

CONSERVATION GENETICS OF AN ENDANGERED CYCAD, *Dioon sonorense* (ZAMIACEAE): IMPLICATIONS FROM VARIATION OF CHLOROPLAST DNA

JOSÉ SAID GUTIÉRREZ-ORTEGA^{1,3}, TADASHI KAJITA¹ AND FRANCISCO E. MOLINA-FREANER²

¹Department of Biology, Graduate School of Sciences, Chiba University, Inage-ku, Chiba, Japan

²Departamento de Ecología de la Biodiversidad, Instituto de Ecología, Universidad Nacional Autónoma de México, Hermosillo, Sonora, Mexico

³Corresponding author: josespo@chiba-u.jp

Abstract: The endemic species *Dioon sonorense* comprises the northernmost group of populations of *Dioon*, in Sonora, Mexico. Although it is endangered, only the southern populations are protected within a preservation area, while the remaining populations have not been taken into account in management plans or genealogical studies. Describing the intraspecific genealogy and the geographical structure of the genetic variation of *D. sonorense* may help in the identification of units for further conservation. Therefore, the main objective of this study was to describe the intraspecific genealogy of populations throughout the range of distribution of the endangered species *D. sonorense*, using haplotypes of the *trnL-F* spacer of the cpDNA. In addition, the levels of diversity of the employed molecular marker were estimated. Low levels of variability were recorded and at least two evolutionary units, corresponding to each haplotype, were identified as components of the species. The inclusion of northern populations into conservation programs is recommended. In order to integrate these findings to the genealogy of genus *Dioon*, *trnL-F* sequences of all the remaining species were merged and analyzed. An ancestral haplotype with wide distribution was detected in several species and one haplotype in *D. sonorense* was found shared with its sister species, *D. tomasellii*. This work represents a first attempt to reconcile a genealogical approach in the identification of important centers of genetic diversity of Mexican cycads.

Key words: chloroplast DNA, conservation genetics, *Dioon sonorense*, endangered species.

Resumen: La especie endémica *Dioon sonorense* representa el grupo de poblaciones más norteño para el género *Dioon*, en Sonora, México. A pesar de su estatus en peligro de extinción, sólo las poblaciones del sur están protegidas dentro de un área de conservación, mientras que las poblaciones restantes no han sido consideradas en los planes de manejo o en estudios genealógicos. La descripción de la genealogía intraespecífica y la estructura geográfica de la variación genética de *D. sonorense* puede ayudar en la identificación de unidades evolutivas para su posterior conservación. Por lo tanto, el objetivo principal de este estudio fue describir la genealogía intraespecífica de las poblaciones de *D. sonorense* a lo largo de su rango de distribución, usando los haplotipos del espacio *trnL-F* del ADNcp. Además, se estimaron los niveles de diversidad genética usando este marcador molecular. Se obtuvieron niveles bajos de variabilidad y se identificaron al menos dos unidades evolutivas, correspondientes a cada haplotipo que componen a la especie. Se recomienda la inclusión de las poblaciones del norte en los programas de conservación. Con el fin de integrar estos hallazgos en el contexto de la genealogía del género *Dioon*, las secuencias del espacio *trnL-F* de todas las especies restantes fueron combinadas y analizadas. Se detectó un haplotipo ancestral con una amplia distribución en varias especies y un haplotipo en *D. sonorense* se encontró compartido con su especie hermana, *D. tomasellii*. Este trabajo representa un primer intento de reconciliar un enfoque genealógico con la identificación de centros importantes de diversidad genética en las cícadas mexicanas.

Palabras clave: ADN del cloroplasto, *Dioon sonorense*, especie en peligro de extinción, genética de la conservación.

Mexico shelters a great diversity of cycads, and all of them are threatened with extinction and subject to conservation efforts (Osborne *et al.*, 2012). Fifty-four species of the genera *Zamia*, *Ceratozamia*, and *Dioon* are distributed within Mexico, while the latter two genera are

endemic to Mexico and Central America. The center of diversification of *Dioon* is assumed to be in Mexico (Moynihan, 2008), where thirteen *Dioon* species are found, while only one more species is found within both Honduras and Nicaragua (Bonta *et al.*, 2006).

Dioon sonorensis (De Luca, Sabato & Vázquez Torres) Chemnick, T.J.Greg. & S.Salas-Mor. is the species with the northernmost range of distribution, in the Sierra Madre Occidental and neighbor mountain ranges in the states of Sonora and Sinaloa, Mexico (Whitelock, 2002; González-Astorga *et al.*, 2009). Only four populations of *D. sonorensis*, labeled as Mazatán, El Novillo, Nuri, and Álamos, have been reported in population genetic studies (González-Astorga *et al.*, 2008, 2009); however, the exact number of populations and their status (number of individuals and present condition) are still uncertain. Although its endangered status is recognized by international institutions (Endangered A2ce+4c; C1 ver 3.1; IUCN, 2013), and by federal laws (Category P with MER method; SEMARNAT, 2010), only the southern populations are protected within a preservation area (Área de Protección de Flora y Fauna Sierra de Álamos-Río Cuachuajui), whereas the remaining populations are still being excluded from federal management plans (CONANP, 2013) or genealogical analyses.

