



Mycoparasitic antagonism of a *Trichoderma harzianum* strain isolated from banana plants in Oaxaca, Mexico

Antagonismo micoparasítico de una cepa de *Trichoderma harzianum* aislada de plantas de plátano en Oaxaca, México

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ABSTRACT

Bananas are important crops in developing countries with tropical climate. In Mexico, the banana production has increased, and it must be guaranteed. The Panama disease, caused by the fungus *Fusarium oxysporum* f.sp. *cubense* threatens the current banana production, for what is necessary to implement methods to protect this crop. Fungi from genus *Trichoderma* are natural residents of the rhizosphere, which comprises mycoparasite species used to control diseases caused by phytopathogenic fungi, and also benefit plant development. Here, we report a novel *Trichoderma harzianum* strain identified by optical microscopy and by DNA sequencing of 3 loci. The antagonistic capacity towards causal fungi of Panama disease was demonstrated by *in vitro* confrontation assay and co-inoculation in roots of 'plátano manzano' plants. Therefore, this fungus can be enabled to obtain organic formulations for biological control that contribute to produce pathogen-free plants and healthy and high quality fruits.

Keywords: Biological control, Panama disease, phytopathogen, plant health

RESUMEN

Los bananos son cultivos de importancia en países en vías de desarrollo con clima tropical. En México, la producción de banano se ha incrementado y debe de garantizarse. La enfermedad de Panamá causada por el hongo *Fusarium oxysporum* f.sp. *cubense* pone en riesgo producción actual de bananos, por lo que es necesario implementar métodos para proteger este cultivo. Hongos del género *Trichoderma* son residentes naturales de la rizosfera. Este género comprende especies micoparásitas que se han usado para controlar enfermedades causados por hongos fitopatógenos y que además benefician el desarrollo de las plantas. Es este reporte presentamos una nueva cepa de *Trichoderma harzianum* identificada por observaciones de microscopía óptica y secuenciación de 3 loci. Mediante ensayos de confrontación *in vitro* y co-inoculación en raíces de plátano manzano se demostró su capacidad antagonista sobre hongos causales de enfermedad de Panamá. Por lo tanto, este hongo puede ser

empleado para obtener formulaciones de control biológico que contribuyan a producir de plantas libres de patógenos y frutos sanos de alta calidad.

Palabras clave: Control biológico, enfermedad de Panamá, fitopatógeno, sanidad vegetal,

INTRODUCTION

Production of bananas and plantains (*Musa* sp.) is a significant income in countries with tropical climate in Latin America, the Caribbean, Africa and southeastern Asia. In 2018, global gross export is estimated in 19.2 millions of tons (FAO 2018). In 2018, Mexico produced 2.8 millions tons in Chiapas, Tabasco, Veracruz and other states of the Pacific coastline such as Oaxaca (SIAP 2018). Out of the 2.8 millions of tons produced in Mexico, 0.5 were exported, but exports increase year after year because of a trade agreement with China (Gobierno de México 2020).

The Panama disease, caused by the ascomycete fungus *Fusarium oxysporum* f.sp. *cubense* (*Foc*) is the more influential factor that has affected the banana production (Ordonez *et al.*, 2015). Based on the banana host cultivar infected, *Foc* is classified into 4 races, being *Foc* tropical race 4 (*FocTR4*) (aka *Fusarium odoratissimum*) the most aggressive one that infects a broad range of cultivars, including plants from the Cavendish group, the most consumed cultivar world-wide (Fourie *et al.*, 2011). *FocTR4* was recently identified in Colombia (Garcia-Bastidas *et al.*, 2020). The fields of Mexico have been free of *FocTR4*, but the dispersion of *FocTR4* in Latin America is unavoidable, which implies future decrease in banana production.

Trichoderma comprises soil-borne and endophytic fungal species with antibiotic and mycoparasitic activities (Mukherjee *et al.*, 2013) *Trichoderma* spp. also facilitate nutrient uptake, produces plant hormones that benefit plant development and enhances molecular responses to biotic and abiotic stress (Contreras-Cornejo *et al.*, 2016). Formulations based on *T. asperellum*, *T. atroviridae*, *T. virens* and *T. harzianum* are commercially available and used in agriculture (Woo *et al.*, 2014).

