

Cyanotoxins bioaccumulation in freshwater ecosystems in Latin America: a review

Bioacumulación de cianotoxinas en ecosistemas dulceacuícolas en América Latina: una revisión

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ABSTRACT

Background: The increasing evidence of risk to the environment and human health by cyanotoxin exposure during cyanobacterial blooms has been reported worldwide. Despite the knowledge of cyanotoxin presence in Latin America, cyanotoxin bioaccumulation from freshwater environments have not been reviewed for the region. **Goals:** To review the current knowledge of cyanotoxin accumulation in tissues of freshwater organisms in field studies in Latin America. **Methods:** An extensive literature search was conducted to construct a database including information on accumulation of cyanotoxins in organisms inhabiting freshwater environments in Latin America (i.e., México to Argentina). **Results:** We found twenty-one studies from 2001 to 2020, including twenty-seven mostly eutrophic water bodies, the majority from Brazil. *Microcystis* was the most reported genus responsible for cyanotoxin production. Fish comprised most of the species accumulating cyanotoxins (20 species). Nile tilapia (*Oreochromis niloticus*) was the most studied species, and 80% of the fish species included have commercial importance, which highlights a potential route of exposure to humans by consumption of contaminated food. Some studies showed the reduction of cyanotoxins in tissues after an experimental depuration time. Also, calculations of the potential human intakes of microcystins by fish consumption exceeded the recommendations of tolerable intakes in most of the cases. **Conclusions:** In Latin America, the geographic extent of studies is narrow, however the summarized information indicates a risk for environment and human health by cyanotoxins bioaccumulation. There is a need for more efforts to generate scientific research on cyanotoxins bioaccumulation, but also for improvement of local level management policies to reduce eutrophication.

Keywords: aquaculture, human health impact, field studies, microcystins, saxitoxins.

RESUMEN

Antecedentes: La creciente evidencia de riesgo para el medio ambiente y la salud humana por la exposición a cianotoxinas durante las floraciones de cianobacterias se ha reportado en todo el mundo. A pesar del conocimiento de la presencia de cianotoxinas en América Latina, la bioacumulación de cianotoxinas en ambientes de agua dulce no ha sido revisada para la región. **Objetivos:** Revisar el conocimiento actual sobre la acumulación de cianotoxinas en tejidos de organismos de agua dulce en estudios de campo en América Latina. **Métodos:** Se realizó una extensa búsqueda bibliográfica para construir una base de datos que incluyera información sobre la acumulación de cianotoxinas en organismos que habitan ambientes de agua dulce en América Latina (de México a Argentina). **Resultados:** Encontramos veintiún estudios de 2001 a 2020, incluidos veintisiete cuerpos de agua en su mayoría eutróficos, la mayoría de Brasil. *Microcystis* fue el género productor de cianotoxinas más reportado. Los peces comprendieron la mayoría de las especies que acumulaban cianotoxinas (20 especies). *Oreochromis niloticus* fue la especie más estudiada, y el 80% de las especies de peces incluidas tienen importancia comercial, lo que destaca una vía potencial de exposición a los humanos. Algunos estudios demostraron la reducción de cianotoxinas en los tejidos después de un tiempo de depuración experimental. Además, los cálculos de las ingestas humanas potenciales de microcistinas por consumo de pescado excedieron las recomendaciones de ingestas tolerables en la mayoría de los casos. **Conclusiones:** En América Latina, los estudios se han realizado en pocos países, sin embargo, estos

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trabajos indican un riesgo para el medio ambiente y la salud humana por la bioacumulación de cianotoxinas. Se necesita más investigación científica sobre la bioacumulación de cianotoxinas, pero también esfuerzos para mejorar la gestión a nivel local con la finalidad de reducir la eutrofización.

Palabras clave: acuicultura, estudios de campo, impacto en la salud humana, microcistinas, saxitoxinas.

INTRODUCTION

Cyanobacteria, often referred as blue-green algae, are photoautotrophic microorganisms distributed ubiquitously in the world and are common inhabitants of a wide range of freshwater bodies. Favorable conditions, such as warm water, a stable water column, high phosphorus and nitrogen concentrations, high pH, low CO₂ availability and low herbivory (Zurawell *et al.*, 2005, Glibert & Burkholder 2018, Metcalf *et al.*, 2021), may lead to increased population growth of cyanobacteria generally called “cyanobacterial blooms” (Mowe *et al.*, 2015, Buratti *et al.*, 2017, Moreira *et al.*, 2022). Increases in cyanobacterial cell concentration reduces oxygen concentration and increases ammonia release when cyanobacteria decay. Blooms result in increased turbidity, leading to the decline of other primary producers, and they can enhance the likelihood of production of unpleasant taste and odor compounds (Hudnell & Dortch 2008, Wood 2016). Of major concern is increased levels of cyanotoxins that can be produced in about 40 cyanobacteria genera (Apeldoorn *et al.*, 2007, Singh & Dhar 2013). Cyanotoxins are secondary metabolites of diverse chemical structure and toxicity, including cyclic peptides, alkaloids, non-proteinogenic amino acids, phosphate ester and lipopolysaccharides (Aráoz *et al.*, 2010, Svirčev *et al.*, 2019). Based on their toxic effects in vertebrates, cyanotoxins are classified as hepatotoxins (microcystins, nodularin), neurotoxins (saxitoxins, anatoxins, homoanatoxin-a, B-methylamino-L-alanine), cytotoxins (cylindrospermopsins), dermatotoxins (aplysiatoxin, debromoaplysiatoxin, lingbyatoxin) and endotoxins (lipopolysaccharides) (Apeldoorn *et al.*, 2007, Svirčev *et al.*, 2019).

The frequency and magnitude of cyanobacterial blooms has received much more scientific attention around the world over the past decades, with blooms generally associated with the increment of eutrophication in freshwater bodies, watershed modifications and climate change (Paerl & Huismann 2009, Brooks *et al.*, 2016, Glibert 2020, Munoz *et al.*, 2021, Chorus *et al.*, 2021). The toxigenic and adverse effects of cyanotoxins in several groups of organisms have been recognized in field and laboratory settings including bacteria, microalgae, zooplankton, fishes, amphibians, birds, mammals and agricultural plants (Apeldoorn *et al.*, 2007, Valdor & Aboal 2007, Tillmanns *et al.*, 2008, Chen *et al.*, 2016, Banerjee *et al.*, 2021, Zhang *et al.*, 2021). This has led to a growing global consensus of the harmful effects for aquatic organisms and human health due to cyanotoxin exposure (Drobac *et al.*, 2013, Wood 2016, Cantoral Uriza *et al.*, 2017, Scarlett *et al.*, 2020). Specifically for humans, there are well documented events of poisoning that indicate that cyanotoxins were the cause of the symptoms (Humpage and Cunliffe, 2021), however, literature related to human intoxication by cyanotoxins exposure must be taken with caution since, usually, other potential causing agents (e.g., other bacteria and pollutants) have not been simultaneously evaluated (Testai *et al.*, 2016; Humpage and Cunliffe, 2021). In addition, there are other social and economic problems

associated to toxic cyanobacterial blooms, including cattle and companion animal deaths, loss of recreational fishing and irrigation value of water bodies, closures of drinking water supplies (Wood 2016, Munoz *et al.*, 2021) and loss of biodiversity (Hudnell & Dortch 2008).

At a global scale, cyanotoxin presence in water bodies (Buratti *et al.*, 2017, Svirčev *et al.*, 2019), accumulation in freshwater organisms (Ettoumi *et al.*, 2011, Testai *et al.*, 2016, Flores *et al.*, 2018, Pham & Utsumi 2018), and their transfer in aquatic food webs (Ferrão-Filho & Kozlowsky-Suzuki 2011, Lance *et al.*, 2014), have been documented and reviewed, highlighting the potential transfer to humans by fish or shellfish consumption (Fig. 1). For example, Sotton *et al.*, (2014) in Lake Hallwil (Switzerland) demonstrated the transfer of microcystins (MCYST's), produced during a bloom of the filamentous cyanobacteria *Planktothrix rubescens* (De Candolle ex Gomont) Anagnostidis & Komárek, from filter-feeders herbivorous zooplankton to predator zooplankton. Consequently, this zooplankton contributed to the contamination with MCYST's of the zooplanktivorous whitefish (*Coregonus suidteri* Fatio, 1885), a species with commercial value in that region. The evidence of biomagnification of cyanotoxins (i.e., the increase of concentrations in organisms as the toxin moves up the food chain) has only partial support and is still debated (Kozlowsky-Suzuki *et al.*, 2012, Flores *et al.*, 2018).

In Latin America (i.e., México to Argentina) the presence of cyanobacterial blooms and cyanotoxins in water bodies have been reported in several countries through field research and review papers, including: Argentina, Brazil, Chile, Costa Rica, Colombia, Cuba, Ecuador, Guatemala, México, Perú, Uruguay, Venezuela (Avendaño Lopez & Arguedas Villa 2006, Dörr *et al.*, 2010, Vasconcelos *et al.*, 2010, Gomez *et al.*, 2012, Mowe *et al.*, 2015, Pérez-Morales *et al.*, 2016, Rico-Martínez *et al.*, 2017, Moura *et al.*, 2018, León & Peñuela 2019, Munoz *et al.*, 2021, Salomón *et al.*, 2020). MCYST's are the most commonly reported cyanotoxins, followed by cylindrospermopsins (CYN's) and saxitoxin (STX's), and less commonly anatoxins (ATX's) and nodularin (NOD) (Galanti *et al.*, 2013, Sunesen *et al.*, 2021).

Despite the knowledge of cyanobacterial blooms and cyanotoxin presence in Latin America, relevant topics related to cyanotoxin impacts on the environment, including field studies on accumulation of cyanotoxins by freshwater organisms (Fig. 1), have not been reviewed for the region. Bioaccumulation of cyanotoxins is directly correlated with the potential human health risk from ingestion of contaminated tissue from organisms such as fish and shrimp. Moreover, in face of the climate change, population increase, watershed modifications and eutrophication increase, it is expected that cyanobacterial blooms will continue. Thus, there is a need of integrative analyses summarizing the knowledge on cyanotoxins bioaccumulation to highlight the potential impacts on freshwater organisms and on human health in Latin America, which could contribute to define a baseline for future studies, and to show pending topics of research. Based on the later ideas, in this review we aimed to summarize the current knowledge of cyanotoxins accumulation in freshwater organisms in Latin America. We analyzed and summarized major topics shared by studies in the region (i.e., depurations of cyanotoxins by organisms in field studies, potential human intake), highlighting the gaps in current knowledge and principal future directions in the region.

