement extends beyond reproduction, influencing energy balance, metabolism, and reproductive behaviors, which points to its broader significance. When compared to other reproductive hormones, kisspeptin stands out due to its upstream regulatory role, controlling the release of GnRH and consequently the entire cascade of reproductive hormones. This unique position justifies the characterization of kisspeptin as the master regulator of reproductive endocrinology.

Future research into the *KISS1-KISS1R* system's mechanisms and potential clinical applications could

open new avenues for the treatment of reproductive disorders, enhancing our understanding of this complex and vital regulatory network. The evidence supports the notion that the *KISS1-KISS1R* system is indeed the “master regulator” of reproductive endocrinology, making it a key focus for ongoing and future studies in reproductive biology.

## Funding

This research has not received any specific grant from agencies in the public, commercial, or for-profit sectors.

## Conflicts of interest

The authors declare that they have no conflicts of interest.

## Ethical considerations

**Protection of humans and animals.** The authors declare that no experiments involving humans or animals were conducted for this research.

**Confidentiality, informed consent, and ethical approval.** The study does not involve patient personal data nor requires ethical approval. The SAGER guidelines do not apply.

**Declaration on the use of artificial intelligence.** The authors declare that ChatGPT 4.0 was used in this work, and it was utilized only for translation purposes from English to Spanish. No other AI tools were used.

## References

- Shcherbakov VP. Biological species is the only possible form of existence for higher organisms: the evolutionary meaning of sexual reproduction. *Biol Direct.* 2010;5:14.
- Xie Q, Kang Y, Zhang C, Xie Y, Wang C, Liu J, et al. The role of Kisspeptin in the Control of the hypothalamic-pituitary-gonadal axis and reproduction. *Front Endocrinol (Lausanne).* 2022;13:925206.
- Kauffman AS. Neuroendocrine mechanisms underlying estrogen positive feedback and the LH surge. *Front Neurosci.* 2022;16:953252.
- Kükürt A, Kuru M, Faruk Ba er Ö, Karapehlivan M. Kisspeptin: role in female infertility. In: *Reproductive Hormones*. London: IntechOpen; 2021.
- Wang HQ, Zhang WD, Yuan B, Zhang JB. Advances in the regulation of mammalian follicle-stimulating hormone secretion. *Animals.* 2021;11:1134.
- Sobrinho V, Avendaño MS, Perdices-López C, Jimenez-Puyet M, Tena-Sempere M. Kisspeptins and the neuroendocrine control of reproduction: recent progress and new frontiers in kisspeptin research. *Front Neuroendocrinol.* 2022;65:100977.
- Zhu N, Zhao M, Song Y, Ding L, Ni Y. The *KISS-1/GPR54* system: essential roles in physiological homeostasis and cancer biology. *Genes Dis.* 2022;9:28-40.
- Novaira HJ, Fadoju D, Diaczok D, Radovick S. Genetic mechanisms mediating Kisspeptin regulation of *GnRH* gene expression. *J Neurosci.* 2012;32:17391-400.
- Zhang C, Roepke TA, Kelly MJ, Rønnekleiv OK. Kisspeptin depolarizes gonadotropin-releasing hormone neurons through activation of TRPC-like cationic channels. *J Neurosci.* 2008;28:4423-34.