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Production of toxic secondary compounds from distinct metabolic pathways produced by *Trichoderma* spp. contribute to suppress the growth of neighbor phytopathogenic fungi at the soil or rhizosphere (Keswani et al., 2014). Mycoparasitism is another biological feature of *Trichoderma* that influences fungal population in soil and rhizosphere. *Trichoderma* can form hook-like structure attached to hyphae of other fungi referred to as 'prey'. Physical pressure derived from the hook-like structures and the activity of cell wall degrading allows acquisition of carbon and nitrogen sources from the prey (Harman et al., 2004).

T. harzianum (teleomorph: *Hypocrea lixii*) can colonize the epidermis and the outer cortex of cucumber roots (Yedidia et al. 2001). While interacting with roots, *Trichoderma* stimulates the execution of the induced systemic resistance (ISR) that strengthen the defense responses (Martínez-Medina et al., 2013) jasmonic acid (JA). Besides the antagonistic activity, *Trichoderma* spp. produce hormones such as auxins that boost plant growth (Contreras-Cornejo et al. 2009).

Due to the need to sustain the production of banana fruits in Mexico, it is necessary to prospecting biocontrol agents, native from the crop area. Here, we report the identification of a potential biocontrol agent isolated from pseudostems of "platano macho" (*Musa* sp. AAB) grown in the coast of Oaxaca. Sequencing of the *internal transcribed spacer* (ITS), *large subunit ribosomal DNA* (LSU) and the *translation elongation factor-1α* (TEF-1α) confirmed the specie as *T. harzianum*. The results validated its beneficial effects and suggest that the exploration of plant-associated microbial communities is a reliable first step to develop novel products of crop protection.

MATERIALS AND METHODS

Isolation and identification of a novel *Trichoderma* strain

The strain was isolated from fragments of pseudostems coming from 'platano macho' (*Musa* sp. AAB cv. Macho) at the municipality of Villa de Tututepec de Melchor Ocampo, Oaxaca, Mexico (16° 08'N, 97° 36' W) in parallel of the experiments of isolation of *Fusarium* spp. previously reported (Maldonado-Bonilla et al., 2019). Cut and washed tissue samples of approximately 0.5 × 0.5 cm were disinfected with 70% ethanol for 1 min, 3% sodium hypochlorite for 10 min, and then, were rinsed three times with sterile distilled water. The pieces of disinfected plant tissue were placed in Potato Dextrose Agar (PDA) plates and incubated for 1 week at room temperature. A fast growing mycelium of white color coming from one of the pieces were transferred to fresh PDA plates, after 1 week, mycelium turns dark green and a concentric ring is formed due to the production of conidia. This fungus was denominated M110, and after identification it was consequently referred to as *ThM110*. Conidia from one PDA plate were harvested with glycerol 50% to store at -80 °C. Morphology was visualized by optical microscopy and lactophenol-cotton blue staining.

Polymerase chain reaction

Extraction of genomic DNA from *ThM110* was performed following the protocol reported to isolate DNA of *Fusarium* spp. (Lin et al., 2009). After extraction, DNA was diluted and subject to PCR by using MyTaq® (Bioline). Amplification of ITS was performed by using the universal primer combination ITS1 (5'-TCCGTAGGTGAACTGCGG-3') and ITS4 (5'-TCCTCCGCTTATTGATATGC-3') under the following conditions: 5 min at 95 °C of initial denaturation; 35 cycles of 94 °C for 30 s (denaturation), 55 °C for 60 s (annealing) and 72 °C for 60 s (extension); final extension of 5 min at 72 °C. Amplification of a LSU fragment was performed with the primer combination LROR (5'-ACCCGCTGAACCTAACG-3') and LR5 (5'-TCCTGAGGGAAACTTCG-3') by the following protocol: 5 min at 95 °C of initial denaturation; 35 cycles of 94 °C for 60 s (denaturation), 51 °C for 60s (annealing) and 72 °C for 60 s (extension); final extension of 5 min at 72 °C. The amplification of the TEF-1α fragment was performed with the primer combination tef71f (5'-CAAAATGGGTAAG-GAGGASAAGAG-3') and tef997R (5'-CAGTACCGGCRGCRA-TRATSAG-3'). The 'touchdown' amplification program was 4 min of initial denaturation at 94 °C; 4 cycles each of 60 s at 94 °C (denaturation), 90 s at 70 °C (annealing) and 90 s at 72 °C (extension); followed by 26 cycles with annealing temperature decreasing by 0.5 °C per cycle from 68 °C to 55 °C, followed by 12 cycles with annealing at 55 °C; and with a final extension of 7 min at 72 °C. These PCR conditions are available in the website of the International Subcommission on *Trichoderma* and *Hypocrea* Taxonomy (http://www.isth.info/isth/methods/method.php?method_id=9) and has been validated at least in another study (Jiang et al. 2016) Ascomycota. Amplification of PCR products was confirmed by agarose gel electrophoresis stained with Eco-Stain (Biobasic).