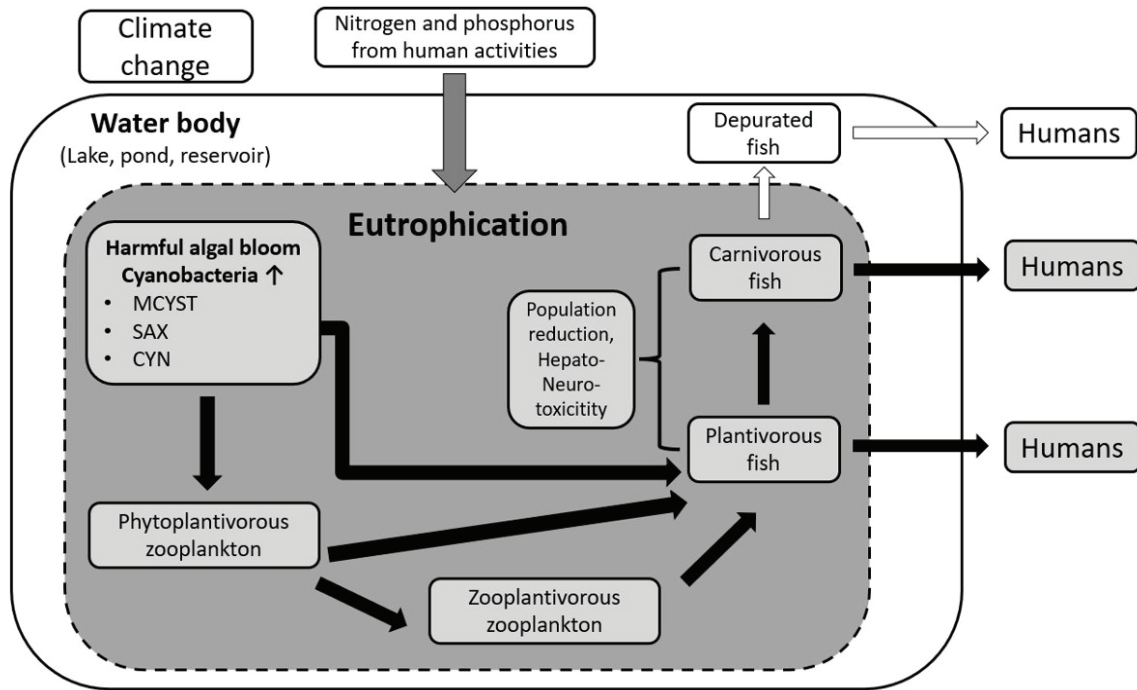


Figure 1. Schematic representation of cyanotoxins production and trophic transfer between several organisms in water bodies. Black arrows represent the cyanotoxins transfer.

MATERIALS AND METHODS

An extensive literature search was conducted to construct a database including information on accumulation of cyanotoxins in organisms inhabiting freshwater environments in Latin America. We collected all published studies to March 2022, using combinations of relevant search terms (i.e., cyanotoxins, Latin America, bioaccumulation, reservoir, aquaculture, cyanobacteria, human health impact) from the Digital Library of Universidad Nacional Autónoma de México, which includes about 170 scientific databases (e.g., Scopus, Web of Science, SciFinder) and Google Scholar. We focused on freshwater field studies in Latin America (from México to Argentina) reporting concentrations of cyanotoxins in organisms and water in natural or artificial/manipulated environments (e.g., ponds for aquaculture, reservoirs for hydroelectric energy). Because analytical methods for determination of concentrations of cyanotoxins in food matrices (e.g., fish and shrimps) is crucial to obtain reliable data, we followed Testai *et al.*, (2016) and Ibelings *et al.*, (2021) on determining the suitability and validation of the analytical methods employed in the studies.

From each study we collected the following information: author, year, country, location, freshwater environment type (e.g., lake, reservoir, pond), uses of the water body and land uses around it, reported nutrient concentrations (e.g., eutrophic), cyanotoxins analyzed and principal cyanobacterial genus/species present known to produce the toxins. For the land uses around the water body, if not reported in the study, we searched for this information in published literature. For the species accumulating cyanotoxins we collected: scientific name, native or introduced, importance in fishing (i.e., aquaculture, commercial/subsistence fishery), tissues and number of samples analyzed and concentrations of cyanotoxin in tissues. When studies reported several species

that accumulate cyanotoxins or the same species from several water bodies, information from each species or water body was registered as a different entry in the database. Data about concentrations of cyanotoxins in water was included as reported: diluted, in seston (particulate material in water) and/or total. When not directly reported, cyanotoxin concentration values were extracted from figures. Additionally, we collected information related to temporal fluctuations of cyanotoxins bioaccumulation, cyanotoxins depuration by organisms in field studies and potential human cyanotoxin intakes by consumption of contaminated organisms, which are developed in the next sections.

RESULTS AND DISCUSSION

Information from studies reporting cyanotoxins accumulation in freshwater organisms in Latin America was extracted from journal articles ($n=20$) and a master thesis ($n=1$). Cyanotoxins bioaccumulation has been studied only recently in Latin America, since the year of publication of studies found ranged from 2001 to 2020. From these, we gathered a total of 50 entries of several species accumulating cyanotoxins in the database in 27 different locations/water bodies (Table S1). The summary and discussion of the information presented below are referred to the 21 studies and 50 entries.

Considerations of analytical methods employed. In order to evaluate the reliability of the analytical methods used in the determination of cyanotoxins concentration in food matrices, including fish and invertebrates, Testai *et al.*, (2016) developed a 1 to 4 score system, where score 1 constitutes a reliable study without restriction (validated method); score 2, a reliable study (fully characterized method); score 3, a reliable study with restriction (only reporting recovery); and 4 denoted

a not reliable study (no report of recovery). Based on that score system, which does not depend exclusively on the analytical method, the main difference between a reliable and a non-reliable study is to report the % of recovery, often by spiking samples with known concentrations of analyte. It allows to detect the losses of cyanotoxin during sample processing and cleaning previous to the analytical determination method. It is in accordance with Flores *et al.*, (2018), which pointed, in their global study of accumulation of MCYST's in fish, that there is a need to standardize cyanotoxin extractions from tissues and analytical methods for quantification, or to report extraction efficiencies, since the method used is likely important in influencing the observed cyanotoxins concentration in tissues.

The analytical methods of cyanotoxins quantification in the database included high-pressure liquid chromatography (HPLC, 12 studies), enzyme-linked immunosorbent assay (ELISA, 10 studies) and liquid chromatography-mass spectrometry (LC-MS, two studies) (Table S1). However, only 5 studies (25%) reported the % of recovery of the corresponding cyanotoxin analyzed, all of them using HPLC (Cazanve *et al.*, 2005, Ame *et al.*, 2010, Galanti *et al.*, 2013, Morandi 2015, Calado *et al.*, 2019). The other 15 studies would be considered including "a not reliable analytical method" based on Testai *et al.*, (2016) score system. Additionally, 50% of the studies included ELISA as analytical method, which has been considered a semiquantitative method, requiring confirmation of identity (i.e., cyanotoxins variants), and quantity, with strategies to determine extraction efficiency and matrix effects, even more when it is used in a human risk assessment (Testai *et al.*, 2016, Lawton *et al.*, 2021, Ibelings *et al.*, 2021). Unfortunately, it is not the case in studies employing ELISA in Latin America: recoveries were not reported and confirmatory identity of cyanotoxins (and their variants) with a chemico-analytical method were usually not carried out. Exceptions are found in Berry & Lind (2010) and Berry *et al.*, (2011) where CYN's (by HPLC-UV and LC-MS) and MCYST variants (by LC-MS) were confirmed respectively.

Despite the noted pattern of limitations in the analytical methods employed in most of the Latin America studies and differences in cyanotoxins extraction methods used that were similar to the patterns found by Testai *et al.*, (2016) in their comprehensive review, we decided to continue summarizing the information about cyanotoxins bioaccumulation of the region, at least in a screening sense, in order to 1) visualize its geographically reported occurrence, 2) to call attention of the potential transfer of cyanotoxins by contaminated fish to humans and 3) to highlight the need of use reliable analytical methods especially when evaluating potential human risk exposure.

Distribution, water bodies and cyanotoxin types. Despite the global distribution of cyanobacteria and cyanotoxins (Mowe *et al.*, 2015, Flores *et al.*, 2018, Svirčev *et al.*, 2019) and the known potential risk for species that are exposed to cyanotoxins, including humans (Codd *et al.*, 2005, Burrati *et al.*, 2017, Cantoral Uriza *et al.*, 2017, Pham & Utsumi 2018, Banerjee *et al.*, 2021), cyanotoxins accumulation in freshwater organisms in Latin America has been evaluated only in four countries. Brazil is the country having more field studies of cyanotoxins bioaccumulation (13 studies), followed by México (4), Argentina (3) and, finally, Uruguay with only one study. Of the entries in the database, 37% comprise natural lake environments, and the others comprise artificial environments, including reservoirs (49%) and ponds for aquaculture (14%). Of special concern is the use of water in lakes and reservoirs for the potential cyanotoxin transfer to humans. For example, in natural lakes, the uses include mainly fishing and tourism, besides irrigation and aquaculture. Reservoir's water is mainly used for electric power generation and drinking water supply (increasing the risk of direct exposition to cyanotoxins by human). However, recreation, aquaculture and fishing are also important activities (Table S1). Notably, 80% of the water bodies included in our database are considered eutrophic, a nutrient condition (i.e., high phosphorus and nitrogen concentrations in water) widely associated with harmful cyanobacterial blooms worldwide (Glibert 2020, Chorus *et al.*, 2021). Clearly, human activities around the water bodies, and its corresponding watershed basin, alter the nutrients dynamics, indirectly by runoff from agriculture fields, pastures for livestock, vicinity of cities, and directly when sewage is discharged in the water bodies (Friesen, 2015). Most of the locations/water bodies included in the database, where information was available (Table S1), are bordered by agricultural fields and/or grassland, sometimes with a large associated human population. For example, Funil reservoir, constructed on the Paraíba do Sul River located near Rio de Janeiro in southern Brazil, has a catchment area of 12800 km² and is one of the most highly industrialized regions in Brazil. Approximately 2 million people live inside the catchment area in 39 cities that depend on the Paraíba do Sul River for their water supply (Deblois *et al.*, 2008, Pacheco *et al.*, 2015).

