



SPECIAL ARTICLE

Ionic and substrate mechanisms of atrial fibrillation: rotors and the excitation frequency approach

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Atrial fibrillation; Dominant frequency; Rotors; Potassium currents; fibrosis; USA.

Abstract

Atrial fibrillation (AF) is the most common sustained cardiac arrhythmia in humans, however its mechanisms are poorly understood and its therapy is often sub-optimal. This article reviews recent experimental, numerical and clinical data on dynamics of wave propagation during AF and its mechanistic link to ionic and structural properties of the atria. At the onset, the article presents numerical and optical mapping data suggesting that a presence of periodic source with increasingly high dominant frequency (DF) of excitation underlies observations of dispersion of local activation rate during AF. Further optical mapping studies in isolated normal sheep hearts in the presence of acetylcholine (ACh) reveals that rotors localized to the left atrium (LA) drive the arrhythmia and are faster than those in the right atrium (RA). Patch-clamp data from isolated cardiomyocytes shows that the ACh-modulated potassium inward rectifier current is higher in the LA than in the RA which may explain the higher DFs and sensitivity of LA rotors to ACh compared with RA rotors. Following, the role of fibrosis in governing the propagation dynamics with a decrease in excitation frequency is presented in AF in failing sheep hearts and complex activation in cell cultures. Translation into the clinical setting is then discussed: DF distribution in patients with paroxysmal AF follows the LA-to-RA gradients found in the acute cholinergic AF of sheep hearts with highest DFs localized primarily to the posterior LA wall and pulmonary veins (PV) region; however in patients with persistent AF, the highest DFs localize mainly outside of the PVs region with possible implication on the outcome of ablation procedures. Next, intravenous injection of adenosine to patients in AF is demonstrated to result in further acceleration of high DF sites and suggests that reentrant activity, rather than triggered or automatic activity, maintains the arrhythmia. Finally, analysis of excitation during AF developed in patients post-cardiac surgery suggests a DF distribution similar to that of patients with paroxysmal AF with dependency on fibrosis as found in sheep failing hearts and cell cultures. In sum, the article presents data demonstrating the use of DF of excitation in linking wave propagation mechanisms to ionic and structural properties in both experimental and human AF.

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PALABRAS CLAVE

Fibrilación auricular;
Frecuencia dominante;
Corrientes de potasio;
Fibrosis; EEUU.

Mecanismos iónicos y sustratos en la fibrilación auricular; rotores y su relación con la frecuencia de excitación**Resumen**

La fibrilación auricular es la arritmia cardíaca más frecuente en los seres humanos; sus mecanismos aún no se entienden totalmente y la terapia utilizada para su tratamiento es, a menudo insuficiente. En este artículo se revisan los resultados numéricos y clínicos de experiencias recientes sobre la dinámica de la propagación de ondas durante la fibrilación auricular, y su relación mecánica con las propiedades iónicas y estructurales de la aurícula. Al inicio, el artículo presenta datos sobre el mapeo numérico y óptico, que sugieren que la presencia de una fuente periódica con un incremento de frecuencia dominante de excitación, sustenta las observaciones de la dispersión de la tasa de activación local durante la fibrilación auricular. Estudios subsecuentes de mapeo óptico en corazones normales de borregos con adición de acetilcolina, revelan que los rotores localizados en la aurícula izquierda impulsan la arritmia y son más rápidos que los de la aurícula derecha. Evidencias de "patch-clamp" en cardiomiocitos aislados muestran que la corriente rectificadora de entrada de potasio modulada por la acetilcolina es más alta en la aurícula izquierda que en la aurícula derecha, lo cual podría explicar las frecuencias dominantes más altas y la mayor sensibilidad a la acetilcolina de los rotores de la aurícula izquierda, comparados con los de la aurícula derecha. A seguir, se presenta el papel de la fibrosis en la dinámica de propagación, con una disminución de la frecuencia de excitación durante la fibrilación auricular en corazones enfermos de borregos, así como la activación completa en cultivos de células. Posteriormente se discute la aplicación al escenario clínico: la distribución de la frecuencia dominante en pacientes con fibrilación auricular paroxística sigue el gradiente: aurícula izquierda hacia la aurícula derecha hallados en la fibrilación auricular colinérgica aguda en corazones de borregos con las frecuencias dominantes más altas, principalmente localizadas en la pared posterior de la aurícula izquierda y en la región de las venas pulmonares; sin embargo en pacientes con fibrilación auricular persistente, la frecuencia dominante más alta se localiza principalmente fuera de la región de las venas pulmonares, con una posible implicación en los resultados de los procedimientos de ablación. A seguir, se demuestra que la inyección intravenosa de adenosina a pacientes con fibrilación auricular, resulta en mayor aceleración de los sitios de frecuencias dominantes altas, lo que sugiere que la actividad de re-entrada más bien mantiene la arritmia y no la actividad automática. Finalmente, el análisis de la excitación durante la fibrilación auricular desarrollada en pacientes post-cirugía cardíaca, sugiere una distribución de frecuencia dominante similar a la de los pacientes con fibrilación auricular paroxística con dependencia de fibrosis, tal como se evidenció en los corazones enfermos de borregos y en los cultivos celulares. En suma, el artículo presenta datos que demuestran el uso de la frecuencia dominante para correlacionar los mecanismos de propagación de onda con las propiedades iónicas y estructurales, tanto en la fibrilación auricular experimental como en la de los humanos.