Although conservation programs are generally founded on morphological taxonomic units, in some cases, morphological traits lead to little or no recognition of intraspecific evolutionary groups (Taberlet and Bouvet, 1994). For that reason, examining patterns of genetic diversity has become an important facet for conservation (Matocq and Villablanca, 2001). Previous studies have described allozyme variation of six *Dioon* species (González-Astorga *et al.*, 2003, 2005, 2008, 2009; Cabrera-Toledo *et al.*, 2008). In combination, the mean values of alleles per locus (A), expected heterozygosity (He) and percentage of polymorphic loci (P) are 1.8, 0.286, and 70.2, respectively (Piñero *et al.*, 2008). For *D. sonorensis*, those values were greater (A = 2, He = 0.314, P = 81.6; González-Astorga *et al.*, 2008, 2009), and showed low levels of differentiation among populations (G_{st} = 0.151). However, the methods used were unable to detect

genealogical differentiation which could distinguish evolutionary units. Also, the number of populations surveyed in those studies was limited and only covered a part of the distribution. From a different perspective, since the chloroplast genome of cycads is maternally inherited (Cafasso *et al.*, 2001) and has low rates of neutral substitution (Ravi *et al.*, 2008), low levels of genetic variation at the intraspecific level are expected and can be directly correlated with historical dispersal of the seeds, allowing for the distinction of sets of populations which have a common recent history and might be considered as evolutionarily significant units (ESU) for conservation (Moritz, 1994).

Identifying evolutionary units may also help solve the relationships within the genus *Dioon*. Although *D. sonorensis* is commonly shown as sister species to *D. tomasellii*, the phylogenetic relationships among the other *Dioon* species still remain unresolved due to low levels of interspecific differentiation and low number of analyzed samples per species in previous phylogenetic studies (Moretti *et al.*, 1993; González *et al.*, 2008; Moynihan, 2008). Sampling schemes that consider different intraspecific groups might provide a better resolution in the understanding of the evolution of this genus. Also, in view of the low levels of variation among *Dioon* species, instead of using a phylogenetic approach aimed to obtain resolution of species trees, a genealogical perspective describing the history of the cpDNA may provide better description of the evolutionary relationships (Rosenberg and Nordborg, 2002).

The main objective of this study was identifying evolutionary units for further conservation, based on the resolution of the intraspecific genealogy of the cpDNA of the species *Dioon sonorensis*. Moreover, in an attempt to glimpse the genealogical relationships of the detected haplotypes at interspecific level, the obtained data were merged with homologous sequences of other *Dioon* species.

Table 1. Estimated number of individuals per population based on field observation (Est), number of sampled (Sam) and analyzed (Ana) individuals per population, haplotype detected (Hap), estimated values of haplotype and nucleotide diversity (Hd, π), Watterson index (θ -W) and Tajima's D (D) are listed. i = parameter considering one indel, SA. = Sierra de Álamos, SM. = Sierra de Mazatán.

Population	Est	Sam	Ana	Hap	Hap _i	Hd	Hd _i	π	θ -W	D
1) Choquincahui	75	38	10	B	B	0	0	0	-	-
2) SA. Cañada del Altar	50	35	10	B	B	0	0	0	-	-
3) Nuri	8	8	8	A	Ai	0	0	0	-	-
4) Bacanora	60	33	10	A	Ai	0	0	0	-	-
5) El Novillo-Bacanora	7	7	6	A	A	0	0	0	-	-
6) El Novillo	2	2	2	A	A	0	0	0	-	-
7) SM. Peñón de las Iglesias	150	60	11	A	A	0	0	0	-	-
8) SM. Cañada de la Leona	100	45	10	A	A	0	0	0	-	-
9) SM. Rancho La Cueva	75	30	10	A	A	0	0	0	-	-
All populations combined	527	258	77	A, B	A, Ai, B	0.390 ± 0.049	0.6295	0.0004 ± 0.00005	0.00021 ± 0.00021	1.10825; P > 0.10

Table 2. *Dioon* species and localities of specimens analyzed. When necessary, the identification number of the specimens recorded by González *et al.* (2008) are indicated in parenthesis with their GenBank access number. ID = identification number of specimen deposited in Jardín Botánico F. J. Clavijero, N = number of samples, Hap = detected haplotypes, ^a = sequence obtained from a sample received from the Jardín Botánico F. J. Clavijero, ^b = identical obtained sequence from a specimen that was also analyzed by González *et al.* (2008), NI = sample from Jardín Botánico F. J. Clavijero without ID, U/D = unavailable data.