Almost all studies reported the principal cyanobacterial species/genus present in water bodies responsible of toxins production. *Microcystis* is the most reported genus producing toxins in Latin America, found in 11 studies and 13 different water bodies, followed by *Cylindropemopsis* (8 studies and 10 water bodies) and *Anabaena* (5 studies, 5 water bodies). Other less reported genera included *Planktothrix*, *Aphanizomenon*, *Pseudanabaena* and *Dolichospermum*, however, more than one genus was also detected within a water body (Table S1). Studies of cyanotoxins accumulation in Latin America have focused on four to-

Table 1. Cyanotoxins analyzed in accumulation studies in freshwater environments in Latin America. Some studies included more than one cyanotoxin or waterbody.

Cyanotoxin	Countries	Number of studies	Number of waterbodies	Group analyzed
MCYST's	Argentina, Brazil, México, Uruguay	15	25	fish, snail, zooplankton
STX's	Brazil, Mexico	6	3	fish, snail, zooplankton
CYN's	Mexico	2	1	fish, snail, zooplankton
NOD	Argentina	1	1	shrimp

xins with MCYST's being the most investigated with 15 studies in 25 different water bodies, followed by STX's, CYN's and NOD (Table 1). This result agrees with global research about cyanotoxin accumulation and distribution, where MCYST's are the most studied (Flores *et al.*, 2018), and highlights the need for more comprehensive studies including other cyanotoxins in Latin America.

Taxonomic groups and species accumulating cyanotoxins. Of the groups and species studied accumulating cyanotoxins in freshwater environments in Latin America, fish comprised most of the entries (90% of entries). Other entries corresponded to invertebrates: 2 entries for both snails and zooplankton (4% each group) and one entry for shrimps (2%). Mexico and Argentina are the countries where the invertebrate species accumulating cyanotoxins have been studied. In Mexico, Berry & Lind (2010) reported concentrations of STX's and CYN's accumulated in the Lake Catemaco endemic snail species, *Pomacea patula catemacensis* (Baker, 1922), locally known as "teogolos", a relevant species because it is both the target of local and commercial fishing. That study was the first report showing evidence of accumulation of STX's and CYN's in any organism in México, and for CYN's the first report in Latin America. Also in México, two studies analyzed concentrations of cyanotoxins in zooplankton, an ecologically important group which can

act as a vector of cyanotoxins, since some zooplankton directly consume cyanobacteria, and also are consumed by other zooplankton or fish (Sotton *et al.*, 2014). Berry *et al.*, (2012) found accumulations of STX's and CYN's in copepods (mainly *Mesocyclops* and *Arctodiaptomus*) from Lake Catemaco, while Zamora-Barrios *et al.*, (2019) reported MCYST's accumulation in copepods (*Acanthocyclops*) and cladoceran (*Daphnia* and *Bosmina*) from Lake Zumpango. In both studies, the concentrations of the cyanotoxins in zooplankton were notably higher than the concentrations in fish tissues. Zooplankton may ingest toxins directly from cyanobacteria and/or absorb dissolved cyanotoxins from water (Karjalainen *et al.*, 2005), which could explain their higher cyanotoxin concentration than fish, but this hypothesis should be tested. Finally, for invertebrates, Galanti *et al.*, (2013) carried out field exposures of the shrimp *Palaemonetes argentinus* (Nobili 1901) in San Roque Reservoir, Argentina, after a cyanobacteria bloom containing NOD. After three weeks of exposure in the reservoir, NOD was detected in *P. argentinus* tissues. Galanti *et al.*, (2013) noted that their study was the first report of NOD in South America freshwaters, and according to the present review, it is the first and unique report to date of accumulation of NOD in any freshwater organism. Also, it is the only report of the accumulation of cyanotoxins in shrimp in Latin America.

Table 2. Fish species included in studies of accumulation of cyanotoxins by commercial importance in Latin America. Countries where the studies were conducted and type of tissues analyzed for each fish species also included. References in table footnote. *indicates introduced species.

Species	Tissues analyzed	Countries	References
Commercial importance			
<i>Astyanax caballeroi</i> (Contreras-Balderas & Rivera-Teillery 1985)	muscle	Mexico	1
<i>Chirostoma jordani</i> Woolman 1894	whole fish	Mexico	2
<i>Chirostoma</i> sp.	Whole fish	Mexico	3
<i>Coptodon rendalli</i> (Boulenger 1897)*	liver, muscle, viscera	Brazil	4,5
<i>Cyprinus carpio</i> Linnaeus 1758*	liver, muscle	Mexico	3
<i>Geophagus brasiliensis</i> (Quoy & Gaimard 1824)	muscle	Brazil	6,7,8,19
<i>Goodea</i> sp.	viscera, muscle	Mexico	3
<i>Hoplias</i> sp.	muscle	Uruguay	9
<i>Hypophthalmichthys molitrix</i> (Valenciennes 1844)*	liver, muscle	Brazil	10
<i>Mayaheros urophthalmus</i> (Günther 1862)	muscle	Mexico	1
<i>Odontesthes bonariensis</i> (Valenciennes 1835)	liver, gills, brain, intestine, muscle	Argentina	11, 12
<i>Oreochromis aureus</i> (Steindachner 1864)*	muscle	Mexico	1
<i>Oreochromis niloticus</i> (Linnaeus 1758)*	liver, muscle, gills, intestine, gonads, bile	Brazil, Mexico	5, 13, 14, 15, 16, 17, 18
<i>Plagioscion squamosissimus</i> (Heckel 1840)*	liver	Brazil	10
<i>Rhamdia</i> sp.	muscle	Mexico	1
<i>Vieja fenestrata</i> (Günther 1860)	muscle	Mexico	1
<i>Vieja</i> sp.	muscle	Mexico	1
No commercial importance			
<i>Dorosoma petenense</i> (Günther 1867)	muscle	Mexico	1
<i>Pseudoxiphophorus jonesii</i> (Günther 1874)	muscle	Mexico	1
<i>Thorichthys helleri</i> (Steindachner 1864)	muscle	Mexico	1

¹Berry *et al.*, 2012, ²Zamora-Barrios *et al.*, 2019, ³Berry *et al.*, 2011, ⁴Magalhães *et al.*, 2001, ⁵Deblois *et al.*, 2008, ⁶Clemente *et al.*, 2010, ⁷Calado *et al.*, 2017, ⁸Calado *et al.*, 2019, ⁹Morandi 2015, ¹⁰Oliveira *et al.*, 2013, ¹¹Cazanave *et al.*, 2005, ¹²Amé *et al.*, 2010, ¹³Chellappa *et al.*, 2008, ¹⁴Galvao *et al.*, 2009, ¹⁵Vasconcelos *et al.*, 2013, ¹⁶Hauser-Davis *et al.*, 2015, ¹⁷Mendes *et al.*, 2016, ¹⁸Lopes *et al.*, 2020, ¹⁹Calado *et al.*, 2018.

Twenty fish species have been studied related to accumulation of cyanotoxins in Latin America, most of which are native species (70%) (Table 2). México is the country with more species (14), followed by Brazil (5). Argentina and Uruguay included only one species. Only four studies included more than one fish species and Nile tilapia, *Oreochromis niloticus* (Linnaeus 1758), is the most studied species, included in 8 studies. Most of the species showing accumulation of cyanotoxins have commercial importance (80%) by aquaculture, commercial and/or subsistence fishing, highlighting the potential exposure of cyanotoxins to humans (Table 2). Muscle is the most analyzed fish tissue for determination of concentration of cyanotoxins in 16 species and 47 entries in database, followed by liver (6 species, 23 entries), other tissues analyzed in some species included gills, intestine, bile, brain, gonads or whole fish (Table 2, Table S1). The analysis of several types of tissues is important because they usually show distinct patterns of cyanotoxins accumulations, and eventually could produce different harmful effects in fish and on human health. For example, among the tissues, the concentrations of MCYST's are often reported to be higher in liver (and viscera) relative to concentrations in the muscle (Romo *et al.*, 2012, Flores *et al.*, 2018). For 20 of 21 entries of our database that reported both muscle and liver concentrations of cyanotoxins (corresponding to 6 species used in aquaculture or commercial fishing), the concentration in the liver was reported to be higher than in the muscle. This is in agreement with the global pattern of Flores *et al.*, (2008). This tendency is important because the liver, and the viscera in general, are normally not eaten and are discarded in aquaculture fish (e.g., Nile tilapia, which comprised 16 of the 20 entries), and it is a manner to reduce the risk of human exposure when eating fish that has accumulated cyanotoxins. There are two studies including whole fish consumption in México. The fish genus *Chirostoma*, locally known as "charales", is widely captured in lakes and reservoirs in Central Mexico; they are small fish which are sold whole, fresh or dried, and are part of the culinary traditions of that region. Berry *et al.*, (2011) from Lake Pátzcuaro and Zamora-Barrios *et al.*, (2019) from Lake Zumpango, two eutrophic lakes with known presence of cyanobacterial blooms, reported high concentrations of MCYST's in whole *Chirostoma*, and suggested a particularly high potential for human exposure to food-derived cyanotoxins. Another case of the relevance of studying different tissues for cyanotoxins accumulations is presented by Hauser-Davis *et al.*, (2015). They studied the accumulation of MCYST's in Nile tilapia in the chronically contaminated and eutrophic Jacarepaguá lagoon, Brazil, finding a greater concentration of MCYST's in gonads than in liver. They suggested that this is of concern since this could signal potential reproductive problems in tilapia, which, as noted above, is an important product of aquaculture (Hauser-Davis *et al.*, 2015). All the above information highlights the relevance of studying several types of tissues depending on research goals. For example, from an ecological point of view of trophic transfer in natural ecosystems, it could be important to study cyanotoxins contains in whole organisms, probably separating different tissues. From a human health risk perspective, it could be relevant to study only muscle, if people are only eating fish fillets, for example.