Introduction

Atrial fibrillation (AF) is to date the most common sustained cardiac arrhythmia in humans and is predicted to dramatically increase its prevalence in the near future.¹ Antiarrhythmic drugs are only partially effective in treating AF, and have the potential for serious side effects, including life-threatening pro-arrhythmia. On the other hand, it has recently been demonstrated that paroxysmal AF in patients is initiated by focal triggers localized usually to one of the pulmonary veins (PVs)² and can be cured by a catheter-based ablation procedure.³ However, in persistent AF, the prevailing theory is that multiple random wavelets of activation coexist to create a chaotic cardiac rhythm,⁴ and therapy is more challenging.⁵

The dismal therapeutic outcome of AF could be in part due to poor understanding of its underlying mechanisms. A commonly accepted mechanism, the multiple wavelet hypothesis,^{6,7} characterizes cardiac fibrillation as randomly propagating waves with intermittent blockades, annihilation and re-generation of discrete waves. A consequence of such postulated dynamics is said to be a self sustained

complex spatio-temporal patterns of excitation lacking a hierarchical organization in the activation. However, the old idea of Lewis⁸ on reentry as the mechanism of fibrillation has re-emerged from theoretical^{9,10} and experimental¹¹ studies showing that wave propagation during AF is not totally random. This postulate is that rotors are the major organizing centers of fibrillation and that certain specific molecular properties of the cardiac muscle at the ion channel level contribute to the establishment of such rotors and to the overall complexity of the arrhythmia.¹² A consequence of such complex dynamics paradigm is a hierarchical distribution of local excitation frequencies. Thus, the early studies on regional differences in activation cycle length during AF by Morillo¹³ and Harada¹⁴ were followed by an increased interest in the analysis of the frequency spectrum as a measure of the activation rate in animal models.¹⁵⁻¹⁸ Those mapping studies have detected a clear hierarchy in the dominant frequency (DF) of excitation across the atria during fibrillation and correlated the location of the highest frequencies with that of relative organized activity and rotors.¹⁵⁻¹⁸ Furthermore, works demonstrating how measurements of AF cycle length in

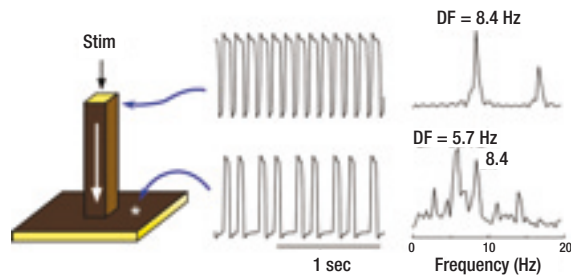


Figure 1. Computer model of action potential propagation from a pectinate muscle to the atrial wall. A 3-dimensional (60x60x60 elements) model includes a 1-dimensional bundle attached to a 2-dimensional sheet (left panel). Periodic stimulation (Stim) was applied at the top edge of the bundle and the impulse was allowed to propagate downward with conduction velocity of -0.29 m/sec and to invade the two dimensional sheet. The voltage time series and corresponding power spectra are shown for a site near the stimulation point and a site at the sheet. Comparison between the points indicates a 3:2 pattern of propagation into the sheet with a concomitant spectral transformation and a DF shift from 8.4 to 5.7 Hz. Reproduced from Jalife et al, *J Cardiovasc Electrophysiol.* 1998;9:S2-S12.12

patients can contribute to its treatment,^{19,20} motivated translation of experimental knowledge and methods into analysis of human AF in the frequency domain.²¹

This article has three major objectives: first, to discuss recent experimental and clinical data on wave propagation dynamics and AF maintenance with gradients of DF across the atria; second, to discuss data that strongly support the hypothesis that the ionic mechanism of AF in the structurally and electrophysiologically normal heart may be explained in part on the basis of chamber specific differences in the level of expression of cardiac potassium channels; and third, to discuss the role of structural fibrosis in the dynamics of AF. As will be demonstrated, spectral analysis with the determination of the DF of excitation provides a systematic approach for linking the dynamics of electrical wave propagation in cell cultures, isolated sheep hearts and humans to underlying ionic and structural determinants of AF.

Frequency-dependent breakdown of wave propagation

To gain insight into the distinct CLs in Morillo's study,¹³ we measured the frequencies at varying locations of both atria and found them to be widely different. Most notably, the activation frequencies in certain areas of the left atrium (LA) were always faster than any other region.^{15,16} Here we use computer simulation to gain more mechanistic insight into the origin of such data. We start with the premise that the underlying mechanism of AF may be a highly periodic stationary source (e.g., a rotor or an automatic focus) located somewhere in the LA.^{17,18} In addition, we assume that, regardless of the nature of the periodic source, the complicated anatomy of the atria plays an integral role in the development of fibrillatory conduction. These assumptions are conveniently considered using a simplified mathematical model of a major pectinate muscle connected to a small area of the atrial wall.¹² The

