

## Effect of tanniferous plants on *in vitro* digestion and methane production

### Efecto de plantas taníferas en la digestión y producción de metano *in vitro*

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**ABSTRACT.** It has been shown that tannins in the foliage of trees and shrubs can reduce methane (CH<sub>4</sub>) production in the rumen. The aim of the present paper was to evaluate the potential of five tanniferous browse plant species, native to temperate central Mexico, to reduce *in vitro* methane production. The *in vitro* gas production technique and gas chromatography were used to evaluate the effect of the inclusion of these plants at three inclusion levels (10, 20 and 30 %) in an experimental diet. A factorial experimental arrangement with the five plant species and four levels of inclusion was used. Gas production was monitored throughout 72 h of incubation and methane production was recorded after 24 h. The results showed that a significant decrease in methane production was observed with all plants at 10 % inclusion level (P < 0.001), and that the highest reduction (> 26 %) was observed with *Amaranthus spinosus* and *Commelina coelestis* at 20 % and 30 % inclusion respectively in comparison to the control diet. However, the digestibility of the experimental diet declined at these inclusion levels. It was concluded that all the tested plants reduced methane production at all inclusion levels; however, the best tradeoff between methane reduction and tannins' associated negative effects on diet digestibility was achieved at the 10 % inclusion level.

**Key words:** Plant phenolics, tannins, *in vitro* gas production, methane emission

**RESUMEN.** Se ha demostrado que los taninos en el follaje de los árboles y arbustos pueden reducir la producción de metano (CH<sub>4</sub>) en el rumen. El objetivo del estudio fue evaluar el potencial de cinco especies de plantas arbustivas, nativas de la región de clima templado del centro de México, para reducir la producción *in vitro* de metano. Los follajes fueron adicionados a 0, 10, 20 y 30 % en base seca a una dieta experimental en un diseño factorial de cinco plantas por cuatro niveles de inclusión. La producción de gas *in vitro* se monitoreó a lo largo de 72 h de incubación y la producción de metano se registró después de 24 h. Los resultados mostraron una disminución significativa en la producción de metano con todas las especies de plantas a un nivel de inclusión de 10 % (P < 0.001), mientras que la reducción más alta (> 26 %) se observó con *Amaranthus spinosus* y *Commelina coelestis* a niveles de inclusión de 20 % y 30 %, respectivamente, en comparación con la dieta control. Sin embargo, la digestibilidad de la dieta experimental disminuyó al aumentar los niveles de inclusión. Se concluyó que las plantas evaluadas en todos los niveles de inclusión redujeron la producción de metano, sin embargo el mejor balance entre la reducción en la producción de metano y los efectos negativos sobre la degradación de la dieta asociados a los taninos se alcanzó a un nivel de inclusión del 10 %.

**Palabras clave:** Fenoles; taninos, producción de gas *in vitro*, Metano, Nutrición

## INTRODUCTION

The accumulation of greenhouse gases (GHG) such as (CO<sub>2</sub>), methane (CH<sub>4</sub>) and nitrous oxide (N<sub>2</sub>O) in the Earth's atmosphere has led to predictions that global surface temperatures will increase between one and six degrees Celsius during the 21st century (IPCC 2007). According to Arbre *et al.* (2016), the contribution of livestock to anthropogenic GHG emissions is estimated at 14.5 % of total emissions, of which 39 % is due to methane (CH<sub>4</sub>) produced by enteric fermentation in ruminants. Approximately 6 % of cattle dietary gross energy intake is lost to the atmosphere in the form of methane (De Ramus *et al.* 2003). Hence, mitigating methane emissions could improve sustainability and profitability of cattle enterprises (Vanlierde *et al.* 2016). Historically, the inhibition of enteric methanogenesis has been approached in regard to livestock nutrition. More recently the reduction of CH<sub>4</sub> formation has garnered interest in relation to reducing GHG emissions. Previous studies have reported that CH<sub>4</sub> production can be reduced by the addition of plant extracts containing tannins to livestock feed (Patra *et al.* 2006, Patra *et al.* 2010). A meta-analysis by Jayanegara *et al.* (2012) showed that condensed and hydrolysable tannins may reduce CH<sub>4</sub> production. Tannins may directly affect methanogenesis by inhibiting the growth, development and activity cellulase enzyme of methanogens, or indirectly by reducing the number of protozoa associated with the methanogens (Makkar *et al.* 1993). Tanniferous plants can represent a sustainable and environmentally friendly option for mitigation; however, significant screening work is needed in order to find the most efficient species. These species have to meet two criteria, one is to reduce CH<sub>4</sub> formation in the rumen and the other is to not diminish animal performance through modification of feedstuffs' fermentation and digestibility in the rumen. *In vivo* evaluation of different species can be expensive and not feasible due to the large number of plants that need to be evaluated. Therefore, *in vitro* gas production can be used as a screening test to select the species with

the greatest potential before *in vivo* evaluation. The objective of the study was to evaluate the potential use of five Mexican tanniferous shrub plants to reduce methane emission and measure the effect on digestibility of the dry matter, organic matter and neutral detergent fiber in the diet, using the *in vitro* gas production technique and gas chromatography.