Taxon and original locality	N	Hap	ID and GenBank sequence accession
<i>D. sonorensis</i> (De Luca, Sabato & Vázq.Torres)			
Chemnick, T.J.Greg. & S.Salas-Mor.			
1) Choquinahui, Sonora	10	B	
2) Álamos, Sonora	12	B	(A) DQ926776; (B) DQ926777
3) Nuri, Sonora	8	A	
4) Bacanora, Sonora	10	A	
5) El Novillo-Bacanora, Sonora	6	A	
6) El Novillo, Sonora	2	A	
7) Peñón de las Iglesias, Sierra de Mazatán, Sonora	11	A	
8) Cañada de la Leona, Sierra de Mazatán, Sonora	10	A	
9) Rancho La Cueva, Sierra de Mazatán, Sonora	10	A	
<i>D. angustifolium</i> Miq.			
10) Rancho Muralla, Linares, Nuevo León	2	C	(A) DQ926750; (B) DQ926751
<i>D. argenteum</i> T.J.Greg, Chemnick, Salas-Mor. & Vovides			
11) San Pedro Yolox, Oaxaca	4	D, E	(A)DQ926752; (B) DQ926753; (C) DQ926754; (D) ^a U/D
<i>D. califanoi</i> De Luca & Sabato			
12) Teotitlán, Oaxaca	2	C	(A) ^{a, b} DQ926755; (B) DQ926756
<i>D. caputoi</i> De Luca, Sabato & Vázq.Torres			
13) Caltepec, Puebla	2	C	(A) DQ926757; (B) DQ926758
14) San Luis Atlotitlán, Puebla	1	C	(NI) ^a N/D
<i>D. edule</i> Lindl.			
15) Monte Oscuro, Veracruz	2	F, G	(A) DQ926759; (B) DQ926760
16) Municipio Casas, Tamaulipas	1	H	(C) DQ926761
17) Tamasopo, San Luis Potosí	1	I	(D) DQ926762
18) Actopan, Veracruz	1	C	(E) DQ926763
19) Coatepec, Veracruz	1	J	(F) ^{a, b} DQ926764
<i>D. holmgrenii</i> De Luca, Sabato & Vázq.Torres			
20) San Gabriel Mixtepec, Oaxaca	3	K, L	(A) DQ926765; (B) DQ926766; (C) ^a N/D
<i>D. mejiae</i> Standl. & L.O.Williams			
21) Gualaco, Honduras	1	M	(A) DQ926767
22) Esquipulas, Honduras	1	C	(B) DQ926768
23) Olanchito, Honduras	1	C	(C) DQ926769
<i>D. merolae</i> De Luca, Sabato & Vázq.Torres			
24) Jiquipilas, Chiapas	2	N	(A) DQ926770; (B) DQ926771
<i>D. purpusii</i> Rose			
25) Cuicatlán, Oaxaca	2	C	(A) DQ926772; (B) DQ926773
26) Pápalo, Oaxaca	1	O	(S) ^a U/D
<i>D. rzedowskii</i> De Luca, A. Moretti, Sabato & Vázq.Torres			
27) San Bartolomé, Oaxaca	2	C, P	(A) DQ926774; (B) DQ926775
<i>D. spinulosum</i> Dyer & Eichler			
28) San Juan Bautista Tuxtepec, La Mina, Oaxaca	1	Q	(B) DQ926778
<i>D. stevensonii</i> Nicolalde-Morejón & Vovides			
29) El Escondido, Guerrero	1	R	(J) ^a U/D
<i>D. tomasellii</i> De Luca, Sabato & Vázq.Torres			
30) Cabo Corrientes, Jalisco	1	C	(A) DQ926779
31) Zirándaro, Guerrero	1	C	(C) DQ926780
32) Compostela, Nayarit	1	A	(F) ^a U/D

Materials and methods

Sampling. One leaflet per individual of each patch from nine populations of *Dioon sonorensis* was sampled throughout the range of distribution in Sonora, Mexico (Table 1). The names of the localities were recorded and the number of individuals was estimated by direct observation. For additional analyses, eight samples of other *Dioon* species (including information of their original localities) were received from the “Colección Nacional de Cycadas” of Jardín Botánico Francisco Javier Clavijero, Instituto de Ecología, A.C., Xalapa, Veracruz, Mexico (SEMARNAT Colección Científica VER.-FLO-228-09-09; Table 2).