three-dimensional model consisted therefore of a one-dimensional thin bundle attached to a two-dimensional sheet (Figure 1). Periodic stimulation was applied to the top free edge of the thin (25 mm^2) bundle and the impulse was allowed to propagate downstream to invade the two-dimensional sheet.¹² The traces on the right show the action potentials and corresponding power spectra of sites in the bundle and in the sheet. As shown by the top and bottom time series, stimulation at a constant period of 0.119 sec resulted in a 3:2 propagation pattern across the boundary between the thin bundle and the sheet. This is reflected also in the corresponding power spectra: While the source region (i.e., the thin bundle) displayed a DF of 8.4 Hz, the geometrical expansion into the sheet imposed a spectral transformation whereby the DF shifted to 5.7 Hz. The two power spectra display additional peaks originating from the combined effect of the sharp action potential deflections and the inter-beat cycle length variations. The constant cycle length at the thin bundle results in a narrow peak at a DF that is the exact inverse of the cycle length (CL, $1/0.119 = 8.4$ Hz) with an additional smaller peak at about 16.8 Hz, which is an integer multiple of the DF (i.e., a harmonic). The CL of the activity in the sheet on the other hand is not constant and can be seen to alternate between short and long values. This in turn gives rise to a more complex profile: Several peaks are seen in the power spectrum, consequent of the combination of various intervals in the time series including not only the long and short CLs, but also their sum and difference. The Fourier algorithm, nevertheless, considers the most stationary combination of the CLs to be the DF at 5.7 Hz, which is the average number of local activations per second,²² corresponding to the 3:2 ratio of the input cycle length (frequency) of 0.119 sec (8.4 Hz) represented by a smaller peak.

Keeping in mind all the mathematical and clinical factors discussed above, in the following sections I discuss recent studies that used power spectral analysis of electrical activity during AF. As the reader will be able to tell, in those studies, investigators were able to successfully assign the dominant frequency an electrophysiological meaning to determine AF mechanisms and to guide the AF ablation procedure. It is well known that during atrial flutter both atria are excited in a 1:1 manner at frequencies as high as 300 beats/min (5 Hz). On the other hand, data obtained from experimentally induced AF in the sheep heart do show that stationary reentrant sources in the LA can rotate at frequencies >7 Hz and generate impulses that propagate toward the right atrium (RA).^{17,18} Yet the RA in induced AF in sheep experiments is activated at much lower frequencies. This suggests that there must be a critical frequency at which the 1:1 input-output relation between the LA and RA breaks down.

Figure 2 shows results of optical mapping experiments and DF analysis in the isolated RA of a sheep heart addressing the mechanism underlying of such lack of 1:1 input-output relationship. The preparation was subjected to periodic pacing through a bipolar electrode on Bachmann's Bundle (BB) to simulate activity arriving from a periodic LA source across BB into the RA.²³ As shown in panel A, stimulation at 5.0 Hz resulted in 1:1 activation of the entire right atrium and thus the output DF was also 5.0 Hz

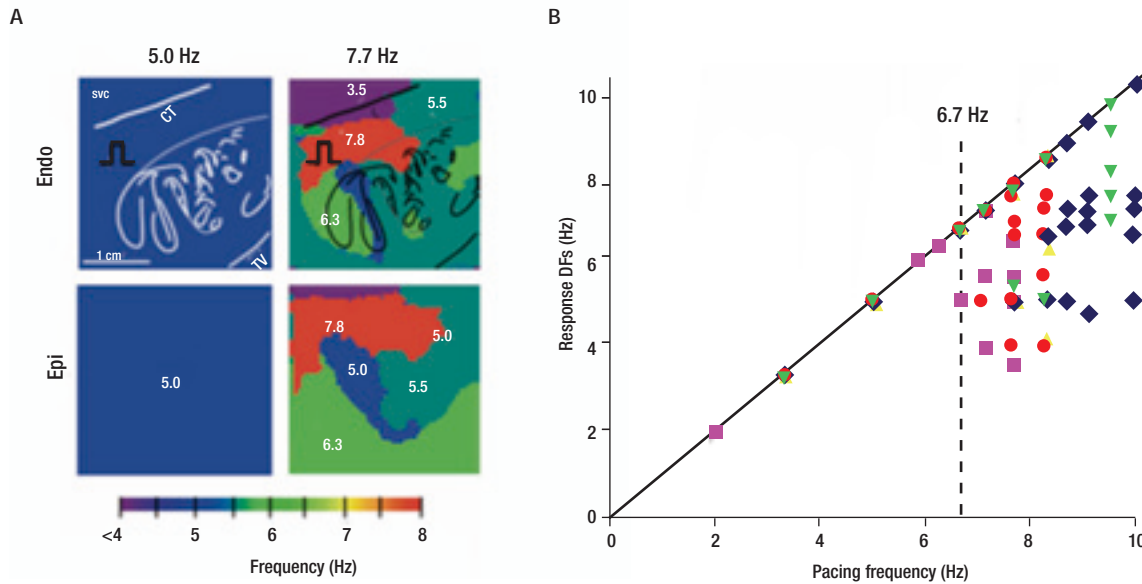


Figure 2. The ‘breakdown frequency’ in a sheep heart. A, Endocardial and epicardial DF maps of same isolated RA preparation paced at 5.0 and 7.7 Hz. Note appearance of heterogeneous DF domains at 7.7 Hz. B, Response DFs versus the pacing rate ($n=5$). Each symbol represents one experiment. Pacing BB at rates below ~ 6.7 Hz, results in 1:1 activation. At higher rates, the number of domains increases but the DFs’ value decrease. SVC, superior vena cava; CT, crista terminalis. Reproduced from Berenfeld et al, *Circ Res* 2002;90:1173-1180.²³