## MATERIALS AND METHODS

The study was conducted at the animal nutrition laboratory of the Faculty of Veterinary Medicine of the Autonomous University of the State of Mexico, and complied with the ethical standards set by the institution.

### Sample collection and preparation

The browse plant species used in the study were *Amaranthus spinosus*, *Cosmos bipinnatus*, *Commelina coelestis*, *Eupatorium glabratum* and *Galinsoga parviflora*. Plant samples were collected from six different locations randomly selected in the temperate region of the State of Mexico, located at 19° 17' 32" N and 99° 39' 14" W and at 2663 masl. For each species, six kilograms of fresh plant matter (one for each location) were collected and cut into small pieces (3-5 cm). Samples were then dried at 50°C for 48 h using a forced air oven in order to prevent enzymatic degradation of the phenolic compounds present in the plant matter (Makkar *et al.* 1993). Once dry, 400 g of plant matter were ground in a Lab-Willey Grinder (code MSW-342-IN; 10122740) and sieved through a 2-mm screen. The grounded material was mixed well and then 100 g were sub-sampled, reground and passed through a 0.5-mm screen sieve. These finely ground sub-samples were used in tannin analysis while the rest of the material was used for *in vitro* gas production analysis.

### Treatments

The experimental diet (control) was composed of 64.6 % ryegrass (*Lolium perenne*), 20.8 % corn grain (*Zea mays*), 8.3 % canola meal (*Brassica napus*) and 6.3 % corn stover. In the experimen-

tal treatments 0, 10, 20 and 30 % ryegrass was replaced by equivalent amounts of the test plants; thus, 20 treatments were tested.

### Chemical composition of plants

Dry matter (DM) was determined by drying the samples at 135 °C for 2 h. Organic matter (OM) was calculated as weight lost at sample ignition at 600 °C (AOAC 1995). Neutral detergent fiber (NDF) and acid detergent fiber (ADF) were determined using the ANKOM fiber technology technique (Robinson *et al.* 1999) without using alpha amylase. Crude protein (CP) was determined by the Kjeldahl method (AOAC 1995).

### *In vitro* gas production

Gas production (GP) was determined following the Theodorou *et al.* (1994) technique. Rumen fluid was collected before the morning feeding from two ruminally-fistulated, non-lactating and non-gravid Holstein cows. All cows had access to fresh water at all times, were fed the same experimental diet, and grazed on a ryegrass sward from 6:00 until 16:00 h daily. Samples weighing 0.999 g of the control and experimental diets were placed in 125-ml serum glass bottles, and approximately 90 ml of buffered rumen fluid was added to each bottle. Once closed, the bottles were gently shaken and placed in a water bath at 39 °C. Six independent runs were completed. The first three were incubated for 24 h in order to simulate normal residence time of feed in the rumen, and were used to measure methane production at 24 h post-incubation. The other three runs were incubated for 72 h. A total of 288 bottles were used and all incubations were completed in triplicate within the run. Gas production measurements, for the samples incubated 72 h, were taken hourly (up to 8 h post-incubation), then every four (12- 28), eight (36-60) and 72 h post-incubation. After 24 and 72 h, the incubation residue was analyzed for digestibility of dry matter (DMD), organic matter (OMD) and neutral-detergent fiber (DNDF) content using the ANKOM fiber technology technique (Robinson *et al.* 1999).

### Methane measurement

Using a gas-tight syringe, gas samples were collected from each bottle at 24 h post-incubation as in Bhatta *et al.* (2015) and Kaya *et al.* (2016). After the volume of gas was recorded, and the sample removed for methane analysis, the remaining gas was released. CH<sub>4</sub> content was determined by injecting 1 ml of gas into a Perkin Elmer gas chromatograph (model: Clarus 500 series) equipped with a flame ionized detector (FID). Separation was achieved using an Elite-Q Plot Capillary Column (Perkin Elmer) packed with a 60/80 mesh carboxenTM-1000 stationary phase. Nitrogen was used as the carrier gas with a flow rate of 30 mL<sup>-1</sup>min, an isothermal oven temperature of 50 °C, and an injector temperature of 250 °C. The calibration curve using a regression equation was completed with standard CH<sub>4</sub> (99.99 % from ALTECH).