Cloning of TEF-1α

We decided to clone the TEF-1α amplicon since it is longer than 900bp which could hinder the full sequencing by using the above mentioned tef71f and tef997R. After the amplification, the PCR product was cloned into pGEM T-Easy (Promega) following the manufacturer's instructions. Ligation products were used to transform chemically competent *E. coli* JM109 cells. The transformation reactions were plated out in LB agar supplemented with 100 mg mL⁻¹ ampicillin, 10 µL of 100mM IPTG (Isopropyl β-D-1-thiogalactopyranoside) and 50 µL of 20 mg µL⁻¹ of X-gal (5-bromo-4-chloro-3-indolyl-β-D-galactopyranoside). Plasmids of white colonies were extracted and digested with EcoRI to screen for clones carrying the TEF-1α PCR product. The plasmid of the selected colony (pGEM-TEF-1α) was purified with the QIAprep Spin Miniprep Kit (QIAGEN).

DNA sequencing and analysis

The amplicons of the ITS and LSU regions were directly sequenced by capillary electrophoresis using the primers mentioned for the PCR. The purified pGEM- TEF-1α was sequenced by capillary electrophoresis with the primers

T7promoter (5'-TAATACGACTCACTATAGGG-3') and SP6 (5'-ATTTAGGTGACACTATAG-3'), both of them are suitable for the sequencing of DNA cloned into pGEM-T-Easy. Sequencing service was provided by Macrogen (<https://dna.macrogen.com/eng/index.jsp>). After the assembly of the full sequences, they were used as queries to search for nearest matches in Genebank using the BLASTN algorithm optimized for highly similar sequences in the nucleotide collection database (nr/nt) of the National Center for Biotechnology Information (<https://blast.ncbi.nlm.nih.gov/Blast.cgi>). Sequences were considered conspecific when the percentage of identity was higher than 97% between a given record and our queries (Ávila-Díaz et al., 2013). The sequences of *ITS*, *LSU* and *TEF-1 α* were submitted to Genebank with the accession numbers MN55334.1, MT549065.1 and MN557854.1, respectively.

***In vitro* antagonistic activity**

In vitro confrontation assays were performed to get a first insight into the antifungal capabilities of M110. The growth of the wilting-causing strains M1a, M5, M7, M104 and M108 of *Fusarium* spp. (Maldonado-Bonilla et al., 2019) a sampling was performed in symptomatic plantains growing in fields of Oaxaca, a coastal southern state of Mexico. A phylogenetic analysis based on the sequences of TEF 1- α and IGS revealed that three isolates belonged to the *Fusarium oxysporum* species complex, while two other isolates were identified as members of the *Fusarium fujikuroi* species complex. Furthermore, isolates from the same complex shared the same ITS2 sequence. Inoculation using spores of each isolate on the roots of *Musa* sp. AAB cv. Manzano produced wilting symptoms of varying severity, suggesting that the *Fusarium* wilt might not be caused only by *Fusarium oxysporum* f. sp. *cubense*. PCR-based detection of Secreted in Xylem (SIX) was determined in the presence of M110. *Fusarium* spp. and M110 spores were spread in PDA plates and incubated at 28°C for one week. After this period, the mycelia cover the whole plate. Squares of 0.5 cm² of newly grown mycelium from each phytopathogen were inoculated close to the margin of a 10 cm diameter Petri dish with PDA. At the opposite margin of the same Petri dish, a similar square of newly grown mycelium of M110 was inoculated. A Petri dish only inoculated with the phytopathogens were used as negative controls. The growth of phytopathogens was recorded at 7 days after inoculation. Every assay included three technical replicates and was repeated three times. The percent of growth was calculated by dividing the growth of the phytopathogen in presence of *ThM110* by the growth of the phytopathogen alone (Askew and Laing, 1994).