10. Pinilla L, Aguilar E, Dieguez C, Millar RP, Tena-Sempere M. Kisspeptin and reproduction: physiological roles and regulatory mechanisms. *Physiol Rev.* 2012;92:1235-316.
11. Roa J, Tena-Sempere M. KiSS-1 system and reproduction: comparative aspects and roles in the control of female gonadotropic axis in mammals. *Gen Comp Endocrinol.* 2007;153:132-40.
12. Seminara SB, Messenger S, Chatzidaki EE, Thresher RR, Acierno JS, Shagoury JK, et al. The *GPR54* gene as a regulator of puberty. *N Engl J Med.* 2003;349:1614-27.
13. Dungan HM, Clifton DK, Steiner RA. Minireview: kisspeptin neurons as central processors in the regulation of gonadotropin-releasing hormone secretion. *Endocrinology.* 2006;147:1154-8.
14. Uenoyama Y, Inoue N, Nakamura S, Tsukamura H. Central mechanism controlling pubertal onset in mammals: a triggering role of kisspeptin. *Front Endocrinol (Lausanne).* 2019;10:312.
15. Ohga H, Sakanoue R, Ohta K, Matsuyama M. Molecular characterization of kisspeptin 2 dodecapeptide in sixteen species of *Scombridae*. *Fish Sci.* 2020;86:437-44.
16. Ohtaki T, Shintani Y, Honda S, Matsumoto H, Hori A, Kanehashi K, et al. Metastasis suppressor gene *KISS-1* encodes peptide ligand of a G-protein-coupled receptor. *Nature.* 2001;411:613-7.
17. Kotani M, Detheux M, Vandenbogaerde A, Communi D, Vanderwinden JM, Le Poul E, et al. The metastasis suppressor gene *KISS-1* encodes kisspeptins, the natural ligands of the orphan G protein-coupled receptor *GPR54*. *J Biol Chem.* 2001;276:34631-6.
18. Morris PG, Herbison AE. Mechanism of arcuate kisspeptin neuron synchronization in acute brain slices from female mice. *Endocrinology.* 2023;164:bqad167.
19. Li M, Chen Y, Liao B, Tang J, Zhong J, Lan D. The role of kisspeptin and MKRN3 in the diagnosis of central precocious puberty in girls. *Endocr Connect.* 2021;10:1147-54.
20. Lehman MN, Merkley CM, Coolen LM, Goodman RL. Anatomy of the kisspeptin neural network in mammals. *Brain Res.* 2010;1364:90-102.
21. Rønnekleiv OK, Qiu J, Kelly MJ. Hypothalamic kisspeptin neurons and the control of homeostasis. *Endocrinology.* 2022;163:bqab253.
22. Clarke IJ, Smith JT. The role of kisspeptin and gonadotropin inhibitory hormone (GnIH) in the seasonality of reproduction in sheep. *Soc Reprod Fertil Suppl.* 2010;67:159-69.
23. Lents CA, Heidorn NL, Barb CR, Ford JJ. Central and peripheral administration of kisspeptin activates gonadotropin but not somatotropin secretion in prepubertal gilts. *Reproduction.* 2008;135:879-87.
24. Clarkson J, Han SK, Liu X, Lee K, Herbison AE. Neurobiological mechanisms underlying kisspeptin activation of gonadotropin-releasing hormone (GnRH) neurons at puberty. *Mol Cell Endocrinol.* 2010;324:45-50.
25. Gottsch ML, Clifton DK, Steiner RA. From KISS1 to kisspeptins: an historical perspective and suggested nomenclature. *Peptides (NY).* 2009;30(1):4-9.
26. Navarro VM, Gottsch ML, Chavkin C, Okamura H, Clifton DK, Steiner RA. Regulation of gonadotropin-releasing hormone secretion by kisspeptin/dynorphin/neurokinin B neurons in the arcuate nucleus of the mouse. *J Neurosci.* 2009;29:11859-66.
27. Kauffman AS, Gottsch ML, Roa J, Byquist AC, Crown A, Clifton DK, et al. Sexual differentiation of *KISS1* gene expression in the brain of the rat. *Endocrinology.* 2007;148:1774-83.
28. Plant TM, Ramaswamy S, DiPietro MJ. Repetitive activation of hypothalamic G protein-coupled receptor 54 with intravenous pulses of kisspeptin in the juvenile monkey (*Macaca mulatta*) elicits a sustained train of gonadotropin-releasing hormone discharges. *Endocrinology.* 2006;147:1007-13.