DNA isolation, amplification, and sequencing. For each sample, total DNA was isolated following the protocol by Doyle and Doyle (1987). In a preliminary survey designed to detect variable sites on *Dioon sonorensis*, double stranded DNA of non-coding regions of the chloroplast DNA (cpDNA) was examined using at least one sample per population. The original references and primer sequences are listed in Appendix 1. The regions *atpB-rbcL* (765 bp), *atpF-H* (565 bp), *trnT-L* (456 bp), *trnH-psbA* (582 bp), and *rpl20-rps12* (770 bp), showed no polymorphisms. The intron *trnL* and the intergenic spacer *trnL-trnF* (978 bp) showed one transversion and one indel on a mononucleotide repeat region; and when possible, at least ten individuals per population were randomly selected for further analysis. Polymerase Chain Reactions (PCRs) were performed in a Takara PCR Thermal Cycler Dice at 10 μ L volumes containing 10X EX Taq buffer (Takara), 2.5mM MgCl₂, 250 μ M of each dNTP, 0.4 μ M of each primer, 0.2U rTaq DNA Polymerase (Takara) and 1 μ L of template DNA. Thermocycling conditions were 95 °C for 1 min; 35 cycles of 95 °C for 45 sec, 51 °C for 45 sec and 72 °C for 1 min; and a final extension at 72 °C for 5 min. Amplification products were confirmed by electrophoresis in 0.8% agarose gel with ethidium bromide 1%. Purification of PCR products was carried out using ExoSAP-IT (USB Corp., Cleveland, Ohio, USA) and cycle sequencing-reactions were produced using BigDye Terminator v.3.1 Cycle Sequencing Kit (Applied Biosystems) following the manufacturer's instructions. Sequencing-reaction products were purified by ethanol precipitation and Sanger sequencing was determined with an ABI 3500 DNA sequencer (Applied Biosystems).

Parameter estimation. All sequences were edited using the Bioedit v7.1.11 software (Hall, 1999). Two matrix data were built for parallel analyses: one including only *Dioon sonorensis* sequences obtained in this study; the second one included the *D. sonorensis* sequences, the eight samples of the other *Dioon* species received from Jardín Botánico Francisco Javier Clavijero, and the sequences of the *trnL-F* spacer of cpDNA from González *et al.* (2008), published

in GenBank. The original localities of the specimens were checked in Appendix 1 of that publication and located with Google Earth (Google Inc., 2013). In both sets of data, the alignment of sequences was performed with the Clustal X program (Thompson *et al.*, 1997) with minimal manual adjustments (Table 2).

On *Dioon sonorensis*, the DnaSP v5.10.01 software (Librado and Rozas, 2009) was used to calculate nucleotide diversity (π), haplotype diversity (H_d), haplotype diversity considering indel (H_{di}), the Watterson estimator ($\theta-W$), and Tajima's statistical test (D). Also, the haplotype corresponding to each sample was identified using the Generate Haplotype Data command. All these parameters were estimated for both disregarding and considering the indel in the mononucleotide repeat region (site 657). For the alignment data of all *Dioon* sequences, 118 sites with at least one ambiguous character (base N) in the borrowed sequences from GenBank were considered as equal to the consensus. Because the length of the sequences from GenBank was shorter in the 3' extreme, the total length of the alignment was cut in order to fit as many sequences as possible. Since consideration of mononucleotide repeat regions is not suitable for genealogical studies (Selkoe and Toonen, 2006), the mononucleotide repeat in the alignment data of total samples of *Dioon* was deleted (12 bp starting from site 657). The haplotype corresponding to each sample was identified using the DnaSP v5.10.01 software (Librado and Rozas, 2009) with the Generate Haplotype Data command. Also, the TCS v1.18 software (Clement *et al.*, 2000) was used to generate a parsimonious net of haplotypes at 95% of confidence, considering gaps as fifth state.

Results

Field work and observations. Nine localities along the range of distribution of the endangered cycad *Dioon sonorensis* were surveyed: the four reported by González-Astorga *et al.* (2008, 2009), and five more populations. All the patches found were sampled, and the estimated number of individuals in each population is listed in Table 1.

Nucleotide and haplotype diversity within *Dioon sonorensis*. A total of 4,115 bp of non-coding regions of the cpDNA of *D. sonorensis* were explored in the preliminary analysis in order to detect intraspecific variability. Only two polymorphic sites (one transversion and one mononucleotide repeat indel) were found within the intron *trnL* and the intergenic spacer *trnL-trnF* (978 bp). For further analysis, a total of 77 sequences of this region were obtained from nine populations of *D. sonorensis*. Haplotype diversity considering indel (H_{di}) and nucleotide diversity (π) were estimated to be 0.6295 and 0.0004, respectively (Table 1). All the populations were monomorphic.