Assessment of plant protection

Musa sp. AAB cv. 'manzano' ('plátano manzano') plants used for this assay were developed by following the *in vitro* micropropagation method established in our workgroup (Maldonado-Bonilla et al., 2019) a sampling was performed in symptomatic plantains growing in fields of Oaxaca, a coastal southern state of Mexico. A phylogenetic analysis based on

the sequences of TEF 1- α and IGS revealed that three isolates belonged to the *Fusarium oxysporum* species complex, while two other isolates were identified as members of the *Fusarium fujikuroi* species complex. Furthermore, isolates from the same complex shared the same ITS2 sequence. Inoculation using spores of each isolate on the roots of *Musa* sp. AAB cv. Manzano produced wilting symptoms of varying severity, suggesting that the *Fusarium* wilt might not be caused only by *Fusarium oxysporum* f. sp. *cubense*. PCR-based detection of Secreted in Xylem (SIX). *In vitro* rooted plants produced by this method were transferred into covered plastic containers with vermiculite rinsed with 25% MS solution without sucrose, and incubated at 26 °C with a 16 h light and 8 h darkness photoperiod. The covers of the containers were gradually removed to prevent dryness. After three weeks of incubation, the plants already had cuticle and produced two newly emerging leaves. Then, the plants were taken from the plastic container. The two older leaves formed during the *in vitro* micropropagation were excised, and the roots were thoroughly washed with water to eliminate the vermiculite. In parallel, the M110 conidia were harvested with sterile distilled water from a Petri dish with one week-old cultures grown in PDA. The suspension was adjusted to 10⁶ conidia mL⁻¹. Twenty plants were inoculated with M110 by submerging the roots into the conidia suspension for 1 min. Additional 20 plants were mock-treated with water. Later on, 10 plants inoculated with M110 and 10 mock-treated plants were infected by adding 10 mL of a *Fusarium* sp spores suspension. M5 adjusted to 10⁶ conidia mL⁻¹. The growth of *Fusarium* sp. M5 and preparation of spore suspension was performed as mentioned above for M110. Out the remaining plants, 10 were reference plants inoculated with M110 and 10 plants only inoculated with water were kept as additional testing conditions. Symptoms were recorded 10 weeks after inoculation by sorting plants into 4 categories: Healthy, yellow leaves, yellow leaves with partial necrosis and necrosis in whole leaves (Maldonado-Bonilla et al., 2019). The experiment was repeated twice.

RESULTS AND DISCUSSION

T. harzianum is a cosmopolitan fungus found as saprophyte in soils, rotting plants, wood, bark as well as associated with living organisms such as plants and other fungi (Chaverri et al., 2015). A close interaction between *T. harzianum* strain T-2013 and cucumber roots induces defense responses, solubilizes phosphate and increases its concentration in roots and shoots, which in turns enhances the plant growth (Yedidia et al., 2001). The detection of different *T. harzianum* strains in roots, stems, leaves and plumules of cacao plants suggests its adaptation to set an endophytic growth and a tight interaction with plants (Bailey et al., 2008). Furthermore, *T. harzianum* as one of the species most commonly isolated as endophytes in the sapwood of tropical trees (Almeida et al., 2018), which might be an indirect evidence of translocation *in planta*. Such conserved feature of this species might be the crucial factor to isolate M110 from living pseudotems of 'plátano macho'.

Lactophenol-cotton blue staining was performed in order to identify the M110 species. *Trichoderma harzianum* M110 is recognized in PDA culture medium for its rapid growth and its yellow-green coloration with concentric growth rings formed by the production of conidia. Microscopically, it is characterized by erect, hyaline conidiophores with bottle-shaped phialides (5.5-) 6-6.8 (-8.5) μm in length and (1.5-) 1.6-1.7 (-2.8) μm in the upper part (Figure 1A). The conidia are hyaline to slightly greenish in KOH (5%), subglobose to slightly oblong, 2.4-2.9 (-3.3) \times (2.0-) 2.3-2.7 μm , with a smooth wall (Figure 1B). The description agrees with that made by Gamms and Bissett (2002). This morphology suggested that the isolated species is *T. harzianum*.

T. harzianum and closely-related species such as *T. guizhouense*, *T. lixii*, and *T. inhamatum* form the *T. harzianum* species complex (THSC). It is proposed that every species of the THSC has a unique geographic distribution, for example *T. harzianum* is distributed in North America and Europe, while *T. inhamatum* is distributed in South America. In this context, the location of the isolation of M110 (the Pacific coastline of Mexico) agrees with the expected distribution of this species (Chaverri et al., 2015).