### Extraction of tannins and measurement of total phenolics and tannins content

Phenols content was determined using the Folin-Ciocalteu method and tannins were measured using polyvinylpolypyrrolidone (PVPP) as described by Makkar *et al.* (1993).

### Experimental design and analysis of results

A completely randomized design with a 5 x 4 factorial arrangement was used in which both the plants used and their inclusion levels are treatments. The general linear model was:

$$Y_{ij} = \mu + T_i + B_j + (T_i * B_j) + e_{ij}$$

Where: Y<sub>ij</sub>= independent variable, μ= overall mean, T<sub>i</sub>= effect of plant (i=1...5), B<sub>j</sub>= effect of the level of inclusion (j=1...4), (T<sub>i</sub> \* B<sub>j</sub>)= Effect of the interaction between plant species and the level of inclusion and e<sub>ij</sub>= True error in the i level inclusion of j treatment.

Results were analyzed with analysis of variance using the general linear model command of Minitab (2013) for a factorial experimental design; where significant differences between species and

**Table 1.** Chemical composition, Phenols and tannins content of the experimental diet and plant species (g kg<sup>-1</sup> DM).

Feed	Dry matter	Organic matter	Crude protein	NDF	ADF	Total phenols N	on-tannins phenols	Total tannins	Condensed tannins
Diet	909 <sup>bd</sup>	810 <sup>d</sup>	160 <sup>e</sup>	439 <sup>a</sup>	181 <sup>e</sup>	34.4 <sup>f</sup>	11.2 <sup>a</sup>	22.2 <sup>c</sup>	1.19 <sup>d</sup>
<i>Amaranthus spinosus</i>	930 <sup>a</sup>	760 <sup>a</sup>	184 <sup>a</sup>	448 <sup>a</sup>	220 <sup>a</sup>	48.4 <sup>a</sup>	10.9 <sup>a</sup>	37.5 <sup>a</sup>	5.5 <sup>a</sup>
<i>Cosmos bipinnatus</i>	901 <sup>cb</sup>	810 <sup>b</sup>	84 <sup>b</sup>	481 <sup>b</sup>	327 <sup>b</sup>	90.7 <sup>b</sup>	19.0 <sup>b</sup>	71.7 <sup>b</sup>	8.4 <sup>b</sup>
<i>Commelina coelestis</i>	926 <sup>ad</sup>	780 <sup>c</sup>	93 <sup>c</sup>	550 <sup>c</sup>	360 <sup>c</sup>	32.3 <sup>c</sup>	11.2 <sup>a</sup>	21.1 <sup>c</sup>	5.4 <sup>a</sup>
<i>Ebatorium glabratium</i>	928 <sup>a</sup>	824 <sup>b</sup>	100 <sup>c</sup>	370 <sup>d</sup>	209 <sup>a</sup>	11.2 <sup>d</sup>	11.3 <sup>a</sup>	18.2 <sup>d</sup>	0.6 <sup>c</sup>
<i>Galinosa parviflora</i>	918 <sup>ac</sup>	809 <sup>bd</sup>	109 <sup>d</sup>	488 <sup>b</sup>	306 <sup>d</sup>	11.3 <sup>e</sup>	17.3 <sup>b</sup>	43.2 <sup>e</sup>	1.0 <sup>d</sup>

NDF = neutral detergent fiber, ADF = Acid detergent fiber, <sup>a,b,c,d,e,f</sup> Different superscripts following means between plants species' in the row indicate differences at  $p < 0.05$ .

level were found, a Tukey test was applied. Individual analysis of variance for level within species was also conducted using a completely randomized design for a significance level of  $p < 0.05$ ; where significant differences between levels were found, a Tukey test was also applied. The GP curves were not fitted to mathematical models in order to determine fermentation kinetics because the aim of the work was not to describe the effect of tanniferous plants on diet's kinetics, but their effect on reducing methane production in the rumen. We used a simpler, original and intuitive approach; we wanted to know the volume of gas produced from the feed sample in order to have an indicator of the extent of digestion (fermentation) of the diet in the rumen (bottles). The results were analyzed by ANOVA as described above.