everywhere on the endocardial and epicardial surfaces. However, at 7.7 Hz, propagation into the RA was no longer 1:1. Instead, a heterogeneous distribution of DF domains was established both in the endocardium and the epicardium, with frequencies ranging between 3.5 and 7.7 Hz. Composite data from five experiments are presented in panel B. The DFs measured on the endocardium, are plotted as a function of the pacing frequency. Clearly, below 6.7 Hz the response DF showed no dispersion in any of the experiments, which meant that activation was 1:1 everywhere in the RA. Above the ‘breakdown frequency’ of 6.7 Hz, there was a large DF dispersion manifested as multiple domains whose individual frequencies were either equal or lower than the pacing frequency. In addition, we found that intermittent blocks occur primarily at branching sites of the pectinate bundles of the atria. Those structurally related blockades lead to a significant loss of consistency in the beat-to-beat direction of wave propagation and provided a direct explanation for the difficulty in tracing back the origin of the activation during fibrillatory conduction.²³ We also found that the spatial distribution of action potential duration at a low pacing rate of 3.3 Hz was different from the distribution of DF domains, which led us to suggest that dispersion of refractoriness at normal frequencies is a poor predictor of the spatial distribution of intermittent block patterns that characterize AF outside the source region.²³

Reentrant activity during acute AF in the isolated sheep heart

The notion that a localized source of reentrant activity could maintain AF was first postulated by Lewis⁸ in the early part of the twentieth century and subsequently by Scherf.²⁴ Many years later, Morillo et al targeted ablation

to sites of short cycle length activity in the posterior LA and observed the termination of arrhythmia in a canine model of AF.¹³ Using a sterile pericarditis model, Kumagai et al identified in the septum dominant unstable reentrant circuits of very short CL that maintained AF and could be successfully terminated by focal ablation.^{25,26} Others have reported that in some patients sustained focal activity at the PVs, coronary sinus (CS) or superior vena cava initiated and maintained AF, and could be eliminated by discrete ablation.²⁷ However, to this date, whether such sites are either automatic, triggered or reentrant and whether changes in the driver activity would alter spatial frequency gradients, remains unresolved.

The general working hypothesis that acute AF results from activity of a small number of high frequency reentrant sources localized in one atrium, with fibrillatory conduction to the other atrium is based primarily on results obtained in the isolated, Langendorff-perfused, sheep heart in the presence of acetylcholine (ACh).¹⁷ We localized the sources that maintain AF by a combined use of optical mapping and frequency analysis. Figure 3 shows data from on the rotation frequency of rotors during AF episodes in which the site of the high-frequency periodic activity was localized.¹⁷ In Figure 3A, an isochrone map from the LA shows one cycle of a vortex that rotated clockwise at a period of 68.6 ± 8.9 ms (~ 14.7 Hz) and persisted for the entire length of the episode (25 min). The fact that the frequency of this reentrant source (rotor) was equal to the highest and narrowest DF peak (most regular signal) recorded from all other atrial sites (both optical and electrical recordings; see Mandapati et al¹⁷ for details) provides direct evidence that it was the mechanism underlying AF in this episode.¹²

Mapping an isolated canine atrial preparation, Schuessler et al²⁸ found that with increasing concentrations

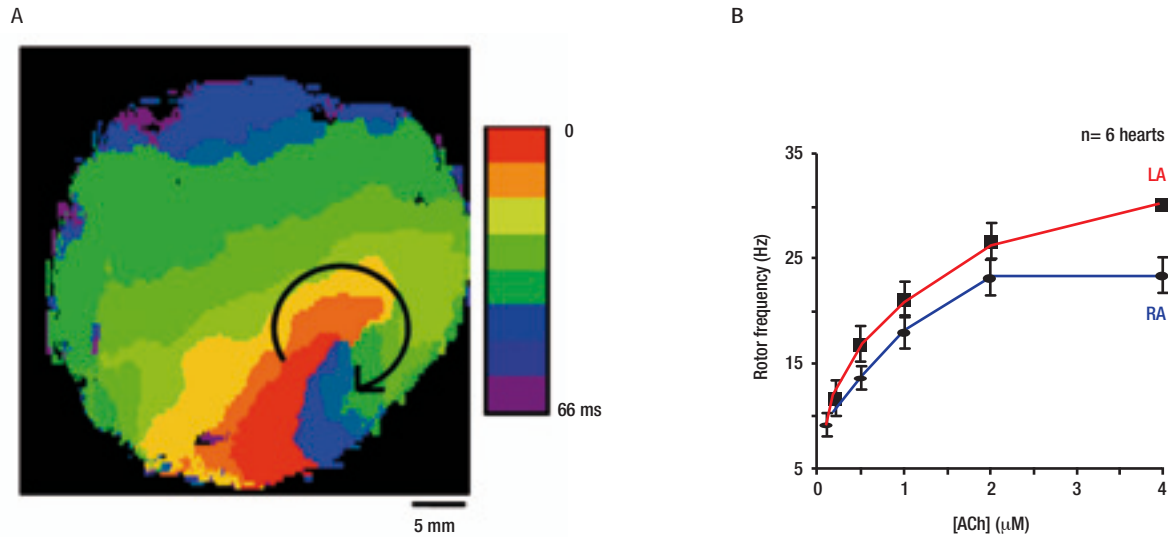


Figure 3. Rotor activity in AF and ACh concentration. A, Isochrone map of optical activity shows a microreentrant AF driver localized to the LA appendage in the form of a rotor rotating clockwise in the presence of ACh. Reproduced from Mandapati et al, *Circulation* 2000;101:194-199.¹⁷ B, Relationship between ACh concentration and rotor frequency created using frequencies from the 5 longest living rotors for each of 6 experiments, at each ACh concentration. There is an increase in rotor frequency with ACh, as well as a LA predominance over the RA. Reproduced from Sarmast et al, *Cardiovasc Res* 2003;59:863-873.²⁹