A bias in the distribution of the species of the THSC might be influenced by the climate conditions, soil composition and the plant diversity that can allow of the endophytic growth. Members of THSC share a quite similar morphological characters, thus, additional criteria such as sequencing of phylogenetically informative loci are necessary to define the species. The *ITS* is considered the fungal barcode sequence (Ratnasingham and Hebert 2007). The full *ITS* sequence is composed by the 5.8S ribosomal gene flanked by the region *ITS1* at the 5' and the *ITS2* at the 3' (Pryce et al., 2003). *ITS1* and *ITS2* are transcribed together with the rest ribosomal operon including the 18S (SSU), 5.8S and the 28S (LSU) rRNAs. During maturation of rRNAs, the *ITS1* and *ITS2* are degraded. The *LSU* sequence is used as a marker for fungal molecular taxonomy, but, due to the lack of evolutionary pressure, most of the polymorphisms are located into the *ITS1* and *ITS2*.

The full *ITS* and a fragment derived from the *LSU* were amplified by PCR and sequenced. Both sequences were

submitted as query in Genebank to search for highly similar sequences by using the BLASTN algorithm. Table 1 shows the two matches of highest identity. The values of percent identity above 99% suggest the identified species is *T. harzianum*. In order to discriminate between members of the THSC and confirm the identity of M110, a fragment of the *TEF-1 α* gene was amplified, cloned and analyzed. The sequenced *TEF-1 α* fragment of M110 was subject to the workflow used to analyze *ITS* and *LSU*. The results are presented in Table 1. The best matches correspond to collection strains *T. harzianum*, thus, M110 is a novel strain of *T. harzianum* isolated from pseudostems of banana. To our knowledge, this is the first report is this species associated to banana plants in Mexico.

We performed confrontation assays in order to demonstrate its potential antagonistic activity. M110 and 5 strains of phytopathogenic *Fusarium* spp. were co-inoculated in PDA. Negative controls of PDA plates solely inoculated with the phytopathogens were run in parallel. After 4 days, when fungi start to touch each other, the growth of *Fusarium* spp. slightly decrease in the presence of M110 (data not shown), and the difference is enhanced at 7 days post inoculation when M110 covers the plate and the growth of *Fusarium* spp. is fully arrested. The results of representative plates after 7 days of inoculation are shown in Figure 2. Table 2 shows the results of *Fusarium* spp. growth percentage in the presence of M110. A scale of 5 classes based on the phytopathogen growth percentage with *Trichoderma* respect to the phytopathogen growth alone is recommended to rank the antagonism (Askew and Laing, 1994). The Class 1 is the most effective and does not allow the growth of the phytopathogen. In Class 2, the pathogen growth is up to 25%, in Class 3, the pathogen growth reaches up to 50%. In Class 4, both fungi locked at the point of contact; and Class 5 indicates the phytopathogen entirely grew in the medium, inhibiting the *Trichoderma* growth. The results of the *Fusarium* spp. growth percentage in the presence of M110 are shown in Table 2. The interactions M1a vs. M110, M5 vs M110, M7 vs. M110 and M108 vs. M110 displayed quite consistent results below 50% of growth. The interaction M104 vs. ThM110 gave high standard deviation, but the inhibition is visually evident. Therefore, M110 is associated to the Class 3 of antagonism.

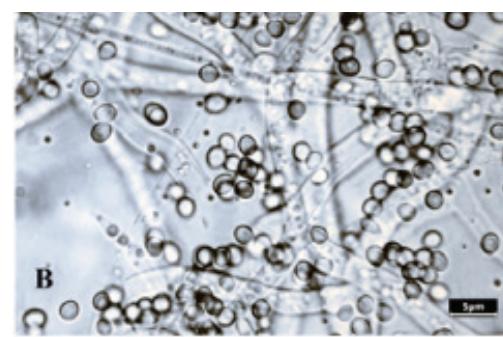


Figure 1. Optical microscopy reveals that M110 has characteristic features of *T. harzianum*. **A)** Bottle-shaped phialides. **B)** Subglobose to slightly oblong conidia. Bar: 5 mm.

Figure 1. Microscopía óptica revela que la cepa M110 tiene características típicas de *T. harzianum*. **A)** Fílides con forma de botella. **B)** Conidios subglobosos a ligeramente oblongos. Barra: 5 mm.

Table 1: Identity of the full length sequence *ITS*, and the derived fragments from *LSU* and *TEF-1α* of M110. The first two sequences of highest percent identity and coverage are shown. The access numbers of the corresponding sequences are indicated at the far right column.