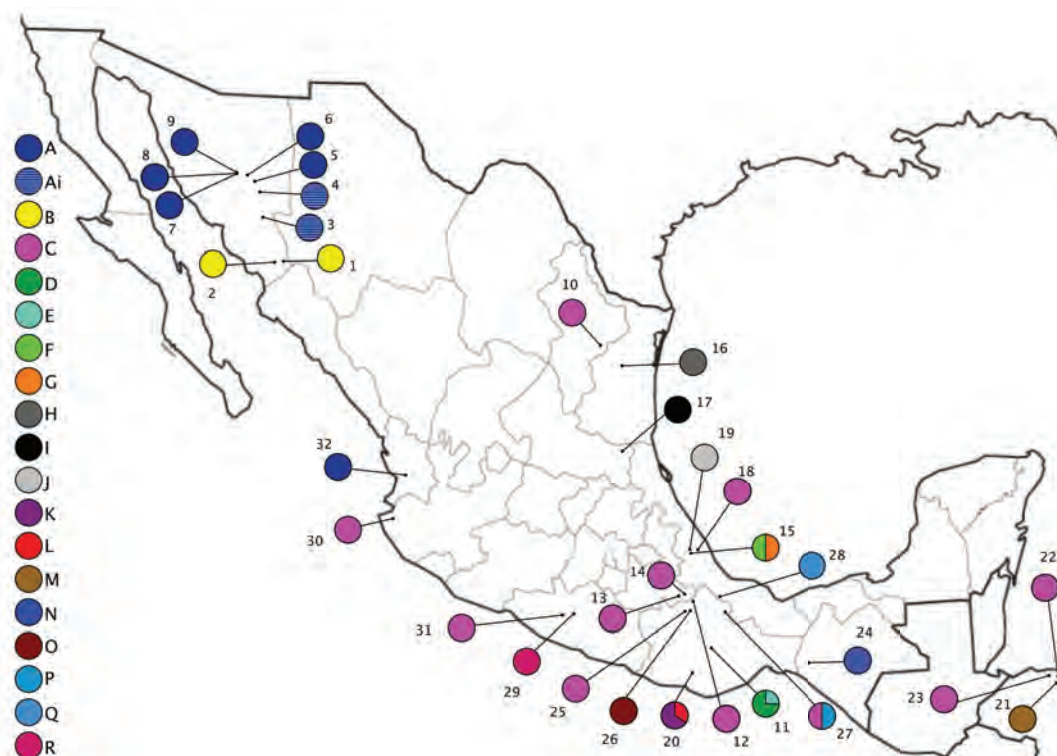


Figure 1. Distribution of 19 haplotypes of the *trnL-F* spacer recovered from all 14 currently described species of the genus *Dioon* (haplotypes labeled A-R). Pie charts reflect the proportion of occurrence of each haplotype in each locality. Location numbers correspond to those listed in Table 1.

*Geographical distribution of the haplotypes of *Dioon sonorensis*.* For the *trnL* intron and the non-coding region *trnL-F* of the cpDNA of *D. sonorensis*, three haplotypes were found among nine populations. Haplotypes A, Ai, and B, corresponded to northern, central, and southern populations, respectively (Figure 1). Haplotype A was found in three populations from Sierra de Mazatán, in El Novillo, and in the population next to the road El Novillo-Bacanora. Haplotype Ai, which was detected in the middle range of distribution, Nuri and Bacanora, was differentiated from haplotype A only by one indel event; however, since indels in mononucleotide repeat regions are not suitable for genealogical analysis due to possible homoplasy biases (Selkoe and Toonen, 2006), haplotypes A and Ai were considered as equal in further analyses. Haplotype B was found in the two southern protected populations: Sierra de Álamos and Choquinahui, and sequences were congruent with those borrowed from GenBank (Specimen A = DQ926776; Specimen B = DQ926777).

Haplotype network. A total of 114 sequences of the *trnL* intron and *trnL-F* region were compiled for the haplotype network. It comprised 77 sequences of *Dioon sonorensis*, six new sequences of other *Dioon* species, and 31 sequences borrowed from GenBank previously analyzed by González *et al.* (2008; Table 2). Thirty-two populations among all the

14 species of the genus *Dioon* were covered. The length of the alignment, excluding the mononucleotide repeat region, was 976 bp. In total, 19 haplotypes were detected in all the distribution range of *Dioon* (Table 3; Figures 1, 2). Whereas haplotype B was found exclusively in *D. sonorensis*, haplotype A was shared between the northern populations of *D. sonorensis* and one sample of *D. tomasellii*. Haplotype C was detected as shared among eight species, being the most widespread haplotype (Table 2; Figures 1, 2). The remaining haplotypes were found exclusively in other species.

Discussion

An extensive survey throughout nine populations of *Dioon sonorensis* was carried out. Besides the four populations analyzed by González-Astorga *et al.* (2008, 2009), five new populations analyzed with molecular markers are reported here. The sampling of these nine populations represents a clearer resolution in the identification of intraspecific groups on *D. sonorensis*. The analyses revealed low levels of genetic diversity for the *trnL* intron and *trnL-F* intergenic spacer of the cpDNA; however, three geographically structured haplotypes, A, Ai and B, were detected throughout its range of distribution, corresponding to northern, central and southern populations, respectively. Estimated genetic diversity and its conservation implications are discussed below.