Fish trophic habits may potentially influence the accumulation of cyanotoxins (Zhang *et al.*, 2009, Flores *et al.*, 2018). For example, fish that feed on cyanobacteria that produce toxins have a direct path of cyanotoxin exposure (i.e., planktivorous fish), which is not present in piscivores or omnivorous fish. Two studies in the database deal with this topic in Latin America. Berry *et al.*, (2011) analyzed the concentra-

tion of MCYST's in three fish species of different trophic habits in Lake Patzcuaro: the mainly phytoplanktivorous *Goodea* sp., the zooplanktivorous *Chirostoma* sp. and the omnivorous *Cyprinus carpio* Linnaeus 1758. All three species accumulated MCYST's, and its content correlated with fish trophic level, with concentrations of cyanotoxin measured as phytoplanktivorous > omnivorous > zooplanktivorous. The authors suggest that, although phytoplanktivorous zooplankton could be a source of MCYST's for the fish, and could increase the exposure for zooplanktivorous fish, the microfauna of the lake is characterized by species that are not efficient grazers on cyanobacteria. Also, Berry *et al.*, (2011) indicated that the accumulation of the cyanotoxin may be mainly associated with direct consumption of the cyanobacteria rather than to biomagnification to higher trophic levels. On the other side, Zamora-Barrios *et al.*, (2019) found a higher concentration of MCYST's in the zooplanktivorous *Chirostoma jordani* Woolman 1894 than in the omnivorous Nile tilapia (even liver) in Lake Zumpango. Indeed, future studies should specifically test the hypothesis of a relation between the trophic habit of fish and the accumulation of several cyanotoxins.

Fluctuations in bioaccumulation and depuration of cyanotoxins.

Patterns of temporal changes of cyanotoxins bioaccumulation in the field have been analyzed in seven studies in Latin America focusing on MCYST (Magalhaes *et al.*, 2001, Cazenave *et al.*, 2005, Ame *et al.*, 2010, Oliveira *et al.*, 2013, Zamora-Barrios *et al.*, 2019) and STX's (Clemente *et al.*, 2010, Calado *et al.*, 2017). Among them, some studies compared the accumulation of cyanotoxins in fish tissues between dry and wet season, however, although there were differences, there was not an obvious pattern of accumulation associated with a particular season. For example, Ame *et al.*, (2010) found accumulation of MCYST's in muscle of *Odontesthes bonariensis* (Valenciennes 1835) during both wet and dry season. The authors found temporal changes in the concentrations of MCYST variants (MCYST-LR, -RR, -LA and -YR), and suggested the need for an intensive monitoring program in that lake to ensure the health of people living in its surrounding. Also in *O. bonariensis*, Cazenave *et al.*, (2005) found a higher concentration of MCYST's in wet season in muscle, liver and gills, while Zamora-Barrios *et al.*, (2019) sampled Nile tilapia in three sampling dates and *C. jordani* in six, finding a higher concentration of MCYST's in tissues after the rains in both species when the decomposition of the *Microcystis* bloom occurred. Another study (Morandi 2015) suggested temporal fluctuations in MCYST's accumulation, since toxin presence and cyanobacterial blooms were previously reported at the Reservoir Rincón del Bonete in Uruguay. However intense sampling in the muscle of *Hoplias* sp. found no toxins, highlighting the importance of sampling in different seasons. Contrastingly, Calado *et al.*, (2017) did not find differences in the concentrations of STX's between dry and wet season, and, Clemente *et al.*, (2010) reported no significant difference in the STX's concentrations in muscle among the three samplings seasons (summer, spring, autumn), both studies carried out in Alagados Reservoir, Brazil with *Geophagus brasiliensis* (Quoy & Gaimard 1824).

Two studies included a long sampling period: Magalhaes *et al.*, (2001) sampled the accumulation of MCYST's in tissues of *Coptodon rendalli* (Boulenger 1897) for 40 months and Oliveira *et al.*, (2013) determined the accumulation of MCYST's in the phytoplanktivorous fish *Hypophthalmichthys molitrix* (Valenciennes 1844) during one year. Sampling in both studies was every two weeks. Six distinct phases based on MCYST's accumulations in different tissues were identified in Magalhães *et al.*, (2001). Although MCYST's were not detected in

Table 3. CYN's and STX's concentration ($\mu\text{g}/\text{kg}$) in muscle of different fish species by water body in Latin America including the corresponding daily intake by an adult and child ($\mu\text{g}/\text{kg}$ body weight). Adult intake calculated based on consuming 300g by an 70kg body weight (bw) person and child intake based on consuming 200g by an 30kg bw person. Intake in bold exceed the ARfD of 0.5 $\mu\text{g}/\text{kg}$ bw for STX's equivalents (EFSA, 2009). *denotes the use of reliable analytical methods

Cyanotoxin/Fish species	Water body	Toxin conc. ($\mu\text{g}/\text{kg}$)	Daily intake adult ($\mu\text{g}/\text{kg}$ bw)	Daily intake child ($\mu\text{g}/\text{kg}$ bw)	Reference
CYN's					
<i>Astyanax caballeroi</i> (Contreras-Balderas & Rivera-Teillery 1985)		0.81	0.003	0.005	
<i>Thorichthys helleri</i> (Steindachner 1864)		0.15	0.001	0.001	
<i>Mayaheros urophthalmus</i> (Günther 1862)		0.26	0.001	0.002	
<i>Dorosoma petenense</i> (Günther 1867)		0.8	0.003	0.005	
<i>Pseudoxiphophorus jonesii</i> (Günther 1874)	Catemaco Lake, México	1.26	0.005	0.008	Berry <i>et al.</i> , 2012
<i>Oreochromis aureus</i> (Steindachner 1864)		0.09	0.0004	0.001	
<i>Rhamdia</i> sp.		0.24	0.001	0.002	
<i>Vieja fenestrata</i> (Günther 1860)		0.81	0.003	0.005	
<i>Vieja</i> sp.		0.42	0.002	0.003	
STX's					
<i>Geophagus brasiliensis</i> (Quoy & Gaimard 1824)	Alagados Reservoir, Brazil	48	0.206	0.320	Calado <i>et al.</i> , 2017
		187.3	0.803	1.249	Calado <i>et al.</i> , 2019*
		12.2	0.052	0.081	Clemente <i>et al.</i> , 2010
<i>Astyanax caballeroi</i> (Contreras-Balderas & Rivera-Teillery 1985)		0.71	0.003	0.005	
<i>Thorichthys helleri</i> (Steindachner 1864)		0.06	0.0003	0.0003	
<i>Mayaheros urophthalmus</i> (Günther 1862)		0.32	0.001	0.002	
<i>Dorosoma petenense</i> (Günther 1867)		0.33	0.001	0.002	
<i>Pseudoxiphophorus jonesii</i> (Günther 1874)	Catemaco Lake, México	0.36	0.002	0.002	Berry <i>et al.</i> , 2012
<i>Oreochromis aureus</i> (Steindachner 1864)		0.03	0.0001	0.0001	
<i>Rhamdia</i> sp.		0.1	0.0004	0.001	
<i>Vieja fenestrata</i> (Günther 1860)		0.3	0.001	0.002	
<i>Vieja</i> sp.		0.22	0.001	0.001	

viscera and liver in some phases, they were always detected in muscle, with different concentrations of MCYST's depending on the phase (Magalhaes *et al.*, 2001). For *H. molitrix*, Oliveira *et al.*, (2013) found that during the drought months (April–September), the concentrations of toxins in muscle and liver were higher than in other months of the study period.

In general, the studies summarized here concerning fluctuations of cyanotoxins accumulation could reflect variations associated with type of cyanotoxin, accumulating species and/or geographic zone, which could be addressed in future analyses. Also, they highlight the need of site-specific studies of fluctuations of cyanotoxins accumulation in fish for a better understanding of potential human exposure by consumption of fish during annual changes in weather patterns.

Four studies analyzed the depuration of cyanotoxins (i.e., reduction or elimination of cyanotoxins after stop exposure during a specific time), finding contrasting results. First, Galvao *et al.*, (2009) analyzed in Nile tilapia from an artificial lake in Brazil the presence of MCYST's and STX's, NOD, ATX's and CYN's, and its depuration. Only STX variants were

detected in fish from the lake and, after a depuration time of five days without food in clean running water, the fish completely eliminated the STX's in muscle and liver (Galvao *et al.*, 2009). The authors suggested that depuration is a simple process that can be readily adopted by Nile tilapia producers as a way to eliminate STX's. In contrast, Calado *et al.*, (2017, 2019) analyzed the depuration of STX's in *G. brasiliensis* from Alagados Reservoir, Brazil, during 40 and 90 days respectively, finding that although there was a reduction of STX's in depurated fish, toxins were still present in fish muscle. Calado *et al.*, (2017) found a reduction in the percentage of specimens with the STX variant in the depurated fish, while the concentrations of the gonyautoxin 2 (GTX2) variant increased. They suggested that STX variant could be transformed to GTX2 and such transformation may decrease the toxicity of cyanotoxins to fish, since GTX2 is less toxic than STX. In addition, Calado *et al.*, (2019) determined the concentrations of STX's in the water and fish feces during the depuration time, suggesting that the fish biotransformed and eliminated STX's during the detoxification process and that the elimination of these toxins is possible but it takes a long time. Finally, Calado *et al.*, (2018) studied the depuration of MCYST's in *G. brasiliensis* during

90 days from Irai Reservoir, Brazil. They found that MCYST's concentrations increased in fish muscles during the depuration time at around 30 days, then MCYST's decreased but were still present in fish muscle at 90 days when the experiment finished. Calado *et al.*, (2018) argued that when MCYST's enter into cells they can be bound to phosphatase proteins and glutathione (GSH), and, during the depuration process, the toxins were metabolized and released. Thus, they initially increased in fish muscles, and finally MCYST's were excreted via feces and urine.

Future studies should focus on depuration patterns of other species and cyanotoxins, in order to determine the utility of the depuration process in reducing the exposure to humans when consuming fish exposed to cyanotoxins. Also, the suggested pattern of depuration of cyanotoxins by aquatic organisms in the field is promissory. It provides an option for species in natural environments to recover after a potential change in conditions of the waterbodies (e.g., after a management program to stop or reduce the eutrophication caused by human activities).