of ACh, activation patterns characterized by multiple re-entrant circuits converted to a single, relatively stable, high-frequency re-entrant circuit that resulted in fibrillatory conduction. More recently a systematic evaluation of the effect of ACh on the frequency of rotation in isolated sheep hearts was performed.²⁹ Figure 3B demonstrates the direct ACh concentration-dependence of rotor frequency in both atria of 6 hearts, with frequency being significantly higher in the LA than RA at all ACh concentrations. Moreover, the LA-to-RA rotor frequency gradient also increased with ACh concentration from 1.0 Hz at 0.1 μM ACh to 6.2 Hz at 4.0 μM ACh.

An ionic mechanism underlying the DF distribution

The data presented in Figure 3 suggest that different responses of the rotors frequency in the LA and RA to ACh during AF are somehow related to the ACh-modulated potassium current, $I_{K,ACh}$. A numerical simulation study using canine atrial cells with realistic ionic and coupling properties, including vagal actions that were formulated based on patch-clamp studies of ACh effects³⁰ showed that $I_{K,ACh}$ is a determinant of frequency and stability of rotors during AF. Nevertheless, that study did not account for the differential levels of that current in the two atria. To provide a more definite evidence for such contention we studied the effects of ACh (0.05, 0.1 and 0.5 μM) on $I_{K,ACh}$ density of sheep LA and RA myocytes.²⁹ We did not find any significant differences in $I_{K,ACh}$ density at 0.05 μM (not shown), however $I_{K,ACh}$ was significantly higher in the LA than in the RA at 0.1 and 0.5 μM. Figure 4 shows ramp-generated current density-voltage relationships in the LA and RA. In Figure 4A, the mean current density-voltage relationship at 0.1 μM ACh shows that inward (at -100 mV)

and peak outward (at -40 mV) current densities were significantly higher in LA than RA ($p < 0.05$). Figure 4B shows mean $I_{K,ACh}$ -voltage relationships at 0.5 mM ACh. Clearly, the LA has a higher density than the RA also at this concentration of ACh in both inward and outward currents ($p < 0.05$). Also, note that at the higher ACh concentration, there was an apparent reduction in current rectification. As the strong inward rectifier potassium current, I_{K1} , was found to underlie DF gradients during ventricular fibrillation in the guinea pigs,³¹ we also investigated its heterogeneity in across the sheep atria. However, additional set of voltage-clamp ramp measurements in the sheep atria found I_{K1} density to be similar in LA and RA cells.

Taken together, the patch-clamp results support the hypothesis that in the sheep heart, LA myocytes can adapt to higher excitation frequencies than RA myocytes. We further studied those findings using a simplified in-silico 2D model of the atria that was used to provide mechanistic insight into the effects of ACh on the maximal DF and LA-to-RA DF gradient by the activation of $I_{K,ACh}$.³² We used a 5x5 cm² model with realistic human atrial kinetics³³ with heterogeneous $I_{K,ACh}$ density³⁰ and an increased ACh concentration from 0.003 to 0.1 μM, simulating the transition from baseline to increased vagal tone. The model was divided in two whereby one half, representing the LA, was assigned 2 and 1.6 fold the maximal conductance for $I_{K,ACh}$ and I_{Kr} of the other half, respectively.^{29,34} Figure 5 shows consecutive snapshots during a simulation of paroxysmal AF at baseline (Figure 5A) and at increased ACh concentration (Figure 5B). In A, stable reentry in the left half of the sheet acted as the high frequency source (a; DF = 9 Hz) of waves that propagated toward the right (b; DF = 4.4 Hz). In Panel B, $I_{K,ACh}$ activation increased the rotation frequency and, consequently, the DFs in the left (15.9 Hz) and right atria (9 Hz). When conditions of persistent AF

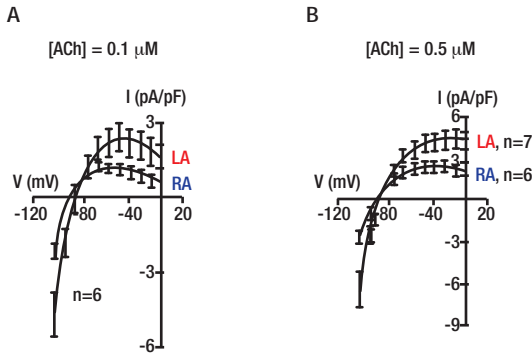


Figure 4. $I_{K,ACh}$ density in sheep atrial cells. **A**, V-clamp ramp generated currents in the presence of $0.1 \mu\text{M}$ ACh. Shown are current density in left (LA, $n=6$) and right (RA, $n=6$) atrial cells. **B**, Current density in left (LA, $n=7$) and right (RA, $n=6$) atrial cells in the presence of $0.5 \mu\text{M}$ ACh. LA $I_{K,ACh}$ density is larger than the RA at both concentrations ($p < 0.05$). Note the apparent reduction in current rectification at the higher ACh concentration. Reproduced from Sarmast et al., *Cardiovasc Res* 2003;59:863-873.²⁹

where established and a reentrant source was simulated in the LA, $I_{K,ACh}$ activation increased the DF in the left (11 to 14.6 Hz) and unified the DFs in the RA from a range of 8.8-11 Hz during baseline to 9.8 Hz during $I_{K,ACh}$ activation. Overall, while the AF frequencies are somewhat different in the simulations than in our sheep experiments and higher than the patients described below,³² qualitatively, the numerical results demonstrate that $I_{K,ACh}$ activation increased the source region DF and consequently the left-to-right DF gradient in both paroxysmal (4.6 to 6.9 Hz) and persistent AF (maximum 2.2 to 4.8 Hz).