## RESULTS

**Chemical composition and polyphenols contents:** Table 1 shows the chemical composition of the experimental diet and plant species. The highest CP content was observed in *A. spinosus* ( $p < 0.05$ ), followed by *G. parviflora*. It can also be observed that the NDF content for all analyzed plant species is moderate. Table 1 also shows the content (g kg<sup>-1</sup> DM) of total phenols, non-tannin phenols, total tannins and condensed tannins for all the experimental plants used. It can be observed that the highest concentration of total tannins ( $p < 0.05$ ) was in *C. bipinnatus*, and the lowest in *E. glabratium*. The highest concentration of condensed tannins ( $p < 0.05$ ) was also observed in *C.*

*bipinnatus*, followed by *A. spinosus* and *C. coelestis*, both with the same content.

**Gas and methane production:** Table 2 shows the CH<sub>4</sub> production after 24 h of fermentation and the cumulative volume of gas produced at 24 and 72 h of fermentation for the five species at the different inclusion levels. The inclusion of all plant species ( $p < 0.001$ ) at all inclusion levels resulted in a significant reduction ( $p < 0.001$ ) of methane produced. Individual analysis by species shows that the highest reduction in methane production (26.2 %) was observed when *C. coelestis* was included at a level of 30 % ( $p < 0.05$ ) followed by including *A. spinosus* at 20 % (26 % less CH<sub>4</sub> produced). The other three plants also significantly reduced ( $p < 0.05$ ) CH<sub>4</sub> production by 20 % when included at 30 % level. Further, *G. parviflora* and *A. spinosus* reduced methane production by 15 % and 17 %, respectively, when included at 10% level.

The cumulative gas produced after 72 h of incubation for all five species at all inclusion levels followed the same trend as did the gas produced at 24 h of incubation; for example, the highest volume of gas produced was observed with *C. coelestis* at 20 % inclusion level ( $p < 0.05$ ) for both incubation periods. The inclusion of *A. spinosus*, *C. bipinnatus* and *G. parviflora* at 30 % level and at 24 h of incubation did not reduce gas production ( $p < 0.05$ ). On the contrary, *C. coelestis* and *E. Glabratium* reduced ( $p < 0.05$ ) gas production at the same level of inclusion. The overall statistical analysis column in Table 2 shows that the inclusion levels of all plant species did not significantly reduce gas

**Table 2.** Effect of the inclusion of five tanniferous plant species on methane production at 24 h and cumulative gas production at 24 and 72 h of incubation.