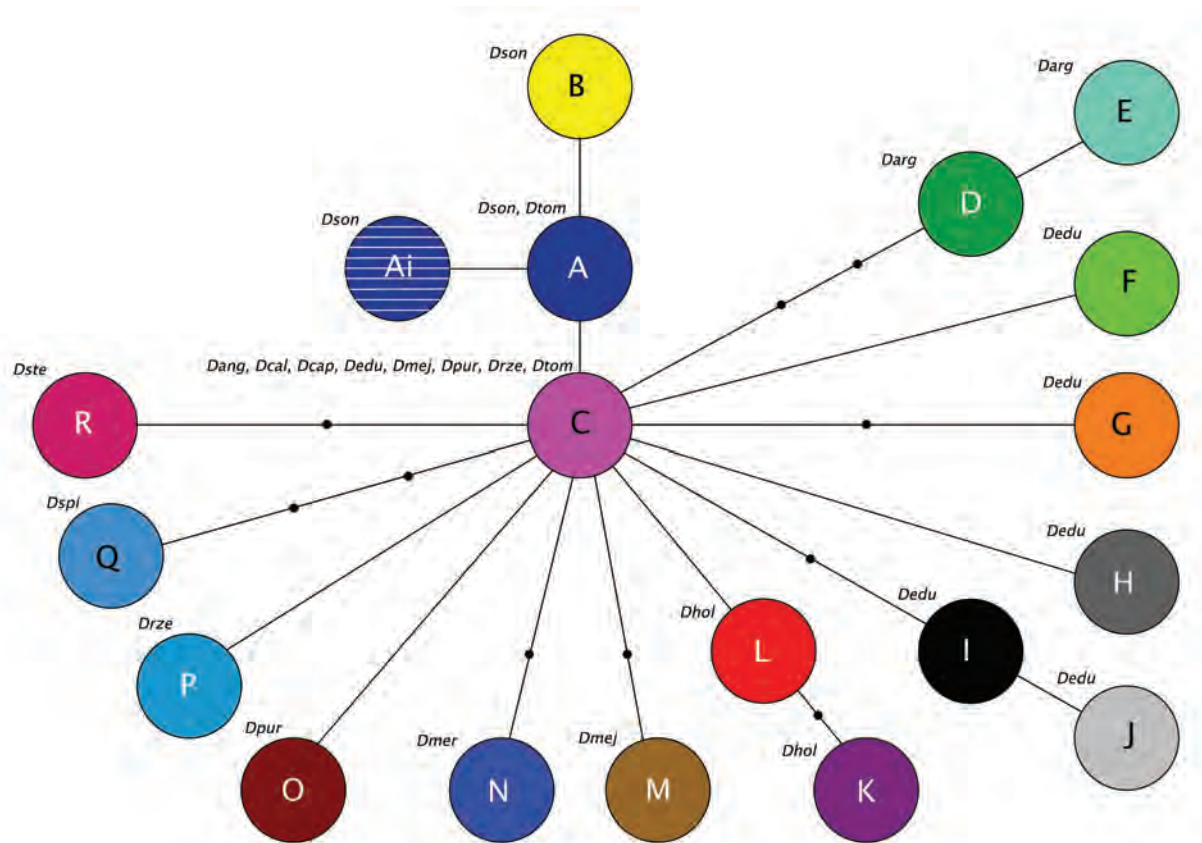


Figure 2. Haplotype network constructed under a criterion of statistical parsimony depicting mutational relationships among haplotypes recovered from *trnL* intron and *trnL-trnF* spacer from all 14 current described species of the genus *Dioon*. Letters as in Figure 1 identify haplotypes. Names of the species where the haplotypes were detected are noted next to the haplotype label: Dang = *D. angustifolium*, Darg = *D. argenteum*, Dcal = *D. califanoi*, Dcap = *D. caputoi*, Dedu = *D. edule*, Dhol = *D. holmgrenii*, Dmej = *D. mejiae*, Dmer = *D. merolae*, Dpur = *D. purpusii*, Drze = *D. rzedowskii*, Dson = *D. sonorensis*, Dspi = *D. spinulosum*, Dste = *D. stevensonii*, Dtom = *D. tomasellii*.

Genetic diversity and geographic structure. In a total of 4,115 bp from six non-coding regions of the cpDNA of the endangered cycad species *Dioon sonorensis*, only two sites were detected as variable. Both sites, one transversion and one indel, were found in the *trnL* intron and *trnL-F* region (978 bp) and allowed the estimation of low levels of genetic variation ($Hd_i = 0.6295$, $\pi = 0.0004$). Values found in *D. sonorensis* were similar to those found in the northernmost Asian cycad species *Cycas revoluta*, in Japan ($Hd = 0.641$, $\pi = 0.00071$; Kyoda and Setoguchi, 2010). For *C. revoluta*, the preliminary analyses surveyed a total of 14,130 bp of organelle DNA and only one polymorphic site and one indel were detected, resulting in three haplotypes showing geographic structure. In contrast, higher levels of cpDNA diversity are reported for *C. taitungensis*, a relict endemic species of Taiwan ($Hd = 0.998$, $\pi = 0.01268$), resulting in 97 haplotypes (Huang *et al.*, 2001). Variation in the levels of diversity of cpDNA in these species may be due to both demographic and genetic factors.

Because the estimation is done by pairwise comparison, the value of nucleotide diversity might be affected by the

presence of common haplotypes (no rare haplotypes were detected at the intraspecific level on *Dioon sonorensis*), therefore, low levels were expected (Nei, 1987). Usually, low values for π and Hd are interpreted as resulting from recent demographic bottlenecks, followed by a rapid expansion of populations (Avise, 2000). This idea is supported by Tajima's statistical test (D). Tajima's D test evaluates the neutrality of a molecular marker under the assumption of equilibrium conditions (when $\theta - W = \pi$). Although the sequences used in this study are from a non-coding region, which is not affected by natural selection, this spacer showed a deviated value (1.10825; not significant, $P > 0.10$), suggesting that populations may have been affected by historical events, such as demographic bottlenecks (Hedrick, 2005).