The computer simulations suggest that the activation at extremely fast rates by stationary rotors may be the

result of the strong repolarizing influence exerted by their core, which abbreviates the action potential duration (APD) in its proximity.³⁵ This is explained by the fact that during reentry there is a sink-to-source mismatch at the highly curved tip of the pivoting wave creating a central core with excitable but unexcited cells that remain near the resting potential. The slightly depolarized resting potential in the core results in a sustained electrotonic currents between cells in the core and cells in the immediate vicinity of the core. This current in turn drives neighboring cells towards repolarized potentials, and hence, shortens APD. As the inward rectifier current $I_{K,ACh}$ is time-independent it may be able to exert its repolarizing influence and abbreviate APD even at high rates of activity and as such to allow the acceleration of the rotors. However, with increasing distance from the core, this influence weakens and the APD progressively increases.³⁵ Consequently, the tissue close to the core achieves very fast CL's, whereas, far from the core the myocardium cannot conduct at the rate of the rotor and non-uniform (i.e. other than 1:1) conduction develops. In theory, this effect may contribute to the gradient in DFs observed in during fibrillation. As illustrated Figure 3 and in the study by Mandapati et al on AF in isolated sheep hearts¹⁷ (as well as in the study by Samie et al on ventricular fibrillation in isolated guinea pig hearts,³¹) propagation near the rotor is usually 1:1, however, at a certain distance from the rotor, intermittent conduction block and wavebreaks develop and slower DF domains are formed. Nevertheless, while this mechanism may account for the shortening of the APD and the formation of DF domains, it does not explain the consistent localization of the fastest DF domain to the LA which may be further based on dispersion of ionic properties as illustrated in Figure 4.²⁹

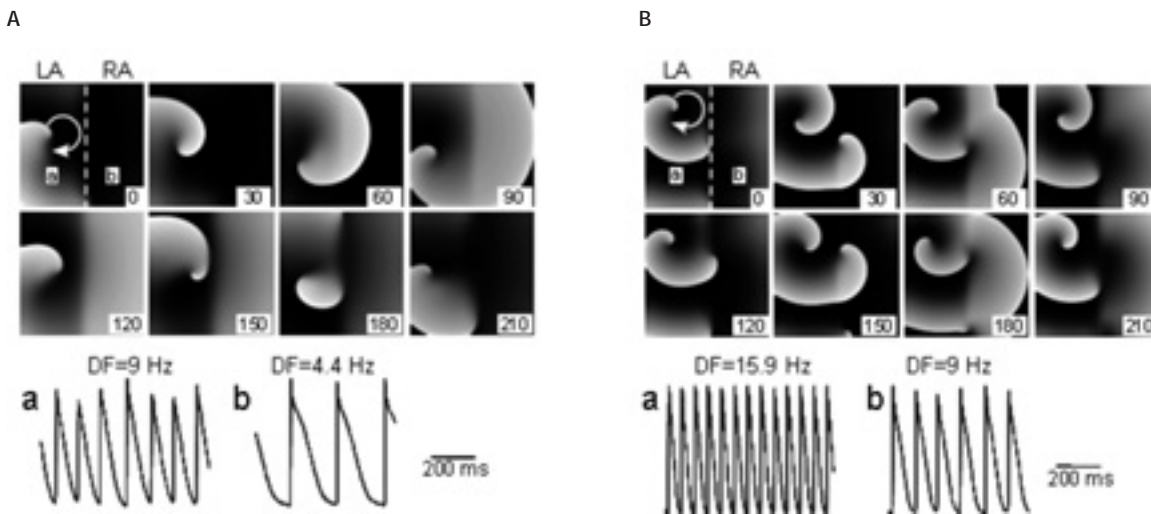


Figure 5. Computer simulations in a 2D sheet of paroxysmal AF at baseline (A) and increased $I_{K,ACh}$ (B). For each snapshot, stable reentry in the left half of the sheet acted as the high-frequency source of fibrillatory waves in the LA (a) that propagated toward the RA (b) in the right part of the sheet. At baseline ($[\text{ACh}] = 0.003 \mu\text{M}$) (A), LA DF (a; 9 Hz) is higher than RA DF (b; 4.4 Hz); left-to-right gradient is 4.6 Hz. At increased ACh ($0.1 \mu\text{M}$) (B), DFs in the LA (a) is 15.9 Hz and in RA (b) 9 Hz; the left-to-right gradient is 6.9 Hz. Numbers in frames indicate relative time in milliseconds. Reproduced from Atienza et al, *Circulation* 2006;114:2434-2442.³²