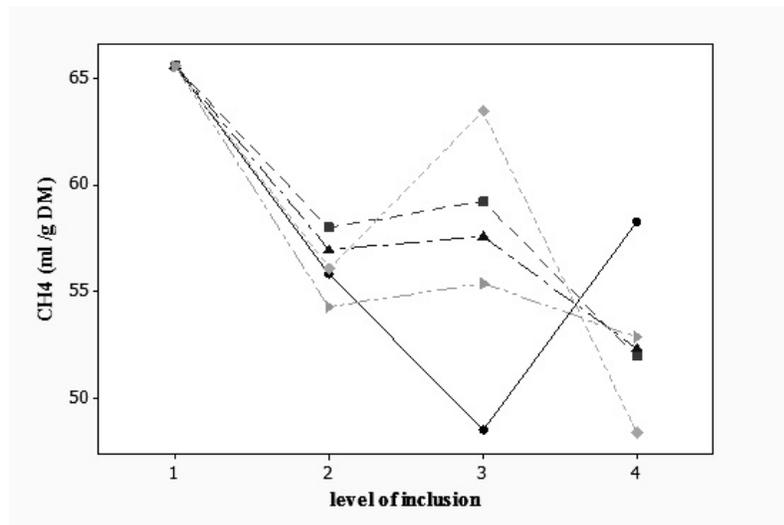
Inclusion level	Species					$\bar{x}$ for level	P value		
	<i>Amaranthus spinosus</i>	<i>Cosmos bipinnatus</i>	<i>Commelina coelestis</i>	<i>Ebatorium glabratium</i>	<i>Galinsoga parviflora</i>		Species	Level	Interaction
Methane production (ml of CH <sub>4</sub> g <sup>-1</sup> DM)									
0 %	65.6 <sup>a</sup>	65.6 <sup>a</sup>	65.6 <sup>a</sup>	65.6 <sup>a</sup>	65.6 <sup>a</sup>	65.6 <sup>a</sup>	0.001	0.002	0.008
10 %	55.8 <sup>b</sup>	58.0 <sup>b</sup>	56.1 <sup>b</sup>	56.9 <sup>b</sup>	54.3 <sup>b</sup>	56.2 <sup>b</sup>			
20 %	48.5 <sup>c</sup>	59.2 <sup>b</sup>	63.5 <sup>a</sup>	57.6 <sup>b</sup>	55.4 <sup>bc</sup>	56.8 <sup>b</sup>			
30 %	58.3 <sup>b</sup>	52.0 <sup>c</sup>	48.4 <sup>c</sup>	52.3 <sup>c</sup>	52.9 <sup>b</sup>	52.7 <sup>c</sup>			
$\bar{x}$ for species SED	57.0 <sup>a</sup>	58.7 <sup>b</sup>	58.4 <sup>b</sup>	58.1 <sup>b</sup>	57.0 <sup>a</sup>				
Gas production at 24 h(ml g <sup>-1</sup> DM)									
0 %	177.9 <sup>a</sup>	177.9 <sup>a</sup>	177.9 <sup>a</sup>	177.9 <sup>a</sup>	177.9 <sup>a</sup>	177.9 <sup>a</sup>	0.001	0.072	0.001
10 %	170.2 <sup>a</sup>	180.1 <sup>a</sup>	186.3 <sup>ab</sup>	168.5 <sup>b</sup>	183.2 <sup>ab</sup>	177.6 <sup>a</sup>			
20 %	149.8 <sup>b</sup>	183.4 <sup>ab</sup>	186.7 <sup>b</sup>	170.5 <sup>bc</sup>	181.4 <sup>a</sup>	174.3 <sup>a</sup>			
30 %	177.9 <sup>a</sup>	164.4 <sup>ac</sup>	160.9 <sup>c</sup>	163.9 <sup>b</sup>	175.8 <sup>ad</sup>	168.5 <sup>b</sup>			
$\bar{x}$ for species SED	169.0 <sup>a</sup>	176.4 <sup>b</sup>	178.0 <sup>c</sup>	170.2 <sup>a</sup>	179.5 <sup>dc</sup>				
Gas production at 72 h(ml g <sup>-1</sup> DM)									
0 %	265.1 <sup>a</sup>	265.1 <sup>a</sup>	265.1 <sup>a</sup>	265.1 <sup>a</sup>	265.1 <sup>a</sup>	265.0 <sup>a</sup>	0.001	0.067	0.001
10 %	246.9 <sup>b</sup>	258.4 <sup>b</sup>	270.9 <sup>a</sup>	249.7 <sup>b</sup>	256.5 <sup>d</sup>	256.4 <sup>b</sup>			
20 %	221.8 <sup>c</sup>	264.5 <sup>a</sup>	276.7 <sup>a</sup>	250.1 <sup>b</sup>	251.7 <sup>bd</sup>	253.0 <sup>cb</sup>			
30 %	268.4 <sup>a</sup>	250.8 <sup>c</sup>	249.0 <sup>b</sup>	241.6 <sup>c</sup>	248.0 <sup>cd</sup>	251.5 <sup>db</sup>			
$\bar{x}$ for species SED	250.5 <sup>a</sup>	259.6 <sup>b</sup>	265.4 <sup>c</sup>	251.6 <sup>a</sup>	255.3 <sup>a</sup>				

SED= standard error of the difference, <sup>a,b,c,d</sup> Different superscripts following means within plant species' inclusion levels in the column indicate differences at  $p < 0.05$ .

production for both incubation periods,  $p > 0.07$  and  $p > 0.06$ , respectively. By contrast, significant differences were observed between species ( $p < 0.001$ ), with the largest gas production volumes being observed in *C. coelestis* and *G. parviflora*.

**Effect on the digestibility of the diet:** Table 3 shows the digestibility (g kg<sup>-1</sup> DM) of the DM, OM and NDF at 24 h of incubation for all species at all inclusion levels. The DMD and the OMD of the experimental diet was reduced by all species at all inclusion levels ( $p < 0.001$ ) except for *G. parviflora* at 30 % inclusion where no change was observed in DMD ( $p > 0.05$ ). However, OMD and DNDF were not affected when using *C. bipinnatus* at 20 % inclusion level ( $p > 0.05$ ) (Table 3). This may be an important indicator that this plant can decrease methane production *in vivo* but not the overall digestive ability of cattle. In contrast, *C. coelestis* significantly reduced DMD, OMD and DNDF at 30 % inclusion level, which was accompanied by the highest reduction of methane production in the experimental diet (Table 2). The DNDF increased by