Tabla 1. Identidad de la secuencia completa *ITS* y de los fragmentos derivados de *LSU* y *TEF-1α* de la cepa M110. Se muestran las primeras 2 secuencias con mayor porcentaje de identidad y cobertura. Los números de acceso de las correspondientes secuencias se indican en la última columna de la derecha.

Sequence	% Identity	% Coverage	E-value	Closest species match	Accession number
ITS	99.68	100	0	<i>T. harzianum</i> NECC30498	MH153641.1
	99.68	100	0	<i>T. harzianum</i> AA1I10F1	KX421472.1
LSU	99.88	100	0	<i>T. harzianum</i> CBS 130681	MH877292.1
	99.88	100	0	<i>T. harzianum</i> CBS 226.95	MH874152.1
<i>TEF-1α</i>	97.30	93	0	<i>T. harzianum</i> (<i>H. lixii</i>) CIB T131	EU279988.1
	96.14	94	0	<i>T. harzianum</i> JBT1244	AY605784.1

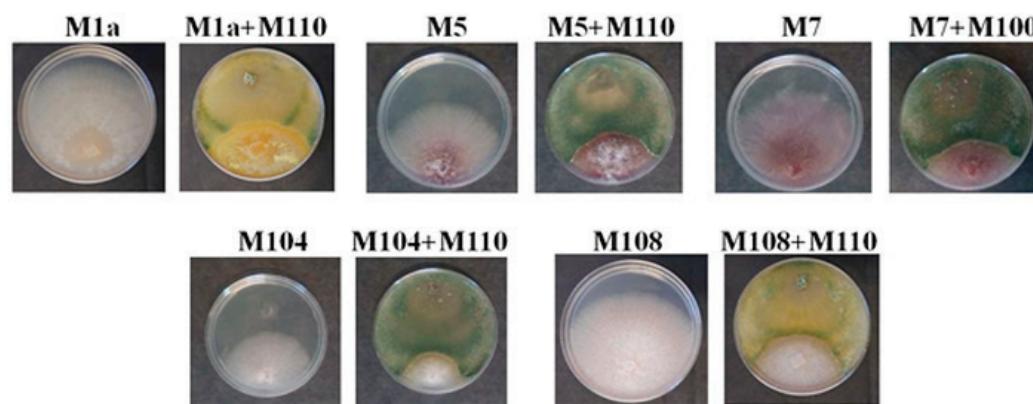


Figure 2. Representative photographs of the confrontation assays *Fusarium* spp. vs. *T. harzianum* M110 show that the mycelial growth of *Fusarium* spp. is inhibited by M110. Plates inoculated only with *Fusarium* were used as controls. The name of the inoculated strains is indicated above each plate. Photographs were taken 7 days post inoculation.

Figura 2. Fotos representativas de ensayos de confrontación *Fusarium* spp. vs. *T. harzianum* M110 muestran que el crecimiento micelial de las cepas de *Fusarium* spp. se inhibe en presencia de M110. Placas inoculadas sólo con *Fusarium* se usaron como controles. El nombre de las cepas inoculadas se indica sobre cada placa. Las fotografías se tomaron 7 días después de la inoculación.

Table 2. Percent of mycelial growth of *Fusarium* spp. in PDA plates co-inoculated with M110 respect to mycelial growth of *Fusarium* spp. in PDA plates without M110. See material and methods for details.

Tabla 2. Porcentaje de crecimiento micelial de *Fusarium* spp. en placas de PDA co-inoculadas con la cepa M110 respecto al crecimiento micelial de *Fusarium* spp. en placas PDA sin M110. Para detalles, ver materiales y métodos.

<i>Fusarium</i> strain	Percent of growth (DAY 7)
M1a	34.85 ± 5.52
M5	37.46 ± 4.20
M7	33.14 ± 5.49
M104	53.05 ± 12.93
M108	29.56 ± 5.26