Potential human intake of cyanotoxins in Latin America. Several routes of human exposure have been recognized from water bodies containing cyanotoxins: use of drinking water, skin or nasal mucous membrane contact during recreational activities (e.g., swimming, canoeing or bathing), consumption of irrigated vegetables or fruits, consumption of aquatic organisms including fish, oral intake of cyanobacterial dietary supplements and dialysis (Drobac *et al.*, 2013, Ibelings *et al.*, 2021). In the case of the present review related to accumulation of cyanotoxins in freshwater organisms with some of them used as food, a tolerable daily intake (TDI) for lifetime has been recommended as a provisional guideline value for human consumption of contaminated organisms based on body weight (bw). For MCYST's the recommended TDI is 0.04 µg/kg bw by World Health Organization (WHO) (Falconer *et al.* 1999), and for CYN's of 0.03 µg/kg bw (Humpage & Falconer, 2003, Ibelings & Chorus, 2007). Also an acute reference dose (ARfD) of 0.5 µg STX's equivalents/kg bw by the European Food Safety Authority (EFSA, 2009) associated to the no-observed- adverse-effect level (NOAEL). In addition for MCYST's, Ibelings & Chorus (2007) calculated 2.5 µg/kg bw for the maximum tolerable intake to avoid an acute exposure (ATI) by a single consumption and 0.4 µg/kg bw for short term exposure tolerable intakes (STI), for example during a cyanobacterial bloom.

As noted in Section 3.1.1 above, almost all the Latin America studies of bioaccumulation of cyanotoxins showed limitations and flaws in the analytical methods employed, precluding a reliable risk assessment due to the consumption of contaminated fish based on these studies. However, in a screening sense and to alert to the risk of cyanotoxins through food in the region, we calculated potential human intakes for studies in the database which reported potential daily intakes and compared them with some guideline values.

We recalculated the potential human intake of cyanotoxins by assuming an adult to weigh 70 kg, consuming 300 g of fish (muscle or whole fish as usually eaten) and for a child weighing 30 kg and consuming 200 g of fish. We compared the potential intake values calculated from field studies with TDI, STI and ATI thresholds. When studies reported concentrations in tissues by sample points in a waterbody, several seasons, or different fish stages, we calculated the intake considering the higher concentrations reported. When reported several water bodies in a study for the same species, we calculated the daily intake for each water body.

Based on the only study reporting accumulation of CYN's in fish species in Latin America (Berry *et al.*, 2012), no CYN's concentration in muscle of the nine species reported from Catemaco Lake, México exceeded the TDI suggested (Humpage & Falconer, 2003; Ibelings & Chorus, 2007) (Table 3). For STX's, four studies (including a total of 10 species) reported intake calculations (Table 3), however, only Calado *et al.*, (2017) included concentrations in tissues exceeding the ARfD for *G. brasiliensis* from Alagados Reservoir, Brazil for both adult and child intake (exceeding 1.6 and 2.5-fold respectively).

Studies reporting intakes of MCYST's included 10 species in several waterbodies from Brazil, Mexico and Argentina (Table 4). From all these countries, some studies reported fish intakes exceeding TDI, some even exceeding STI and ATI. In Argentina, Cazanave *et al.*, (2005) reported potential intakes of MCYST's due to consumption of *O. bonariensis* from San Roque Reservoir to be higher than TDI, while for the same species from Los Padres Lake, Ame *et al.*, (2010) found intakes lower than TDI (Table 4). Most of the studies, including potential human intakes of MCYST's, come from Brazil for four species and 18 water bodies (Table 4). Among them, Oliveira *et al.*, (2013) reported the highest concentrations in muscle and, consequently, the highest daily intakes for a fish (*H. molitrix*) in Latin America, which exceeded by 168 and 261-fold the TDI respectively for adult and child. In this case, the intake also exceeded the ATI, and represented a potentially high risk of intoxication by consumption of this planktivorous species. Contradictingly, *H. molitrix* was introduced in Paranoá Lake as an attempt to reduce the amount of cyanobacteria, and became an important issue of health risks for its consumption by local people (Oliveira *et al.*, 2013).

The MCYST's intakes by consumption of Nile tilapia were reported in 19 cases (18 from Brazil), and in 10 of them exceeded the TDI (Table 4). Moreover, it is notably the study of Lopes *et al.*, (2020) where the MCYST's intakes of Nile tilapia from eight water bodies exceeded even the ATI. From Mexico, the MCYST's intake by consumption of *Goodea* sp. and *Chirostoma* sp. from Pátzcuaro lake (Berry *et al.*, 2011) and by *C. jordani* from Zumpango lake (Zamora-Barrios *et al.*, 2019) exceeded the TDI (also the STI for *Goodea* sp.). Related to differences between adult and child intakes, two studies including Nile tilapia from Brazil (DeBlois *et al.*, 2008) and *C. jordani* (Zamora-Barrios *et al.*, 2019) from Mexico showed intakes exceeding TDI for child but not exceeding for adults, highlighting the higher risk to children of exposure to harmful levels.

Similar to the summarized data reported by Ibelings and Chorus (2007) on their review of accumulation of cyanotoxins in freshwater seafood and its consequences for public health, there are variations in cyanotoxins intakes depending on fish species and water body, in many cases exceeding the TDI, and some doses exceeding ATI. For example, DeBlois *et al.*, (2010) found intakes exceeding TDI for Nile tilapia, but lower (and not exceeding TDI) for *C. rendalli* in Furnas Reservoir, suggesting that certain fish known to feed on cyanobacteria might be safer for consumption than others, which could be considered in the formulation of public health guidelines. Indeed, more research is needed in order to evaluate the health problems for people consuming exposed fish on a local scale and for particular waterbodies, and its correlation to the provisional guideline values suggested. Also, specific factors of water bodies and consuming habits of people including the frequency and for which time spans people will be exposed by their diet, the du-

ration of cyanotoxins occurrence in the water bodies where people get the fish, and other exposure routes acting synergistically, together with other risk conditions of expose people (i.e., health condition, age), will influence the potential risk for humans (Ibelings and Chorus 2007). Finally, reliable analytical methods for cyanotoxins concentrations in tissues of expose organisms should be implemented in future assessment of risk to human health in Latin America, even more considering the summarized patterns of risk in some cases using screening methods.

Future directions. The bioaccumulation of cyanotoxins and potential impacts on environment and human health constitute a complex scenario, where biological processes, but also social, economic, cultural, management, conservation and regulatory factors are involved. Some of these factors are out of the scope of the present review, however, we identified some future directions in order to reduce specifically for Latin America the potential harmful effects of cyanotoxin on the environment and humans.

Table 4. MCYST's concentration ($\mu\text{g}/\text{kg}$) in muscle of different fish species by country and water body in Latin America, including the corresponding daily intake by an adult and child ($\mu\text{g}/\text{kg}$ of body weight). Adult intake calculated based on consuming 300 g by a 70 kg body weight (bw) person and child intake based on consuming 200 g by a 30 kg bw person. Intakes in bold exceed the TDI of 0.04 $\mu\text{g}/\text{kg}$ bw for a lifetime suggested by the World Health Organization (Falconer *et al.* 1999). (All concentration for muscle except for *Chirostoma jordani* and *Chirostoma* sp., which includes the whole fish). *Denotes the use of reliable analytical methods, ^aExceed the "short-term" daily intake of 0.4 $\mu\text{g}/\text{kg}$ bw, ^bExceeded maximum tolerable intake to avoid an acute exposure by a single consumption of 2.5 $\mu\text{g}/\text{kg}$ bw.

Country/Species	Water body	Toxin conc. ($\mu\text{g}/\text{kg}$)	Daily intake adult ($\mu\text{g}/\text{kg}$ bw)	Daily intake child ($\mu\text{g}/\text{kg}$ bw)	Reference	
Argentina						
<i>Odontesthes bonariensis</i> (Valenciennes 1835)	Los Padres Lake	4.9	0.021	0.033	Ame <i>et al.</i> , 2010*	
	San Roque Reservoir	50	0.214	0.333	Cazanave <i>et al.</i> , 2005*	
Brazil						
<i>Coptodon rendalli</i> (Boulenger 1897)	Furnas Reservoir	1.7	0.007	0.011	DeBlois <i>et al.</i> , 2008	
	The Jacarepaguá lagoon	337.3	1.44^a	2.24^a	Magalhães <i>et al.</i> , 2001	
<i>Hypophthalmichthys molitrix</i> (Valenciennes 1844)	Paranoá Lake	1570	6.72^b	10.46^b	Oliveira <i>et al.</i> , 2013	
<i>Geophagus brasiliensis</i> (Quoy & Gaimard 1824)	Iraí Reservoir	4.6	0.01	0.03	Calado <i>et al.</i> , 2018	
	Acauã Reservoir	0.006	<0.002	<0.002	Mendes <i>et al.</i> , 2006	
	Acauã Reservoir	0.37	<0.002	0.002	Vasconcelos <i>et al.</i> , 2013	
	Araçagi Reservoir	0.019	<0.002	<0.002	Mendes <i>et al.</i> , 2006	
	Aracoíaba pond	570	2.44^a	3.8^b	Lopes <i>et al.</i> , 2020	
	Boqueirão do Casi Reservoir	0.018	<0.002	<0.002	Mendes <i>et al.</i> , 2006	
	Cacimba de Varzea Reservoir	0.03	<0.002	<0.002	Mendes <i>et al.</i> , 2006	
	Camalau Reservoir	0.16	<0.002	<0.002	Vasconcelos <i>et al.</i> , 2013	
	Castanhão pond	1040	4.45^b	6.93^b	Lopes <i>et al.</i> , 2020	
	Cordeiro Reservoir	0.005	<0.002	<0.002	Mendes <i>et al.</i> , 2006	
	<i>Oreochromis niloticus</i> (Linnaeus 1758)	Cordeiro Reservoir	0.23	<0.002	<0.002	Vasconcelos <i>et al.</i> , 2013
		Funil Reservoir	6.1	0.026	0.041	DeBlois <i>et al.</i> , 2008
		Furnas Reservoir	11.7	0.05	0.078	DeBlois <i>et al.</i> , 2008
		Furnas Reservoir	710	3.04^b	4.73^b	Lopes <i>et al.</i> , 2020
		Ilha Solteira Reservoir	350	1.5^a	2.33^a	Lopes <i>et al.</i> , 2020
Juara lake		625	2.67^b	4.16^b	Lopes <i>et al.</i> , 2020	
Linhares lake		740	3.17^b	4.93^b	Lopes <i>et al.</i> , 2020	
Orós pond		655	2.81^b	4.36^b	Lopes <i>et al.</i> , 2020	
The Jacarepaguá lagoon		2.75	0.011	0.018	Hauser-Davis <i>et al.</i> , 2015	
Tres Marias Reservoir		780	3.34^b	5.2^b	Lopes <i>et al.</i> , 2020	
México						
<i>Chirostoma jordani</i> Woolman 1894	Zumpango Lake	7	0.03	0.046	Zamora-Barrios <i>et al.</i> , 2019	
<i>Chirostoma</i> sp.	Pátzcuaro Lake	18.5	0.079	0.123	Berry <i>et al.</i> , 2011	
<i>Cyprinus carpio</i> Linnaeus 1758	Pátzcuaro Lake	4.99	0.021	0.033	Berry <i>et al.</i> , 2011	
<i>Goodea</i> sp.	Pátzcuaro Lake	157	0.67^a	1.047^a	Berry <i>et al.</i> , 2011	
<i>Oreochromis niloticus</i> (Linnaeus 1758)	Zumpango Lake	2	0.008	0.013	Zamora-Barrios <i>et al.</i> , 2019	