Fibrosis in experimental models of CHF sheep and cell cultures

AF is often associated with congestive heart failure (CHF), which induces extracellular matrix remodeling involving atrial fibrosis and dilatation.³⁶⁻³⁸ In addition to CHF, cardiac fibroblast proliferation and concomitant collagenous matrix accumulation (fibrosis) develop during various insults to the myocardium in ischemic, hypertensive, hypertrophic, and dilated cardiomyopathies³⁹ and arrhythmogenic right ventricular cardiomyopathy.^{40,41} Fibrosis per-se has been implicated in arrhythmia initiation and maintenance⁴² affecting electrical propagation through slow, discontinuous conduction with “zigzag” propagation,^{43,44} reduced regional coupling,⁴⁵ abrupt changes in fibrotic bundle size,⁴⁶ and micro-anatomical reentry.⁴⁷ Most clinical, experimental, and numerical studies have regarded fibroblasts and fibrosis as electrically insulating obstacles,^{48,49} although some evidence of fibroblast-to-myocyte electrical coupling was shown in the rabbit sino-atrial node.⁵⁰ However, heart injury promotes fibroblast differentiation into myofibroblasts,⁵¹ which have been shown to be coupled electrotonically to myocytes in vitro.⁵²⁻⁵⁶ Despite accumulating evidence of potential heterocellular electrical coupling in the diseased myocardium and its implications in arrhythmogenesis, the electrophysiological interplay between myofibroblasts and their neighboring myocytes has not been studied in detail. Myofibroblasts are unexcitable cells; their resting membrane potential is less negative than myocytes, and their membrane resistance is higher.^{55,57,58} These characteristics suggest that myofibroblasts may function as a sink for electrical charge and as short and long-range conductors.⁵⁹

Our experimental studies on AF included sheep with atria remodeled by ventricular tachypacing to induce CHF conditions.⁶⁰ Those experiments demonstrated that posterior LA (PLA) sources maintain AF in both normal and CHF hearts.⁶⁰ In both groups, AF waves were visualized as breakthroughs traveling from inside to outside the field of view corroborating previous experimental findings^{61,62} confirming that the PLA plays a key role in maintaining AF activity. Nevertheless the origin sites of AF breakthrough waves in CHF hearts tended to cluster peripherally, in closer vicinity to the PV ostia (PVO) compared with breakthrough waves in control. The shift in the breakthrough sites toward the periphery of the PLA in the CHF vs. control hearts coincided with histological observations demonstrating an increased fibrosis particularly near the PVO in the CHF hearts.⁶⁰ Spectral analysis revealed a significantly higher DF in the posterior LA compared with other atrial areas in both groups, however significant differences between the groups were also observed: As previously described,^{61,63,64} regional maximal DF (DF_{max}) values at other locations (PLA, Bachmann's Bundle and RA appendage) were significantly lower in the CHF group compared with the control group. The reduced local frequency of excitation in our CHF may be related to reduced conduction velocity during AF and pacing, as observed recently in mice pretreated with angiotensin II to increase fibrosis.⁶⁵ Correlation with anatomical markers of the PLA in the sheep hearts revealed that in control the DF_{max} domain extended over the left PVO in 2/6

experiments and over both left PVO and center of the PLA in 4/6 experiments. In contrast, in CHF, the DF_{max} domain localized mainly to the margin of the PLA, enclosing one or several PVO (5/7 experiments), or extended over the entire PLA (2/7 experiments). Interestingly, in all CHF experiments, the signals recorded in the DF_{max} area were more fractionated than those recorded in other areas. Conversely, the signals from the DF_{max} area in control hearts were the most regular, in agreement with another report on fractionation in the periphery of DF_{max} domains during AF.⁶⁶

Overall, the isolated sheep heart experiments demonstrated that in the PLA major changes in AF dynamics are caused by an increased amount and a different architecture of fibrosis caused by CHF.⁶⁰ Structural remodeling has been observed in both clinical and experimental AF and is an important feature of the AF substrate producing fibrosis that alters atrial tissue composition and function, however the precise mechanisms underlying AF in the setting of atrial fibrosis are not fully elucidated.⁶⁷

To characterize the effect of fibrosis separately from electrical and other structural remodeling in the myocardium we studied patterns of propagation in monolayers cultured with varying ratios in myocytes and myofibroblasts.⁶⁸ The number of myofibroblasts in the co-cultures significantly modified the spatio-temporal characteristics of reentry in the monolayers. Figure 6 shows analysis of propagation patterns and frequency of excitation using optical mapping of such monolayers. Panel A shows representative examples of phase maps during spontaneous activity in the monolayers. Each color represents a phase of the action potential; the convergence of all colors at any given location is a phase singularity (PS).⁶⁹ Clearly, the number of PSs and wavelets multiplied as the myofibroblast/myocytes ratio increased from 0.1 to 0.5 and 0.7. In panel B the complexity in the patterns of propagation is quantified by counting the number of PSs and the frequency of reentrant activity; as shown, maximal number of PSs counted in five randomly selected snapshots of a 4-sec-long phase movie (left) and rotation frequency as determined by the DFs across the monolayers (right) are plotted against myofibroblast/myocyte ratio. Each point corresponds to an individual experiment. The graphs demonstrate that as the myofibroblast/myocyte ratio increased, the number of PSs, and thus the complexity of propagation, increased, but reentry frequency decreased. These results are consistent with the results of the optical mapping in the CHF sheep hearts.⁶⁰ They suggest that fibrosis by itself is an important factor in increasing the complexity of the propagation and the slowing of the excitation frequency, independent of other ionic and structural remodeling that characterize the atria in ventricular tachypaced hearts.⁷⁰