The northernmost populations that have haplotype A are located in Sierra de Mazatán. Three localities, one with about 75 individuals and two with more than 100 individuals, are reported here (Table 1). The distance between Rancho La Cueva and Cañada de la Leona is 6 km, which in turn, is 1 km from Peñón de las Iglesias. Despite their closeness, it is unknown whether gene flow by seed or pollen dispersal

Table 3. Variable sites of *trnL* intron and *trnL-F* used to determine haplotypes in genus *Dioon* (haplotypes labeled A-R). Hap = haplotype, NR= first site of mononucleotide repeat region, dots (.) = same character state as for Hap A, hyphen (—) = indel site.

Hap\ site	8	34	36	42	119	122	188	223	302	330	331	377	391	445	480	494	558	572	574	636	NR	657	715	848	850	895	932	939
Hap A	A	C	A	T	T	C	A	A	T	T	C	T	G	T	A	A	C	T	T	C	A	T	G	T	C	—	T	T
Hap Ai	—
Hap B	A
Hap C	C
Hap D	C	.	.	C	T	T	.	.	.	—
Hap E	C	.	.	C	T	.	.	C	.	.	T	.	.	.	—
Hap F	.	.	.	C	.	.	C	—
Hap G	C	T	—
Hap H	C	.	C
Hap I	C	C	C
Hap J	T	C	C	C
Hap K	C	C	C	.	—	G
Hap L	C	C	—
Hap M	.	A	C	.	.	.	C	—
Hap N	C	G	A
Hap O	C	T
Hap P	C	—	T	.	.
Hap Q	C	C	T	.	G	.
Hap R	C	.	C	C	.

occurs within this set of populations. For that purpose, other molecular markers, such as microsatellites, would give a better resolution (Selkoe and Toonen, 2006). In addition, haplotype A was also identified in El Novillo and next to the road El Novillo-Bacanora. In these localities, only two and seven individuals were found, respectively. There, local dwellers associate this low number of individuals to illegal collection for ornamental use and for the preparation of alcoholic beverages. Based on field observations of populations that share haplotype A, populations from the Sierra de Mazatán seem to be well conserved and their difficult accesses keep them largely away from illegal collectors. However, demographic studies are needed to know the structure of these populations, patterns of recruitment, and demographic tendencies.

Haplotype Ai occurs in populations from Bacanora and Nuri. The population of Bacanora is distributed only in the northern face of a mountain, where local people have commented that the number of individuals has decreased while elements of “monte mojino” (represented by thornscrub vegetation and *Prosopis* spp.) have been colonizing during the past 40 years. The population of Nuri was small, as only eight individuals were found next to a rural road. In this locality, signs of a recent extraction of plants were observed, this being a common practice, according to local testimonies. No significant geographical barriers exist between the ranges of distribution of haplotypes A and Ai; moreover, although the molecular marker employed in this study separates haplotypes A and Ai by one indel event in a mo-

nonucleotide repeat region, such difference is not suitable for genealogical analyses due to possible homoplasy biases (Selkoe and Toonen, 2006). For these reasons, haplotypes A and Ai may be considered a unique evolutionary unit.

Haplotype B was found in the two southern populations analyzed in this study. Both populations are protected within the range of the Área de Protección de Flora y Fauna Sierra de Álamos-Río Cuchujaqui (CONANP, 2013). The local people from Ejido Choquinahui have been involved in the conservation of the nearby populations of *Dioon sonorense*. However, the demographic study by Álvarez-Yépez *et al.* (2011) suggests a low long-term persistence probability. Since low number of adults, low quality of habitat, and low fecundity and recruitment were detected in that study, more management measures and a greater effort to avoid illegal collections is necessary for most of these populations. On the other hand, it is uncertain whether gene flow exists among this set of populations. A larger number of populations analyzed with different molecular markers would give a better resolution in the detection of important spots of genetic diversity in this conservation area.

Genealogy and species relationships. The data matrix which merged all the *Dioon* species consisted of 114 sequences of the *trnL-F* region of cpDNA. Thirty-one sequences used in this analysis were produced by the phylogenetic work of González *et al.* (2008); however, genealogical issues were not discussed in that publication. In that study, most of the resolution of topologies was obtained from the variation of

internal transcribed spacer (ITS) of ribosomal DNA, while *trnL-F* region showed low levels of variation. Although the variation of *trnL-F* region was unable to provide a concrete resolution on the relationships within a phylogenetic tree, its representation as haplotypes on a geographical setting (Figure 1) and on a haplotype network (Figure 2) permitted to formulate general inferences about the genealogy of the genus *Dioon*.