Infection assays in the presence or absence of M110 were performed to validate the ability of M110 to antagonize causal agents of Panama disease in the soil or rhizosphere. After the *Foc* spore germinate, nascent hyphae adhere to the root epidermis, penetrate, and invade apoplast. When xylem vessels are reached, the fungus is dispersed throughout plant, causing discoloration of the rhizome and yellowing of leaves, and later on browning of the vascular bundles, necrosis and splitting of the pseudostem (Ploetz, 2015). Under our experimental conditions, strain M5 displays the more severe symptoms towards *Musa* sp. AAB cv 'manzano'. A M5 spores suspension was inoculated in roots of mock plants and plants previously inoculated with M110. The M110 potential protective effect is shown in Figure 3. Strain M5 causes severe wilting of the leaves, but the symptoms notably decreased in plants pre-inoculated with M110 (Figure 3A). By comparing

M110-inoculated and control plants, we suggest that M110 does not cause visual negative effects to *Musa* sp. AAB 'manzano'. Figure 3B shows the distribution of symptoms in the plants grown under these experimental conditions and confirms the pre-inoculation of M110 decreases of wilting of leaves caused by M5. Under these conditions, no further symptoms were detected. Therefore, M110 seems to be innocuous to plants and its inoculation into roots decreases the Panama disease symptoms, possibly by the antagonism characteristic of the *Trichoderma* genus.

Antifungal compounds such as peptidiols, polyketides and terpenes are produced by *Trichoderma* spp (Keswani et al., 2014). Such metabolites might limit the proliferation of other fungi that compete for carbon and nitrogen source and represent a benefit to the plant when phytopathogens are inhibited. Remodeling of the cell wall by the activity of chitinases, glucanases and proteases is required for hyphal growth and branching (Verdín et al., 2019). Those extracellular enzymes can reach the hyphae of neighbor fungi and

restrict their growth. Coiling around fungal host enhances the antagonistic effect of *Trichoderma* (Guzmán-Guzmán et al., 2018), since physical proximity between *Trichoderma* and host facilitates the effect of antifungal metabolites and the lytic enzymes mentioned above. Therefore, diffusible factors produced by *T. harzianum* M110 and the coiling might contribute to the *in vitro* growth arrest of *Fusarium* spp., and, in the same way, they could obstruct the development of *Fusarium* sp. M5 in the rhizosphere of *Musa* sp. AAB 'manzano'.

Besides the antagonism, *Trichoderma* might elicit mechanisms of protection. Genes encoding biosynthetic enzymes of biotic stress-related plant hormones such as salicylic acid (SA) and jasmonic acid (JA), have been identified in the *T. atroviridae* genome (Guzmán-Guzmán et al., 2018). A number of *Trichoderma*-derived proteins and non-ribosomal peptides can also trigger defense responses such as production of phytoalexins, reactive oxygen species, or induction of genes encoding pathogenesis-related proteins (Ramírez-Valdespino et al., 2019). These reactions have to be