29. Castellano JM, Navarro VM, Fernández-Fernández R, Castaño JP, Malagón MM, Aguilar E, et al. Ontogeny and mechanisms of action for the stimulatory effect of kisspeptin on gonadotropin-releasing hormone system of the rat. *Mol Cell Endocrinol.* 2006;257-258:75-83.
30. Dhillon WS, Chaudhri OB, Patterson M, Thompson EL, Murphy KG, Badman MK, et al. Kisspeptin-54 stimulates the hypothalamic-pituitary gonadal axis in human males. *J Clin Endocrinol Metab.* 2005;90:6609-15.
31. Padda J, Khalid K, Moosa A, Syam M, Kakani V, Imdad U, et al. Role of Kisspeptin on hypothalamic-pituitary-gonadal pathology and its effect on reproduction. *Cureus.* 2021;13:e17600.
32. Singh SP, Kumar A, Sahu M, Sourya N, Singh AK. Application of kisspeptin in domestic animal reproduction. *Pharma Innov J.* 2021;3:1-4.
33. Bano R, Shamas S, Khan SH, Shahab M. Inverse age-related changes between hypothalamic *NPY* and *KISS1* gene expression during pubertal initiation in male rhesus monkey. *Reprod Biol.* 2022;22:100599.
34. Kauffman AS. Coming of age in the Kisspeptin era: sex differences, development, and puberty. *Mol Cell Endocrinol.* 2010;324:51-63.
35. Mayer C, Acosta-Martinez M, Dubois SL, Wolfe A, Radovick S, Boehm U, et al. Timing and completion of puberty in female mice depend on estrogen receptor alpha-signaling in kisspeptin neurons. *Proc Natl Acad Sci USA.* 2010;107:22693-8.
36. Pierantoni R, Cobellis G, Meccariello R, Fasano S. Evolutionary aspects of cellular communication in the vertebrate hypothalamo-hypophysio-gonadal axis. *Int Rev Cytol.* 2002;18:69-143.
37. Chianese R, Cobellis G, Chioccarelli T, Ciarrella V, Migliaccio M, Fasano S, et al. Kisspeptins, estrogens and male fertility. *Curr Med Chem.* 2016;23:4070-91.
38. Sharma A, Thaventhiran T, Minhas S, Dhillon WS, Jayasena CN. Kisspeptin and testicular function-is it necessary? *Int J Mol Sci.* 2020;21:2958.
39. Meccariello R, Fasano S, Pierantoni R. Kisspeptins, new local modulators of male reproduction: a comparative overview. *Gen Comp Endocrinol.* 2020;299:113618.
40. Trevisan CM, Montagna E, De Oliveira R, Christofolini DM, Barbosa CP, Crandall KA, et al. Kisspeptin/*GPR54* system: what do we know about its role in human reproduction? *Cell Physiol Biochem.* 2018;49:1259-76.
41. Ramzan MH, Ramzan M, Ramzan F, Wahab F, Jelani M, Khan MA, et al. Insight into the serum kisspeptin levels in infertile males. *Arch Iran Med.* 2015;18:12-7.
42. Kotani M, Katagiri F, Hirai T, Kagawa J. Plasma kisspeptin levels in male cases with hypogonadism. *Endocr J.* 2014;61:1137-40.
43. Meccariello R. The kisspeptin system in male reproduction. *Endocrines.* 2022;3:168-74.
44. Nimri R, Lebenthal Y, Lazar L, Chevrier L, Phillip M, Bar M, et al. A novel loss-of-function mutation in *GPR54/KISS1R* leads to hypogonadotropic hypogonadism in a highly consanguineous family. *J Clin Endocrinol Metab.* 2011;96:E536-45.
45. León S, Barroso A, Vázquez MJ, García-Galiano D, Manfredi-Lozano M, Ruiz-Pino F, et al. Direct actions of kisspeptins on GnRH neurons permit attainment of fertility but are insufficient to fully preserve gonadotropic axis activity. *Sci Rep.* 2016;6:19206.
46. Goto T, Hirabayashi M, Watanabe Y, Sanbo M, Tomita K, Inoue N, et al. Testosterone supplementation rescues spermatogenesis and *in vitro* fertilizing ability of sperm in KISS1 knockout mice. *Endocrinology.* 2020;161:bqaa092.