the inclusion of all plant species at inclusion level of 10 % at 24 h of incubation but at the 30 % level it decreased DNDF ( $p < 0.001$ ). Table 4 shows the digestibility (g kg<sup>-1</sup> DM) of the DM, OM and NDF after 72 h of incubation for all species at all inclusion levels. A trend similar to that in Table 3 was observed, since the inclusion of all plants at all levels resulted in a significant reduction of the DNDF ( $p < 0.001$ ). Figure 1 shows the effect of interaction between plant species and the inclusion levels on methane production at 24 h ( $p < 0.001$ ); it can be observed that the response is not linear for all the species because a quadratic effect was observed for *A. spinosus*, *C. bipinnatus* and *C. coelestis*. All plants reduced methane production at 10 % inclusion but at 20 % only *Amaranthus spinosus*, *Cosmos bipinnatus*, *Eupatorium glabratum* and *Galinsoga parviflora* kept methane production low. However, a significant interaction can be observed at level three for *A. spinosus* ( $p < 0.001$ ), *C. bipinnatus* ( $p < 0.01$ ) and *C. coelestis* ( $p < 0.001$ ) because lines of these species are not parallel implying that the effect of the species upon CH<sub>4</sub> production depends



**Figure 1.** Interaction plot for methane production at 24 h of incubation for the five tanniferous species at the different inclusion levels.

**Table 3.** Effect of the inclusion of five tanniferous plant species on the digestibility of dry matter, organic matter and NDF after 24 h of incubation ( $\text{g kg}^{-1}$  DM).

Inclusion level	Plant Species					$\bar{x}$ for level	P value for Species	Level	Interaction
	<i>Amaranthus spinosus</i>	<i>Cosmos bipinnatus</i>	<i>Commelina coelestis</i>	<i>Ebatorium glabratium</i>	<i>Galinsoga parviflora</i>				
Dry matter digestibility									
0 %	739 <sup>a</sup>	739 <sup>a</sup>	739 <sup>a</sup>	739 <sup>a</sup>	739 <sup>a</sup>	739.3 <sup>a</sup>	0.001	0.001	0.001
10 %	673 <sup>b</sup>	683 <sup>b</sup>	719 <sup>b</sup>	701 <sup>b</sup>	628 <sup>b</sup>	681.0 <sup>b</sup>			
20 %	686 <sup>c</sup>	732 <sup>c</sup>	726 <sup>c</sup>	717 <sup>c</sup>	680 <sup>d</sup>	708.0 <sup>c</sup>			
30 %	723 <sup>d</sup>	649 <sup>d</sup>	621 <sup>d</sup>	705 <sup>b</sup>	744 <sup>a</sup>	688.4 <sup>d</sup>			
$\bar{x}$ for species	705.3 <sup>a</sup>	700.7 <sup>b</sup>	701.2 <sup>cb</sup>	715.5 <sup>d</sup>	698.0 <sup>eb</sup>				
SED							1.4	1.3	
Organic matter digestibility									
0 %	742 <sup>a</sup>	742 <sup>a</sup>	742 <sup>a</sup>	742 <sup>a</sup>	742 <sup>a</sup>	742.4 <sup>a</sup>	0.001	0.001	0.001
10 %	679 <sup>b</sup>	693 <sup>b</sup>	728 <sup>b</sup>	710 <sup>b</sup>	638 <sup>b</sup>	689.5 <sup>b</sup>			
20 %	690 <sup>c</sup>	740 <sup>a</sup>	731 <sup>b</sup>	729 <sup>ac</sup>	692 <sup>c</sup>	716.5 <sup>c</sup>			
30 %	728 <sup>d</sup>	659 <sup>d</sup>	631 <sup>c</sup>	716 <sup>bc</sup>	753 <sup>d</sup>	697.4 <sup>d</sup>			
$\bar{x}$ for species	710.0 <sup>a</sup>	708.6 <sup>a</sup>	708.0 <sup>a</sup>	724.5 <sup>b</sup>	706.2 <sup>a</sup>				
SED							1.6	1.4	
Digestibility of Neutral detergent fiber									
0 %	657 <sup>a</sup>	657 <sup>a</sup>	657 <sup>a</sup>	657 <sup>a</sup>	657 <sup>a</sup>	657.1 <sup>a</sup>	0.001	0.001	0.001
10 %	680 <sup>b</sup>	725 <sup>b</sup>	655 <sup>a</sup>	665 <sup>a</sup>	715 <sup>b</sup>	688.0 <sup>b</sup>			
20 %	656 <sup>a</sup>	686 <sup>ac</sup>	680 <sup>b</sup>	634 <sup>c</sup>	652 <sup>ab</sup>	661.5 <sup>a</sup>			
30 %	643 <sup>a</sup>	651 <sup>a</sup>	624 <sup>c</sup>	582 <sup>b</sup>	662 <sup>ab</sup>	632.3 <sup>c</sup>			
$\bar{x}$ for species	659.0 <sup>a</sup>	679.5 <sup>b</sup>	654.0 <sup>a</sup>	634.6 <sup>c</sup>	671.3 <sup>ab</sup>				