As highlighted in the present review, bioaccumulation of cyanotoxins have been accessed only in four countries in Latin America, a region comprising ~20 countries and investigations have focused mainly on MCYST's, for example, bioaccumulation of CYN's and NOD have only been analyzed twice and once from México and Argentina, respectively. Accordingly, Zurawell *et al.*, (2005) and Scarlet *et al.*, (2020) stated that some cyanobacterial toxins, including CYN's, are not routinely monitored around the world, and thus the range of toxins concentrations in natural systems is not known. There is a need for studies of bioaccumulation and monitoring, including less studied cyanotoxins and in more water bodies in more countries in Latin America, for a better understanding of potential risk of exposure to cyanotoxins. Aquaculture is a common activity in the region (Morales & Morales, 2006) and, as reported in the studies analyzed here, 80% of the species showing accumulation of cyanotoxins have commercial importance. To what extent the aquaculture constitutes an important route of transfer of cyanotoxins to humans is still unknown, and it should be determined in future studies. Also, as noted by Cantoral Uriza *et al.*, (2017), it is important to investigate unknown cyanobacteria species from tropical areas in Latin America to access their potential toxicity.

From an ecological point of view, there is evidence of behavioral alterations caused by cyanotoxins exposure in fish (Malbrouck & Kestemont 2006), including a decrease in locomotor activity and altered reproductive behavior (reduction in the spawning activity and success) (Baganz *et al.*, 1998). It is still unknown how changes in behavior associated with cyanotoxins exposure may affect fish populations in the wild (e.g., increasing or decreasing populations) and future studies could focus on the potential threat to conservation of native species by cyanotoxin exposure. Also, the trophic transfer of cyanotoxins has been widely documented (Ferrão-Filho & Kozłowski-Suzuki *et al.*, 2011, Lanca *et al.*, 2014), however, comprehensive analyses of food-chain effects of cyanotoxins (Zurawell *et al.*, 2005) and potential negative impacts in a wide range of organisms (i.e., insects, other vertebrates than fish) in freshwater natural systems have not been evaluated and constitute a pending area of research in Latin America. In this sense, the implementation of new approaches, including ecotoxicological signals using omics analyses, allowing the investigation of thousands of molecular responses of the cell to cyanotoxins at the same time (Marie 2020), will be particularly useful in future studies.

The present review summarized the current knowledge of cyanotoxins accumulation in freshwater organisms in Latin America, integrating information of major topics studied in the region, including groups and species accumulating cyanotoxins, fluctuations and depuration of cyanotoxins, and potential human intake of cyanotoxins from field studies. A further understanding and reduction of the harmful effects of cyanotoxins on the environment and on human health, specifically for Latin America, is a promising field for future research. Additionally, in the face of the population increase and watershed modifications in the region, combined with climate change and eutrophication that promotes cyanobacterial blooms, the study and monitoring of bioaccumulation of cyanotoxins should be mandatory. Also, assessing, monitoring and/or managing cyanobacterial/cyanotoxins risks in water-use systems is lacking for most of the countries in the region. This scenario indicates the need for more efforts to generate scientific research, but also, this research needs to be linked with national and local level management policies.

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REFERENCES

- AMÉ, M. V., L. N. GALANTI, M. L. MENONE, M. S. GERPE, V. J. MORENO & D. A. WUNDERLIN. 2010. Microcystin-LR, -RR, -YR and -LA in water samples and fishes from a shallow lake in Argentina. *Harmful Algae* 9 (1): 66-73. DOI:10.1016/j.hal.2009.08.001
- APELDOORN, M. E., H. P. VAN EGMOND, G. J. SPEIJERS & G. J. BAKKER. 2007. Toxins of cyanobacteria. *Molecular Nutrition & Food Research* 51 (1): 7-60. DOI:10.1002/mnfr.200600185
- ARÁOZ, R., J. MOLGÓ & N. T. DE MARSAC. 2010. Neurotoxic cyanobacterial toxins. *Toxicon* 56 (5): 813-828. DOI:10.1016/j.toxicon.2009.07.036
- AVENDAÑO LOPEZ, A., & C. ARGUEDAS VILLA. 2006. Microcistina en plantas de tratamiento de agua para consumo humano en un ambiente tropical: el Área Metropolitana de Costa Rica. *Revista de Biología Tropical* 54 (3): 711-716. DOI:10.15517/rbt.v54i3.12557
- BAGANZ, D., G. STAAKS & C. STEINBERG. 1998. Impact of the cyanobacteria toxin, microcystin-LR on behaviour of zebrafish, *Danio rerio*. *Water research* 32 (3): 948-952. DOI:10.1016/S0043-1354(97)00207-8
- BANERJEE, S., S. MAITY, R. GUCHHAIT, A. CHATTERJEE, C. BISWAS, M. ADHIKARI & K. PRAMANICK. 2021. Toxic effects of cyanotoxins in teleost fish: a comprehensive review. *Aquatic Toxicology* 240: 105971. DOI:10.1016/j.aquatox.2021.105971
- BERRY, J. P., & O. LIND. 2010. First evidence of "paralytic shellfish toxins" and cylindrospermopsin in a Mexican freshwater system, Lago Catemaco, and apparent bioaccumulation of the toxins in "tegogolo" snails (*Pomacea patula catemacensis*). *Toxicon* 55 (5): 930-938. DOI:10.1016/j.toxicon.2009.07.035
- BERRY, J. P., E. LEE, K. WALTON, A. E. WILSON & F. BERNAL-BROOKS. 2011. Bioaccumulation of microcystins by fish associated with a persistent cyanobacterial bloom in Lago de Pátzcuaro (Michoacan, Mexico). *Environmental Toxicology and Chemistry* 30 (7): 1621-1628. DOI:10.1002/etc.548
- BERRY, J. P., A. JAJA-CHIMEDZA, L. DÁVALOS-LIND & O. LIND. 2012. Apparent bioaccumulation of cylindrospermopsin and paralytic shellfish toxins by finfish in Lake Catemaco (Veracruz, Mexico). *Food Additives & Contaminants: Part A* 29 (2): 314-321. DOI:10.1080/19440049.2011.597785
- BROOKS, B. W., J. M. LAZORCHAK, M. D. HOWARD, M. V. V. JOHNSON, S. L. MORTON, D. A. PERKINS & J. A. STEEVENS. 2016. Are harmful algal blooms becoming the greatest inland water quality threat to public health and aquatic ecosystems?. *Environmental Toxicology and Chemistry* 35 (1): 6-13. DOI:10.1002/etc.3220
- BURATTI, F. M., M. MANGANELLI, S. VICHI, M. STEFANELLI, S. SCARDALA, E. TESTAI & E. FUNARI. 2017. Cyanotoxins: Producing organisms, occurrence, toxicity, mechanism of action and human health toxicological risk