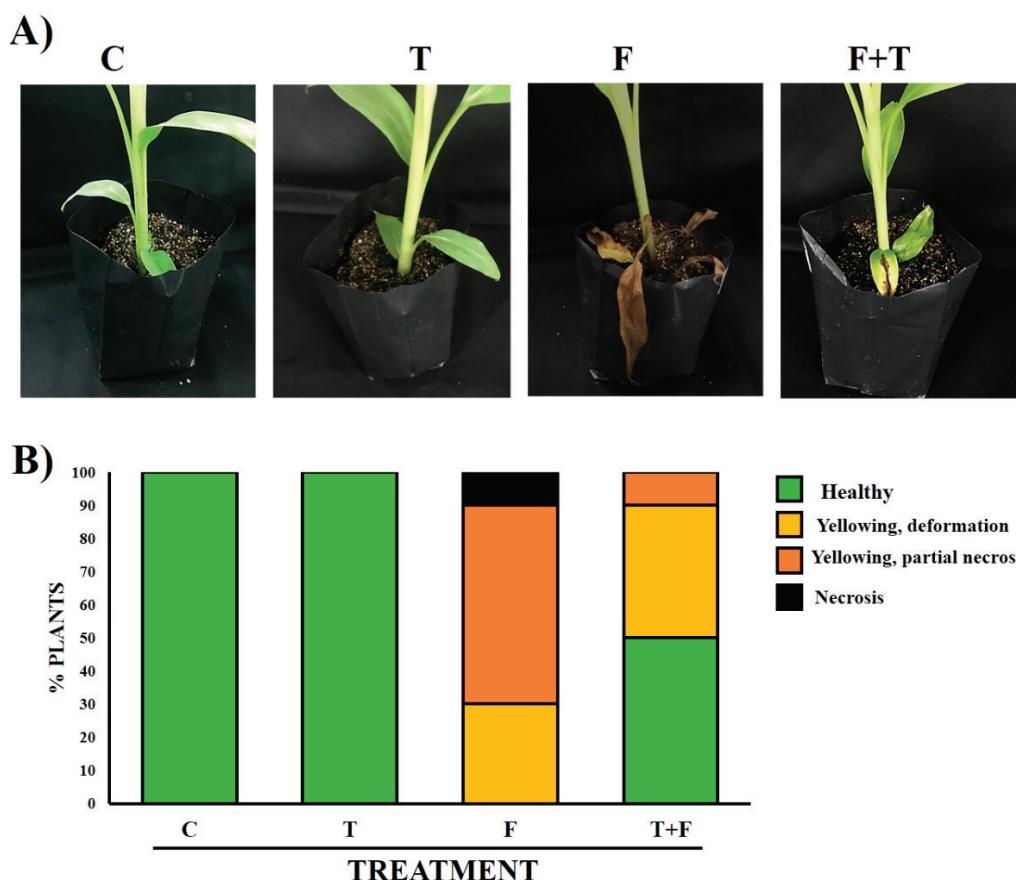


Figure 3. Evidences of the protective effect of M110 against *Fusarium* sp. M5. Spores solutions of M110 and M5 were inoculated in roots of *Musa* sp. AAB cv. 'manzano'. A) The effect is visible in plants 10 weeks after inoculation. B) Plants were classified according to the displayed symptoms of different severity at 10 weeks after inoculation. C, non-inoculated control. T, *T. harzianum* M110. F, *Fusarium* sp. M5. F+T, *T. harzianum* M110 + *Fusarium* sp. M5. See materials and methods for details of the inoculation procedure.

Figura 3. Evidencias del efecto protector de M110 contra *Fusarium* sp. M5. Soluciones de esporas de M110 y M5 se inocularon en raíces de *Musa* sp. AAB. cv 'manzano'. A) Efecto en plantas es visible a 10 semanas después de inocular. B) Las plantas fueron clasificadas de acuerdo a los síntomas mostrados 10 semanas después de la inoculación. C, plantas control no inoculadas, T, plantas inoculadas con *T. harzianum* M110. F, plantas inoculadas con *Fusarium* sp. M5. F+T, *T. harzianum* M110 + *Fusarium* sp. M5. Ver materiales y métodos para detalles de procedimiento de inoculación.

explored in the future in order to have a better understanding about the protective effect of M110 in 'plátano manzano'. Both SA and JA-dependent signal pathways are stimulated by *T. virens* in tomato plants, and induced resistance against *Fusarium oxysporum* f.sp. *lycopersici* (*Fol*) (Jogaiah et al., 2018). As the pathogenicity mechanisms of *Foc* and *Fol* are similar, we have to consider the role of the SA and JA pathways in banana, either elicited by hormones produced by the fungus or by molecules that trigger hormone biosynthesis and signaling in the plant. These possibilities can be investigated in M110 and banana plants.

A combination of *T. harzianum* and *Glomus* spp. suppressed the infection of *FocTR4* towards the 'Lakatan' bananas, a top-selling cultivar in Philippines (Castillo et al., 2019). A combined formulation of *T. harzianum* and *Bacillus subtilis* enhances the uptake of minerals and biomass production in swiss chard (Venegas-Gonzalez et al., 2019). That illustrates additional effects of *T. harzianum* that might be investigated in the future for the interaction between M110 and banana plants.

We report here a *T. harzianum* strain isolated from plants growing in southern Mexico, which has features of a biocontrol agent, and it can be efficient to protect plants against Panama disease. Some banana producers have faced disease-related restrictions and bans due to Panama disease (FAO, 2017). Mexico has an important trade to export bananas to China (Gobierno de México, 2020) thus, it is necessary to be alert in securing the plant health and production of pathogen-free bananas.

CONCLUSIONS

Due to the economic importance of banana in Mexico, and the need to sustain the production of fruits, it is necessary to prospecting biocontrol agents native from the soil of the crop area and adapted to interact with cultivars grown in Mexico. Here, we demonstrated the antagonism of *T. harzianum* M110 towards *Fusarium* spp., and demonstrated the potential of this novel strain as biocontrol agent. These findings encourage us to investigate the molecular basis of its antagonism and interaction with plants. Communication between microbiologists, phytopathologists and biotechnologists is necessary to develop large-scale biopesticide products as alternative to protect plants against Panama disease.

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