It is thought that the center of diversification of the current *Dioon* species was in southern Mexico (Moretti *et al.*, 1993; González *et al.*, 2008; Moynihan, 2008), and very recent migration events have been proposed as a result of successful seed dispersal during interglacial periods of the Pleistocene (Gregory and Chemnick, 2004). This idea seems to be supported by the paleoclimatic characterization of Toledo (1982), and suggests that the Sierra Madre mountain chains have played an important role in the migration of communities that are currently distributed within areas considered as Pleistocene refugia (Sauer, 1988; Contreras-Medina and Luna-Vega, 2007).

Haplotype A was detected in samples from populations of *Dioon sonorensis* and *D. tomasellii* which were uncovered in the phylogenetic analysis by González *et al.* (2008). This haplotype sharing indicates a close relationship between the species, being congruent with the taxonomic description for the taxa (de Luca *et al.*, 1984) and the phylogenetic analysis at genus level, in which these species are shown as sister species (González *et al.*, 2008; Moynihan, 2008). Haplotype C was detected as an ancestral haplotype and was found shared among eight species on a wide distributional range (Figure 1, 2). Recent divergence or ancestral polymorphism might explain this phenomenon, but more samples and different molecular markers are necessary to assess these probable scenarios. Though this analysis represents a first attempt in understanding the genealogy of the entire genus *Dioon*, a deeper sampling scheme including more populations of all species is necessary in order to obtain a robust resolution.

Conservation of populations of Dionn sonorensis. Since haplotypes A and B correspond to lineages that have a distinct evolutionary potential, at least these two sets of populations should be considered as independent ESUs to be included in conservation plans (Moritz, 1994). Although each population represents important spots of genetic diversity, some of them (e.g. El Novillo, El Novillo-Bacanora, and Nuri) seem to be senescent due to anthropogenic impact. In the northern distribution, populations from the Sierra de Mazatán should be conserved, and more studies regarding demography and genetic diversity are necessary to support its importance. Currently, the Sierra de Mazatán is proposed as an area for conservation as Área de Protección de Flora y Fauna, under the name of Sierra Huérfana, but its status has still not been approved by the Federal Govern-

ment (CONANP, 2013). Like the populations from Sierra de Mazatán, the locality in Bacanora is relatively isolated, and thus, illegal collection seems to be infrequent; however, its preservation is recommended and the demography should be studied to determine its current status and long term-persistence. Regarding the southern populations, although they are protected, more effective management plans should be performed for the preservation of *D. sonorensis*. Plans that include the collaboration of local people, like the one that was applied with the species *D. edule* in Veracruz, should be promoted (Vovides and Iglesias, 1994).

Despite the short length of the non-coding region *trnL-F* of the cpDNA of *Dioon sonorensis* (978 bp), it implies the level of variation through 4,115 bp and showed enough variation to distinguish geographical structure at the intraspecific level. For that reason, using it as a concise DNA marker to infer the origin of individuals of *D. sonorensis* seems applicable. The original localities of individuals deposited in collections, as well as individuals illegally collected, would be roughly traceable by the employment of this molecular marker.

Conclusions

In order to include different ESU for the conservation of the species *Dioon sonorensis*, this study proposes the conservation of populations in the Sierra de Mazatán, and while the southern populations are distributed within an official protected area, more efficient efforts are needed for the protection of these populations from anthropogenic impact. This work represents an example of how a genealogical perspective can be applied to the identification of evolutionary units to address both conservation and evolutionary issues on Mexican cycads.

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Appendix 1

Chloroplast DNA regions examined and primers used in this study.

Chloroplast region	Length of PCR products (bp)	Primers	Primer sequence (5'-3')	Primer source
<i>atpB-rbcL</i>	765	<i>atpB</i> -1	ACA TCK ART ACK GGA CCA ATA A	Chiang <i>et al.</i> , 1998
		<i>rbcL</i> -1	AAC ACC AGC TTT RAA TCC AA	
<i>atpF-H</i>	565	<i>atpF</i>	ACTCGCACACACTCCCTTTCC	Lahaye <i>et al.</i> , 2008
		<i>atpH</i>	GCTTTTATGGAAGCTTTAACAAT	
<i>trnT-trnL</i>	456	a	CAT TAC AAA TGC GAT GCT CT	Taberlet <i>et al.</i> , 1991
		b	TCT ACC GAT TTC GCC ATA TC	
Intron <i>trnL</i> + <i>trnL-F</i>	978	c	CGA AAT CGG TAG ACG CTA CG	Taberlet <i>et al.</i> , 1991
		f	ATT TGA ACT GGT GAC ACG AG	
<i>psbA-trnH</i>	582	<i>psbA3'</i> f	GTTATGCATGAACGTAATGCTC	Sang <i>et al.</i> , 1997; Tate and Simpson, 2003
		<i>trnH</i>	CGC GCA TGG TGG ATT CAC AAT CC	
<i>rpl20-rps12</i>	770	<i>rpl20</i>	CGY YAY CGA GCT ATA TAT CC	Shaw <i>et al.</i> , 2005
		5' <i>rps12</i>	ATT AGA AAN TCA AGA CAG CCA AT	