- evaluation. *Archives of Toxicology* 91 (3): 1049-1130. DOI:10.1007/s00204-016-1913-6
- CALADO, S. L., J. WOJCIECHOWSKI, G. S. SANTOS, V. F. DE MAGALHÃES, A. A. PADIAL, M. M. CESTARI & H. C. D. S. DE ASSIS. 2017. Neurotoxins in a water supply reservoir: An alert to environmental and human health. *Toxicicon* 126: 12-22. DOI:10.1016/j.toxicicon.2016.12.002
- CALADO, S. L., G. S. SANTOS, T. P. B. LEITE, J. WOJCIECHOWSKI, M. N. JUNIOR, D. C. BOZZA, V. F. DE MAGALHÃES, M. M. CESTARI, V. PRODUCIMO & H. C. S. DE ASSIS. 2018. Depuration time and sublethal effects of microcystins in a freshwater fish from water supply reservoir. *Chemosphere* 210: 805-815. DOI:10.1016/j.chemosphere.2018.07.075
- CALADO, S. L., G. S. SANTOS, J. WOJCIECHOWSKI, V. F. DE MAGALHÃES, & H. C. S. DE ASSIS. 2019. The accumulation dynamics, elimination and risk assessment of paralytic shellfish toxins in fish from a water supply reservoir. *Science of The Total Environment* 651: 3222-3229. DOI:10.1016/j.scitotenv.2018.10.046
- CANTORAL URIZA, E. A., A. D. ASENSIO MARTÍNEZ & M. ABOAL SANJURJO. 2017. Cianotoxinas: efectos ambientales y sanitarios. Medidas de prevención. *Hidrobiológica* 27 (2): 241-251. DOI:10.24275/uam/izt/dcbis/hidro/2017v27n2/Cantoral
- CAZENAIVE, J., D. A. WUNDERLIN, M. A. BISTONI, M. V. AMÉ, E. KRAUSE, S. PFLUGMACHER & C. WIEGAND. 2005. Uptake, tissue distribution and accumulation of microcystin-RR in *Corydoras paleatus*, *Jenynsia multidentata* and *Odontesthes bonariensis*: a field and laboratory study. *Aquatic Toxicology* 75 (2): 178-190. DOI:10.1016/j.aquatox.2005.08.002
- CHELLAPPA, N. T., S. L. CHELLAPPA & S. CHELLAPPA. 2008. Harmful phytoplankton blooms and fish mortality in a eutrophicated reservoir of Northeast Brazil. *Brazilian Archives of Biology and Technology* 51: 633-641. DOI:10.1590/S1516-89132008000400022
- CHEN, L., J. CHEN, X. ZHANG, & P. XIE. 2016. A review of reproductive toxicity of microcystins. *Journal of Hazardous Materials* 301: 381-399. DOI:10.1016/j.jhazmat.2015.08.041
- FALCONER, I., J. BARTRAM, I. CHORUS, T. KUIPER-GOODMAN, H. UTKILEN, M. BURCH & G. A. CODD. 1999. Safe levels and safe practices. In: Chorus, I. & J. Bartram (Eds.) *Toxic Cyanobacteria in Water: A guide to their public health consequences, monitoring and management*. E & FN Spon, pp. 161-182.
- CHORUS, I., J. FASTNER, & M. WELKER. 2021. Cyanobacteria and cyanotoxins in a changing environment: Concepts, controversies, challenges. *Water* 13 (18): 2463. DOI:10.3390/w13182463
- CLEMENTE, Z., R. H. BUSATO, C. A. O. RIBEIRO, M. M. CESTARI, W. A. RAMSDORF, V. F. MAGALHÃES, & H. C. S. ASSIS. 2010. Analyses of paralytic shellfish toxins and biomarkers in a southern Brazilian reservoir. *Toxicicon* 55 (2-3): 396-406. DOI:10.1016/j.toxicicon.2009.09.003
- CODD, G. A., L. F. MORRISON, & J. S. METCALF. 2005. Cyanobacterial toxins: risk management for health protection. *Toxicology and Applied Pharmacology* 203 (3): 264-272. DOI:10.1016/j.taap.2004.02.016
- DEBLOIS, C. P., R. ARANDA-RODRIGUEZ, A. GIANI & D. F. BIRD. 2008. Microcystin accumulation in liver and muscle of tilapia in two large Brazilian hydroelectric reservoirs. *Toxicicon* 51 (3): 435-448. DOI:10.1016/j.toxicicon.2007.10.017
- DÖRR, F. A., E. PINTO, & R. M. SOARES. 2010. Microcystins in South American aquatic ecosystems: Occurrence, toxicity and toxicological assays. *Toxicicon* 56 (7): 1247-1256. DOI:10.1016/j.toxicicon.2010.03.018
- DROBAC, D., N. TOKODI, J. SIMEUNOVIĆ, V. BALTIC, D. STANIĆ, & Z. SVIRČEV. 2013. Human exposure to cyanotoxins and their effects on health. *Archives of Industrial Hygiene and Toxicology* 64 (2): 305-315. DOI:10.2478/10004-1254-64-2013-2320
- EFSA. 2009. Scientific Opinion: marine biotoxins in shellfish-saxitoxin group. *EFSA Journal* 1019: 1-76. DOI:10.2903/j.efsa.2009.1019
- ETTOUMI, A., F. EL KHALLOUFI, I. EL GHAZALI, B. OUDRA, A. AMRANI, H. NASRI & N. BOUAIÇA. 2011. Bioaccumulation of cyanobacterial toxins in aquatic organisms and its consequences for public health. In: Kattel, G (Ed.) *Zooplankton and Phytoplankton: Types, Characteristics and Ecology*. Nova Science Publishers, pp. 1-33.
- FERRÃO-FILHO, A. D. S., & B. KOZLOWSKY-SUZUKI. 2011. Cyanotoxins: bioaccumulation and effects on aquatic animals. *Marine Drugs* 9 (12): 2729-2772. DOI:10.3390/md9122729
- FLORES, N. M., T. R. MILLER, & J. D. STOCKWELL. 2018. A global analysis of the relationship between concentrations of microcystins in water and fish. *Frontiers in Marine Science* 5: 1-14. DOI:10.3389/fmars.2018.00030
- FRIESEN, C. 2015. Agricultural Runoff, Cyanobacterial Blooms in Lake Erie and Public Health: A Knowledge Synthesis. Master thesis. University of Guelph, Canada. 140p.
- GALANTI, L. N., M. V. AMÉ, & D. A. WUNDERLIN. 2013. Accumulation and detoxification dynamic of cyanotoxins in the freshwater shrimp *Palaeomonetes argentinus*. *Harmful Algae* 27: 88-97. DOI:10.1016/j.hal.2013.05.007
- GALVAO, J. A., M. OETTERER, M. DO CARMO BITTENCOURT-OLIVEIRA, S. GOUVÊA-BARRROS, S. HILLER, K. ERLER & P. KUJBIDA. 2009. Saxitoxins accumulation by freshwater tilapia (*Oreochromis niloticus*) for human consumption. *Toxicicon* 54 (6): 891-894. DOI:10.1016/j.toxicicon.2009.06.021
- GLIBERT, P. M., & J. M. BURKHOLDER. 2018. Causes of harmful algal blooms. In: Shumway, S. E., J. M. Burkholder & S. L. Morton (Eds.) *Harmful Algal Blooms: A Compendium Desk Reference*. John Wiley & Sons, pp. 1-38. DOI:10.1002/9781118994672.ch1
- GLIBERT, P. M. 2020. Harmful algae at the complex nexus of eutrophication and climate change. *Harmful Algae* 91: 101583. DOI:10.1016/j.hal.2019.03.001
- GÓMEZ, L., I. ÁLVAREZ & J. C. RODRÍGUEZ. 2012. CUBA: Toxic cyanobacteria risk assessment, research and management. In: Chorus, I (Ed.) *Current approaches to Cyanotoxin risk assessment, risk management and regulations in different countries*. Federal Environment Agency (Umweltbundesamt), pp. 40-48.
- HAUSER-DAVIS, R. A., R. T. LAVRADAS, R. C. LAVANDIER, E. G. A. ROJAS, A. W. S. GUARINO & R. L. ZIOLLI. 2015. Accumulation and toxic effects of microcystin in tilapia (*Oreochromis niloticus*) from a eutrophic Brazilian lagoon. *Ecotoxicology and Environmental Safety* 112: 132-136. DOI:10.1016/j.ecoenv.2014.10.036
- HUDNELL, H. K., & Q. DORTCH. 2008. A synopsis of research needs identified at the interagency, international symposium on cyanobacterial har-

- mfal algal blooms (ISOC-HAB). *In*: Hudnell, H. K. (Ed). Cyanobacterial harmful algal blooms: state of the science and research needs Springer, pp. 17-43.
- HUMPAGE, A. R., & I. R. FALCONER. 2003. Oral toxicity of the cyanobacterial toxin cylindrospermopsin in male Swiss albino mice: determination of no observed adverse effect level for deriving a drinking water guideline value. *Environmental Toxicology: An International Journal* 18 (2): 94-103. DOI:10.1002/tox.10104
- HUMPAGE, A. R., & D. CUNLIFFE. 2021. Exposure to cyanotoxins: understanding it and short term interventions to prevent it. Drinking water. *In*: Chorus, I. & M. Welker (Eds). Toxic Cyanobacteria in Water: A Guide to Their Public Health Consequences, Monitoring and Management. CRC Press, pp. 305-332.
- IBELINGS, B. W., & I. CHORUS. 2007. Accumulation of cyanobacterial toxins in freshwater "seafood" and its consequences for public health: a review. *Environmental Pollution* 150 (1): 177-192. DOI:10.1016/j.envpol.2007.04.012
- IBELINGS, B. W., A. FOSS & I. CHORUS. 2021. Exposure to cyanotoxins: understanding it and short term interventions to prevent it. Food. *In*: Chorus, I. & M. Welker (Eds). Toxic Cyanobacteria in Water: A Guide to Their Public Health Consequences, Monitoring and Management. CRC Press, pp. 368-384.
- KARJALAINEN, M., M. REINIKAINEN, L. SPOOF, J. A. MERILUOTO, K. SIVONEN & M. VIITASALO. 2005. Trophic transfer of cyanobacterial toxins from zooplankton to planktivores: consequences for pike larvae and mysid shrimps. *Environmental Toxicology* 20 (3): 354-362. DOI:10.1002/tox.20112
- KOZŁOWSKY-SUZUKI, B., A. E. WILSON, & A. DA SILVA FERRAO-FILHO. 2012. Bio-magnification or biodilution of microcystins in aquatic foodwebs? Meta-analyses of laboratory and field studies. *Harmful Algae* 18: 47-55. DOI:10.1016/j.hal.2012.04.002
- LANCE, E., A. PETIT, W. SANCHEZ, C. PATY, C. GERARD & M. BORMANS. 2014. Evidence of trophic transfer of microcystins from the gastropod *Lymnaea stagnalis* to the fish *Gasterosteus aculeatus*. *Harmful Algae* 31: 9-17. DOI:10.1016/j.hal.2013.09.006
- LAWTON, L. A., J. S. METCALF, B. ŽEGURA, R. JUNEK, M. WELKER, A. K. TÖRÖKNÉ, & L. BLÁHA. 2021. Laboratory analysis of cyanobacterial toxins and bioassays. *In*: Chorus, I. & M. Welker (Eds). Toxic Cyanobacteria in Water: A Guide to Their Public Health Consequences, Monitoring and Management. CRC Press, pp. 745-800.
- LEÓN, C., & G. A. PEÑUELA. 2019. Detected cyanotoxins by UHPLC MS/MS technique in tropical reservoirs of northeastern Colombia. *Toxicon* 167: 38-48. DOI:10.1016/j.toxicon.2019.06.010
- LOPES, T. O. M., L. S. PASSOS, L. V. VIEIRA, E. PINTO, F. DORR, R. SCHERER & L. C. GOMES. 2020. Metals, arsenic, pesticides, and microcystins in tilapia (*Oreochromis niloticus*) from aquaculture parks in Brazil. *Environmental Science and Pollution Research* 27 (16): 20187-20200. DOI:10.1007/s11356-020-08493-x
- MAGALHÃES, V. F., R. M. SOARES & S. M. AZEVEDO. 2001. Microcystin contamination in fish from the Jacarepaguá Lagoon (Rio de Janeiro, Brazil): ecological implication and human health risk. *Toxicon* 39 (7): 1077-1085. DOI:10.1016/S0041-0101(00)00251-8
- MALBROUCK, C., & P. KESTEMONT. 2006. Effects of microcystins on fish. *Environmental Toxicology and Chemistry* 25 (1): 72-86. DOI:10.1897/05-029R.1
- MARIE, B. (2020). Disentangling of the ecotoxicological signal using "omics" analyses, a lesson from the survey of the impact of cyanobacterial proliferations on fishes. *Science of The Total Environment* 736: 139701. DOI:10.1016/j.scitotenv.2020.139701
- MENDES, C. F., J. E. L. BARBOSA & J. F. NERY. 2016. Microcystin accumulation and potential depuration on muscle of fishes of fish farm: implications to public health. *International Journal of Innovative Studies in Aquatic Biology and Fisheries* 2: 1-10. DOI:10.20431/2455-7670.0202001
- METCALF, J. S., M. TISCHBEIN, P. A. COX, & E. W. STOMMEL. 2021. Cyanotoxins and the Nervous System. *Toxins* 13(9): 660. DOI:10.3390/toxins13090660
- MORALES, V.V., & R.R. MORALES. 2006. Regional review on aquaculture development. 1. Latin America and the Caribbean 2005. FAO Fisheries Circular. No. 1017/1. Rome. 177 pp
- MORANDI, M. 2015. Bioacumulación de microcistina-LR en tarariras (*Hoplias* sp.) de un embalse eutrófico (Rincón del Bonete, Uruguay) y su riesgo potencial para la salud humana. Tesis de Maestría. Universidad de la República, Uruguay. 49p.
- MOREIRA, C., V. VASCONCELOS, & A. ANTUNES. 2022. Cyanobacterial Blooms: Current Knowledge and New Perspectives. *Earth* 3(1): 127-135. DOI:10.3390/earth3010010
- MOURA, A.N, N. K. ARAGAO-TAVARES & C. A. AMORIM. 2018. Cyanobacterial blooms in freshwater bodies from a semiarid region, Northeast Brazil: A review. *Journal of Limnology* 77 (2): 179-188. DOI:10.4081/jlimnol.2018.1646
- MOWE, M. A., S. M. MITROVIC, R. P. LIM, A. FUREY & D. C. YEO. 2015. Tropical cyanobacterial blooms: a review of prevalence, problem taxa, toxins and influencing environmental factors. *Journal of Limnology* 74 (2): 205-224. DOI:10.4081/jlimnol.2014.1005
- MUNOZ, M., S. CIRÉS, Z. M. DE PEDRO, J. A. COLINA, Y. VELÁSQUEZ-FIGUEROA, J. CARMONA-JIMÉNEZ, A. CARO-BORRERO, A. SALAZAR, M. S. M. FUSTER, D. CONTRERAS, E. PERONA, A. QUESADA, & J. A. CASAS. 2021. Overview of toxic cyanobacteria and cyanotoxins in Ibero-American freshwaters: Challenges for risk management and opportunities for removal by advanced technologies. *Science of The Total Environment* 761: 143197. DOI:10.1016/j.scitotenv.2020.143197
- OLIVEIRA, N. B., C. A. SCHWARTZ, C. BLOCH, L. PAULINO & O. R. PIRES. 2013. Bioaccumulation of cyanotoxins in *Hypophthalmichthys molitrix* (silver carp) in Paranoá Lake, Brasília-DF, Brazil. *Bulletin of Environmental Contamination and Toxicology* 90 (3): 308-313. DOI:10.1007/s00128-012-0873-7
- PACHECO, F. S., M. C. S. SOARES, A. T. ASSIREU, M. P. CURTARELLI, F. ROLAND, G. ABRIL, J. L. STECH, P. C. ALVALÁ, & J. P. OMETTO. 2015. The effects of river inflow and retention time on the spatial heterogeneity of chlorophyll and water-air CO₂ fluxes in a tropical hydropower reservoir. *Biogeosciences* 12 (1): 147-162. DOI:10.5194/bg-12-147-2015
- PAERL, H. W., & J. HUISMAN. 2009. Climate change: a catalyst for global expansion of harmful cyanobacterial blooms. *Environment*