47. Chianese R, Ciarrella V, Fasano S, Pierantoni R, Meccariello R. Kisspeptin receptor, *GPR54*, as a candidate for the regulation of testicular activity in the frog *Rana esculenta*. *Biol Reprod.* 2013;88:73.
48. Chianese R, Ciarrella V, Fasano S, Pierantoni R, Meccariello R. Kisspeptin drives germ cell progression in the anuran amphibian *Pelophylax esculentus*: a study carried out in *ex vivo* testes. *Gen Comp Endocrinol.* 2015;211:81-91.
49. Aytürk N, Firat T, Kükner A, Özoğul C, Töre F, Kandirali IE, et al. The effect of kisspeptin on spermatogenesis and apoptosis in rats. *Turk J Med Sci.* 2017;47:334-42.
50. Wang JY, Hsu MC, Tseng TH, Wu LS, Yang KT, Chiu CH. Kisspeptin expression in mouse Leydig cells correlates with age. *J Chin Med Assoc.* 2015;78:249-57.
51. Fernandois D, Na E, Cuevas F, Cruz G, Lara HE, Paredes AH. Kisspeptin is involved in ovarian follicular development during aging in rats. *J Endocrinol.* 2016;228:161-70.
52. Dewailly D, Andersen CY, Balen A, Broekmans F, Dilaver N, Fanchin R, et al. The physiology and clinical utility of anti-Müllerian hormone in women. *Hum Reprod Update.* 2014;20:370-85.
53. Fleming R. Anti-müllerian hormone: a personal view of the empowering analyte. *J Hum Reprod Sci.* 2020;13:257.
54. Sen A, Caiazza F. Oocyte maturation: a story of arrest and release. *Front Biosci.* 2013;S5:S383.
55. Saadeldin IM, Koo OJ, Kang JT, Kwon DK, Park SJ, Kim SJ, et al. Paradoxical effects of kisspeptin: it enhances oocyte *in vitro* maturation but has an adverse impact on hatched blastocysts during *in vitro* culture. *Reprod Fertil Dev.* 2012;24:656.
56. Persani L, Rossetti R, Di Pasquale E, Cacciatori C, Fabre S. The fundamental role of bone morphogenetic protein 15 in ovarian function and its involvement in female fertility disorders. *Hum Reprod Update.* 2014;20:869-83.
57. McNatty KP, Juengel JL, Reader KL, Lun S, Myllymaa S, Lawrence SB, et al. Bone morphogenetic protein 15 and growth differentiation factor 9 co-operate to regulate granulosa cell function. *Reproduction.* 2005;129:473-80.
58. Chakravarthy VP, Ghosh S, Housami SM, Wang H, Roby KF, Wolfe MW, et al. ER $\beta$  regulated ovarian kisspeptin plays an important role in oocyte maturation. *Mol Cell Endocrinol.* 2021;527:111208.
59. Reader KL, Heath DA, Lun S, McIntosh CJ, Western AH, Littlejohn RP, et al. Signalling pathways involved in the cooperative effects of ovine and murine GDF9+BMP15-stimulated thymidine uptake by rat granulosa cells. *Reproduction.* 2011;142:123-31.
60. Mottershead DG, Ritter LJ, Gilchrist RB. Signalling pathways mediating specific synergistic interactions between GDF9 and BMP15. *Mol Hum Reprod.* 2012;18:121-8.
61. Richards JS, Russell DL, Ochsner S, Espey LL. Ovulation: new dimensions and new regulators of the inflammatory-like response. *Annu Rev Physiol.* 2002;64:69-92.
62. Zeydabadi Nejad S, Ramezani Tehrani F, Zadeh-Vakili A. The role of Kisspeptin in female reproduction. *Int J Endocrinol Metab.* 2017;15:e44337.
63. Sirois J, Sayasith K, Brown KA, Stock AE, Bouchard N, Doré M. Cyclooxygenase-2 and its role in ovulation: A 2004 account. *Hum Reprod Update.* 2004;10:373-85.
64. Caraty A, Smith JT, Lomet D, Ben Saïd S, Morrissey A, Cogne J, et al. Kisspeptin synchronizes preovulatory surges in cyclical ewes and causes ovulation in seasonally acyclic ewes. *Endocrinology.* 2007;148:5258-67.