SED is standard error of difference, <sup>a,b,c,d</sup> Different superscripts following means within plant species' inclusion levels in the column indicate differences at  $p < 0.05$ .

up on the inclusion level. Moreover, the inclusion level effect explains more than two and a half times the total variance explained by the species.

## DISCUSSION

### Polyphenols content

In the present study, we found significantly lower levels of methane released with increasing levels of most tannin-rich plants added to the ex-

**Table 4.** Effect of the inclusion levels of the five plant species on digestibility of dry matter, organic matter and NDF after 72 h of incubation (g kg<sup>-1</sup> DM).

Inclusion level	Plant Species					$\bar{x}$ for level	P value for		
	<i>Amaranthus spinosus</i>	<i>Cosmos bipinnatus</i>	<i>Commelina coelestis</i>	<i>Ebatorium glabratium</i>	<i>Galinsoga parviflora</i>		Species	Level	Interaction
Dry matter digestibility									
0 %	814	814 <sup>a</sup>	814 <sup>a</sup>	814 <sup>a</sup>	814 <sup>a</sup>	814.0 <sup>a</sup>	0.001	0.001	0.001
10 %	823	794 <sup>b</sup>	793 <sup>b</sup>	769 <sup>b</sup>	839 <sup>b</sup>	804.0 <sup>b</sup>			
20 %	812	817 <sup>a</sup>	799 <sup>ab</sup>	794 <sup>ab</sup>	839 <sup>b</sup>	812.6 <sup>a</sup>			
30 %	808	788 <sup>b</sup>	766 <sup>c</sup>	765 <sup>b</sup>	819 <sup>ab</sup>	789.3 <sup>c</sup>			
$\bar{x}$ for species	814.0 <sup>a</sup>	803.6 <sup>a</sup>	793.0 <sup>b</sup>	786.0 <sup>cb</sup>	828.0 <sup>d</sup>				
SED							3.9	3.6	
Organic matter digestibility									
0 %	821 <sup>a</sup>	821 <sup>a</sup>	821 <sup>a</sup>	821 <sup>a</sup>	821 <sup>a</sup>	820 <sup>a</sup>	0.001	0.001	0.001
10 %	843 <sup>b</sup>	799 <sup>b</sup>	797 <sup>b</sup>	778 <sup>a</sup>	847 <sup>b</sup>	813 <sup>a</sup>			
20 %	824 <sup>a</sup>	821 <sup>a</sup>	801 <sup>ab</sup>	808 <sup>a</sup>	847 <sup>b</sup>	820 <sup>a</sup>			
30 %	821 <sup>a</sup>	799 <sup>b</sup>	774 <sup>c</sup>	773 <sup>b</sup>	833 <sup>ab</sup>	800 <sup>b</sup>			
$\bar{x}$ for species	827.0 <sup>a</sup>	810.0 <sup>b</sup>	798.3 <sup>cb</sup>	795.0 <sup>dc</sup>	837.0 <sup>a</sup>				
SED							4.9	4.3	
Digestibility of Neutral detergent fiber									
0 %	755 <sup>a</sup>	755 <sup>a</sup>	755 <sup>a</sup>	755 <sup>a</sup>	755 <sup>a</sup>	754.7 <sup>a</sup>	0.001	0.001	0.001
10 %	779 <sup>b</sup>	728 <sup>b</sup>	726 <sup>b</sup>	698 <sup>b</sup>	774 <sup>ab</sup>	740.7 <sup>b</sup>			
20 %	704 <sup>c</sup>	715 <sup>b</sup>	719 <sup>b</sup>	667 <sup>c</sup>	748 <sup>a</sup>	710.5 <sup>c</sup>			
30 %	685 <sup>c</sup>	658 <sup>c</sup>	674 <sup>c</sup>	579 <sup>d</sup>	742 <sup>ac</sup>	667.6 <sup>d</sup>			
$\bar{x}$ for species	730.6 <sup>a</sup>	714.0 <sup>b</sup>	718.2 <sup>cb</sup>	674.7 <sup>d</sup>	754.3 <sup>e</sup>				