- tal Microbiology Reports*, 1 (1): 27-37. DOI:10.1111/j.1758-2229.2008.00004.x
- PÉREZ-MORALES, A., A. OLIVOS-ORTIZ, S. I. QUIJANO-SCHEGGIA, C. A. ESPINOSA-RODRÍGUEZ & M. A. JIMÉNEZ-SANTOS. 2016. Estado actual del estudio de cianobacterias dulceacuicolas formadoras de florecimientos en el centro de México. *In: García-Mendoza, E., S. I. Quijano-Scheggia, A. Olivos-Ortiz, & E. J. Núñez-Vázquez (Eds.). Florecimientos Algales Nocivos en México. CICESE*, pp. 408-421.
- PHAM, T. L., & M. UTSUMI. 2018. An overview of the accumulation of microcystins in aquatic ecosystems. *Journal of Environmental Management* 213: 520-529. DOI:10.1016/j.jenvman.2018.01.077
- RICO-MARTÍNEZ, R., A. PÉREZ-LEGASPI, G.E. SANTOS-MEDRANO, J. ALVARADO-FLORES & M.A. ARZATE-CÁRDENAS. (2017). Ecotoxicological Studies of Freshwater Ecosystems in Latin America: Diagnosis, Perspectives, and Proposals. *In: Araújo C.V.M. & C.H. Shinn (Eds). Ecotoxicology in Latin America. Nova Science Publishers*, pp. 137-161.
- ROMO, S., F. FERNÁNDEZ, Y. OUAHID & A. BARÓN-SOLA. 2012. Assessment of microcystins in lake water and fish (*Mugilidae, Liza sp.*) in the largest Spanish coastal lake. *Environmental Monitoring and Assessment* 184 (2): 939-949. DOI:10.1007/s10661-011-2011-0
- SCARLETT, K. R., S. KIM, L. M. LOVIN, S. CHATTERJEE, J. T. SCOTT, & B. W. BROOKS. 2020. Global scanning of cylindrospermopsin: Critical review and analysis of aquatic occurrence, bioaccumulation, toxicity and health hazards. *Science of The Total Environment* 738: 139807. DOI:10.1016/j.scitotenv.2020.139807
- SINGH, N. K., & D. W. DHAR. 2013. Cyanotoxins, related health hazards on animals and their management: A review. *The Indian Journal of Animal Sciences* 83: 1111-1127. <https://epubs.icar.org.in/index.php/IJAnS/article/view/34749>
- SALOMÓN, S., C. A. RIVERA-RONDÓN, & Á. M. ZAPATA. 2020. Cyanobacterial blooms in Colombia: State of knowledge and research needs in the context of climate global change. *Revista de la Academia Colombiana de Ciencias Exactas, Físicas y Naturales* 44 (171): 376-391. DOI:10.18257/raccefyn.1050
- SOTTON, B., J. GUILLARD, O. ANNEVILLE, M. MARÉCHAL, O. SAVICHTCHEVA, & I. DOMAZON. 2014. Trophic transfer of microcystins through the lake pelagic food web: evidence for the role of zooplankton as a vector in fish contamination. *Science of the Total Environment* 466: 152-163. DOI:10.1016/j.scitotenv.2013.07.020
- SUNESSEN, I., S. M. MÉNDEZ, J. E. MANCERA-PINEDA, M. Y. D. BOTTEIN & H. ENEVOLDSEN. 2021. The Latin America and Caribbean HAB status report based on OBIS and HAEDAT maps and databases. *Harmful Algae* 102: 101920. DOI:10.1016/j.hal.2020.101920
- SVIRČEV, Z., D. LALIĆ, G. B. SAVIĆ, N. TOKODI, D. D. BACKOVIĆ, L. CHEN, J. MERILUOTO, & G. A. CODD. 2019. Global geographical and historical overview of cyanotoxin distribution and cyanobacterial poisonings. *Archives of Toxicology* 93 (9): 2429-2481. DOI:10.1007/s00204-019-02524-4
- TESTAI, E., F. M. BURATTI, E. FUNARI, M. MANGANELLI, S. VICHI, N. ARNICH, R. BIRÉ, V. FESSARD & A. SIALEHAAMO. 2016. Review and analysis of occurrence, exposure and toxicity of cyanobacteria toxins in food. *EFSA Supporting Publications* 13(2): 998E. DOI:10.2903/sp.efsa.2016.EN-998
- TILLMANN, A. R., A. E. WILSON, F. R. PICK & O. SARNELLE. 2008. Meta-analysis of cyanobacterial effects on zooplankton population growth rate: species-specific responses. *Fundamental and Applied Limnology* 171 (4): 285. DOI:10.1127/1863-9135/2008/0171-0285
- VALDOR, R., & M. ABOAL. 2007. Effects of living cyanobacteria, cyanobacterial extracts and pure microcystins on growth and ultrastructure of microalgae and bacteria. *Toxicon* 49 (6): 769-779. DOI:10.1016/j.toxicon.2006.11.025
- VASCONCELOS, V., A. MARTINS, M. VALE, A. ANTUNES, J. AZEVEDO, M. WELKER, & G. MONTEJANO. 2010. First report on the occurrence of microcystins in planktonic cyanobacteria from Central Mexico. *Toxicon* 56 (3): 425-431. DOI:10.1016/j.toxicon.2010.04.011
- VASCONCELOS, J. F., J. E. L. BARBOSA, W. LIRA, & S. M. F. O. AZEVEDO. 2013. Microcystin bioaccumulation can cause potential mutagenic effects in farm fish. *The Egyptian Journal of Aquatic Research* 39(3): 185-192. DOI:10.1016/j.ejar.2013.11.002
- WOOD, R. 2016. Acute animal and human poisonings from cyanotoxin exposure—A review of the literature. *Environment International* 91: 276-282. DOI:10.1016/j.envint.2016.02.026
- ZAMORA-BARRIOS, C. A., S. NANDINI & S. S. SARMA. 2019. Bioaccumulation of microcystins in seston, zooplankton and fish: A case study in Lake Zumpango, Mexico. *Environmental Pollution* 249: 267-276. DOI:10.1016/j.envpol.2019.03.029
- ZHANG, D., P. XIE, Y. LIU & T. QIU. 2009. Transfer, distribution and bioaccumulation of microcystins in the aquatic food web in Lake Taihu, China, with potential risks to human health. *Science of the Total Environment* 407 (7): 2191-2199. DOI:10.1016/j.scitotenv.2008.12.039
- ZHANG, Y., J. K. WHALEN, & S. SAUVÉ. 2021. Phytotoxicity and bioconcentration of microcystins in agricultural plants: Meta-analysis and risk assessment. *Environmental Pollution* 272: 115966. DOI:10.1016/j.envpol.2020.115966
- ZURAWELL, R. W., H. CHEN, J. M. BURKE, & E. E. PREPAS. 2005. Hepatotoxic cyanobacteria: a review of the biological importance of microcystins in freshwater environments. *Journal of Toxicology and Environmental Health, Part B* 8 (1): 1-37. DOI:10.1080/10937400590889412