65. Matsui H, Takatsu Y, Kumano S, Matsumoto H, Ohtaki T. Peripheral administration of metastatin induces marked gonadotropin release and ovulation in the rat. *Biochem Biophys Res Commun*. 2004;320:383-8.
66. Owen CM, Zhou X, Bernard DJ, Jaffe LA. Kisspeptin-54 injection induces a physiological luteinizing hormone surge and ovulation in mice. *Biol Reprod*. 2021;104:1181-3.
67. Cao Y, Li Z, Jiang W, Ling Y, Kuang H. Reproductive functions of Kisspeptin/*KISS1R* systems in the Periphery. *Reprod Biol Endocrinol*. 2019;17:65.
68. Gaytan F, Garcia-Galiano D, Dorfman MD, Manfredi-Lozano M, Castellano JM, Dissen GA, et al. Kisspeptin receptor haplo-insufficiency causes premature ovarian failure despite preserved gonadotropin secretion. *Endocrinology*. 2014;155:3088-97.
69. Shengbing Z, Jing Feng L, Bin W, Lingyun G, Aimin H. Expression of *KISS-1* gene and its role in invasion and metastasis of human hepatocellular carcinoma. *Anat Rec (Hoboken)*. 2009;292:1128-34.
70. Soundararajan R, Rao AJ. Trophoblast "pseudo-tumorigenesis": significance and contributory factors. *Reprod Biol Endocrinol*. 2004;2:15.
71. Horikoshi Y, Matsumoto H, Takatsu Y, Ohtaki T, Kitada C, Usuki S, et al. Dramatic elevation of plasma metastatin concentrations in human pregnancy: metastatin as a novel placenta-derived hormone in humans. *J Clin Endocrinol Metab*. 2003;88:914-9.
72. Bilban M, Ghaffari-Tabrizi N, Hintermann E, Bauer S, Molzer S, Zoratti C, et al. Kisspeptin-10, a *KISS-1*/metastatin-derived decapeptide, is a physiological invasion inhibitor of primary human trophoblasts. *J Cell Sci*. 2004;117:1319-28.
73. Gomes VC, Sones JL. From inhibition of trophoblast cell invasion to proapoptosis: what are the potential roles of kisspeptins in preeclampsia? *Am J Physiol Regul Integr Comp Physiol*. 2021;321:R41-8.
74. Scott V, Brown CH. Beyond the GnRH axis: kisspeptin regulation of the oxytocin system in pregnancy and lactation. *Adv Exp Med Biol*. 2013;784:201-18.
75. Brown RS, Herbison AE, Grattan DR. Prolactin regulation of kisspeptin neurones in the mouse brain and its role in the lactation-induced suppression of kisspeptin expression. *J Neuroendocrinol*. 2014;26:898-908.
76. Grattan DR. 60 years of neuroendocrinology: the hypothalamo-prolactin axis. *J Endocrinol*. 2015;226:T101-22.
77. Tolson KP, Garcia C, Delgado I, Marooki N, Kauffman AS. Metabolism and energy expenditure, but not feeding or glucose tolerance, are impaired in young *KISS1R* KO female mice. *Endocrinology*. 2016;157:4192-9.
78. Sahin Z, Ozcan M, Ozkaya A, Canpolat S, Kutlu S, Kelestimur H. Percentages of serum, liver and adipose tissue fatty acids and body weight are affected in female rats by long-term Central kisspeptin treatments. *Arch Physiol Biochem*. 2023;129:307-15.
79. Clarkson J, Han SY, Piet R, McLennan T, Kane GM, Ng J, et al. Definition of the hypothalamic GnRH pulse generator in mice. *Proc Natl Acad Sci USA*. 2017;114:E10216-23.