SED is standard error of difference, a,b,c,d Different superscripts following means within plant species' inclusion levels in the column indicate differences at  $p < 0.05$ .

perimental diet. Tannins were frequently observed to reduce structural carbohydrate digestibility by decreasing the number of cellulolytic microbes in rumen fluid (Singleton 1981), inhibiting cellulase enzyme activity (Leinmüller *et al.* 1991, Makkar 1993), preventing adhesion of microbes onto food particles (Leinmüller and Menke 1990) and/or suppressing degradation by the formation of complexes with cellulose (McSweeney *et al.* 2001). However, no information is given by these authors as to whether there is a difference between the effects of hydrolysable and condensed tannins, but Leinmüller *et al.* (1991) considered hydrolysable tannins as being less adverse in this respect. It is worth mentioning that the effect of tannins on DNDF was not the same for all species at 10 % inclusion level (Table 3); for example, the DNDF for *A. spinosus*, and *G. parviflora* increased at this level. This suggests that at low inclusion levels these plants may reduce methane production without significantly affecting the DNDF.

### Methane production

Results of the present study show that methane was consistently reduced by all five tanniferous plants (Table 2), and that the highest reduction was observed by the inclusion of *C. coelestis* and *A. spinosus* at the 30 and 20 % inclusion levels respectively. The same effect of tannin-rich plants was observed by Hatew *et al.* (2016) and Rodríguez *et al.* (2011) who reported reduced methane production with the addition of plant extract containing tannins. The concentration of condensed tannins in both plants species may have also exerted a negative effect on methane production because *C. coelestis* and *A. spinosus* showed the second highest concentrations of condensed tannins just behind *C. bipinnatus*. These results are in line with Tavendale *et al.* (2005) and Gameda and Hassen (2015) who demonstrated for the first time that condensed tannins can inhibit methane production by inhibiting the growth of rumen methanogens and by their indirect effects via reduced hydrogen production (possible by reducing fiber digestibility). In the present study, the largest

reduction in methane production observed with the inclusion of *C. coelestis* was accompanied by a significant reduction in the DNDf at 30 % inclusion level (Tables 2 and 3), which confirms the Gemeda and Hassen (2015) hypothesis. Our results are also in line with Tan *et al.* (2011) who found that a relatively low level of 15 mg of condensed tannins/500 mg of DM reduced methane production by 47 % with only 7 % reduction in digestibility of feed DM. No information was found in the literature reviewed on the effect of the plants analysed in the present work on methane production, so no further discussion was possible.

### Gas production

Gas production is generally recognized as a more sensitive approach for detection of differences in substrate fermentation using *in vitro* systems than DM disappearance (Xu *et al.* 2010). Our results show that GP was negatively affected by some levels of inclusion of tanniferous plants in the experimental diet (Table 2). This effect could be attributed to the antimicrobial properties of tannins and phenolic compounds in the experimental plants, which in turn cause a reduction in gas production (Francis *et al.* 2002). However, the least severe effect on GP was observed at 10 % inclusion level suggesting that the best tradeoff between methane reduction and reduction of diet digestibility can be achieved at low inclusion levels of these plants.

### Interaction plant species x inclusion rate

The significant interaction observed between inclusion level and plant species for methane and gas production indicates that the behaviors of both variables are the result of the combined effect between the plant species and, particularly, the inclusion level of it in the diet. This effect is

clearly seen in the case of *C. bipinnatus* and *C. coelestis*, where at low inclusion level the effect on methane production is moderate; however, at the highest inclusion rate methane production is drastically reduced due to the effect of tannins, especially condensed ones since both plants have a high content of these secondary metabolites. Unfortunately, methane reduction was accompanied by a significant reduction in DMD and OMD, which may prevent the inclusion of these plants at high rates in *in vivo* experiments.

### CONCLUSION

It is concluded that all the tested plants reduced methane production at all inclusion levels; however, the best tradeoff between methane reduction versus negative effects on diet digestibility was achieved at the low and intermediate rather than highest inclusion levels. In this way the most promising result was observed for *A. spinosus* and *C. bipinnatus*. The impact of this inhibition on ruminant whole-animal energy needs to be evaluated *in vivo* in order to use these plants in sustainable methane mitigation strategies provided they show the same effects as *in vitro*.

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