80. Arai AC. The role of kisspeptin and *GPR54* in the hippocampus. *Peptides*. 2009;30:16-25.
81. Telegdy G, Adamik Á. The action of kisspeptin-13 on passive avoidance learning in mice. Involvement of transmitters. *Behav Brain Res*. 2013;243:300-5.
82. Hu KL, Zhao H, Yu Y, Li R. Kisspeptin as a potential biomarker throughout pregnancy. *Eur J Obstet Gynecol Reprod Biol*. 2019;240:261-6.
83. Sullivan-Pyke C, Haisenleder DJ, Senapati S, Nicolais O, Eisenberg E, Sammel MD, et al. Kisspeptin as a new serum biomarker to discriminate miscarriage from viable intrauterine pregnancy. *Fertil Steril*. 2018;109:137-41.e2.
84. Li M, Chen Y, Liao B, Tang J, Zhong J, Lan D. The role of kisspeptin and *MKRN3* in the diagnosis of central precocious puberty in girls. *Endocr Connect*. 2021;10(9):1147-54.
85. Rhie YJ, Lee KH, Eun SH, Choi BM, Chae HW, Kwon AR, et al. Serum Kisspeptin Levels in Korean Girls with Central Precocious Puberty. *J Korean Med Sci*. 2011;26(7):927.
86. Jayasena CN, Abbara A, Cominos AN, Nijher GMK, Christopoulos G, Narayanaswamy S, et al. Kisspeptin-54 triggers egg maturation in women undergoing in vitro fertilization. *Journal of Clinical Investigation*. 2014;124(8):3667-77.
87. Romero-Ruiz A, Skorupskaitė K, Gaytan F, Torres E, Perdices-Lopez C, Mannaerts BM, et al. Kisspeptin treatment induces gonadotropic responses and rescues ovulation in a subset of preclinical models and women with polycystic ovary syndrome. *Human Reproduction*. 2019;34(12):2495-512.
88. Abbara A, Eng PC, Phylactou M, Clarke SA, Richardson R, Sykes CM, et al. Kisspeptin receptor agonist has therapeutic potential for female reproductive disorders. *Journal of Clinical Investigation*. 2020;130(12):6739-53.
89. Sonigo C, Bouilly J, Carré N, Tolle V, Caraty A, Tello J, et al. Hyperprolactinemia-induced ovarian acyclicity is reversed by kisspeptin administration. *Journal of Clinical Investigation*. 2012;122(10):3791-5.
90. Hoskova K, Kayton Bryant N, Chen ME, Nachtigall LB, Lippincott MF, Balasubramanian R, et al. Kisspeptin Overcomes GnRH Neuronal Suppression Secondary to Hyperprolactinemia in Humans. *J Clin Endocrinol Metab*. 2022;107(8):e3515-25.
91. Penning TM, Dettlefsen AJ. Intracrinology-revisited and prostate cancer. *J Steroid Biochem Mol Biol*. 2020;196:105499.
92. Ishikawa K, Tanaka A, Kogame A, Watanabe T, Tagawa Y, Matsui H. Usefulness of pharmacokinetic/efficacy analysis of an investigational kisspeptin analog, TAK-448, in quantitatively evaluating anti-tumor growth effect in the rat VCaP androgen-sensitive prostate cancer model. *Eur J Pharmacol*. 2018;828:126-34.
93. Matsui H, Tanaka A, Yokoyama K, Takatsu Y, Ishikawa K, Asami T, et al. Chronic Administration of the Metastatin/Kisspeptin Analog *KISS1-305* or the Investigational Agent *TAK-448* Suppresses Hypothalamic Pituitary Gonadal Function and Depletes Plasma Testosterone in Adult Male Rats. *Endocrinology* 2012;153(11):5297-308.