The spatial distribution of DFs during AF in patients

Employing the local excitation frequency approach has been demonstrated to be useful in translating mechanistic insights on AF from the experimental to the clinical settings. Several recent studies have characterized the spatial distribution of DF during AF in patients.⁷¹⁻⁷³ Using

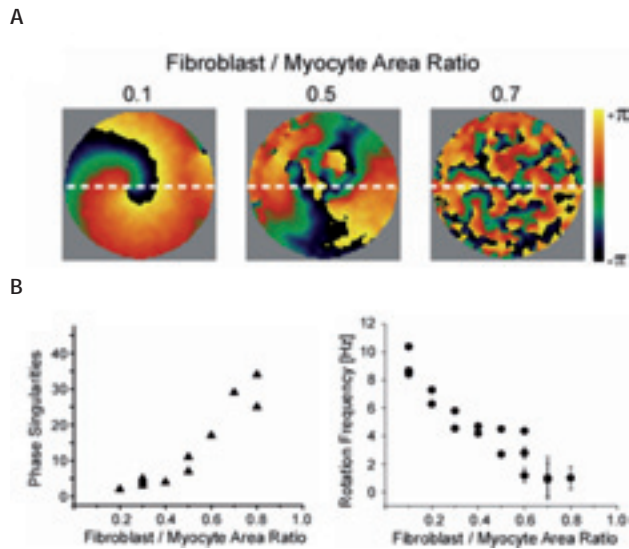


Figure 6. Optical mapping and analysis of complexity and frequency of propagation in heterocellular mixed monolayers of myocytes and fibroblasts. A, Phase maps show increasing number of singularity points with increasing number of myofibroblasts in three experiments; myofibroblast/myocyte area ratios: 0.1, 0.5, and 0.7. B, Number of PSs (left, $n=14$) and rotation frequency and (right, $n=19$) as function of myofibroblast/myocyte area ratio. Reproduced from Zlochiver et al, *Biophys J* 2008;95: 4469- 4480.⁶⁸

the CARTO electroanatomic mapping system, Sanders et al⁷³ sequentially acquired 5-sec long bipolar recordings from about 120 points throughout both atria and the CS in 32 patients during sustained AF. The recorded signals were rectified⁷⁴ and 3-15 Hz band-pass filtered, then fast Fourier transform (FFT) was used to determine the DF of each segment of 4.096 second long with a resolution of 0.24 Hz (Figure 7A).⁷⁵

In all patients, electrograms were collected from multiple sites and their corresponding DFs were superimposed on the atrial geometry to generate color DF maps,⁷³ as illustrated in Figure 7. On each DF map, those sites that demonstrated high frequency activity with a gradient of 20% or more relative to the surrounding atrial tissue were defined as high DF (HDF) sites. Figure 7B reproduces a postero-anterior view⁷³ of a DF map in a paroxysmal AF (PAF) patient. While frequency in the majority of the atria and CS was relatively slow (≤ 5 Hz), the posterior wall of the LA was activated at faster rates (7-8 Hz), with notable HDF sites at each of the PVs. Figure 7C shows a DF map from a patient with permanent AF. Compared with the patient with paroxysmal AF, this patient not only had a higher frequency at the maximal HDF site (13.7 Hz), but also both atria demonstrated higher global frequency of activity and in addition, the HDF sites were located in the atria rather in the PV region.^{32,73} We concluded that in the cohort of patients studied, paroxysmal AF was characterized by the hierarchical spatial distribution of DFs where the LA and PVs were always the fastest regions. By contrast, in persistent AF, a more uniform distribution of higher DF values was observed, where the highest DFs could not be found in the PV region.^{22,32,72,73}

The high DF sites and maintenance of AF in patients

AF mapping studies have recognized the presence of temporally and spatially periodic activity^{17,76-78} emanating from the PV region with regularity,¹⁵ suggesting that these structures may have a role in maintaining AF,¹⁷ by harboring either localized short cycle length reentrant sources and/or focal automatic activity.^{79,80} Indeed, in the patient whose DF map is presented in Figure 7B, focal radiofrequency ablation applied to the HDF site near the right inferior PV (RIPV, red arrow) effectively terminated AF.⁷³ A recent study utilizing a morphologically accurate computer model of the atria has demonstrated that the PV region is a preferential site for anchoring rotors.⁸¹ In the clinic, paroxysms of short cycle length activity have been observed in the PVs of patients undergoing AF ablation.⁸²⁻⁸⁴ In addition, sequential ablation of sites showing the shortest cycle length has been associated with a progressive slowing of AF frequency culminating in termination in 75% of patients with paroxysmal AF.²⁰

Using a blind correlation between atrial DF distribution and ablation, without any attempt at identifying potentially arrhythmogenic sites at the time of the procedure, Sanders et al⁷³ found that ablation at PVs harboring HDF sites resulted in an increase in AFCL (≥ 5 ms) within the CS in 89% of cases. The latter was true in patients with either paroxysmal or permanent AF. However, eventual arrhythmia termination occurred during ablation in 15 (88%) out of 17 patients with paroxysmal but none with permanent AF ($p<0.0001$). In 13 (87%) of the 15 paroxysmal AF patients arrhythmia termination was associated with ablation at a HDF site; 11 localized to a PV and 2 to the LA roof and the fossa ovalis. A more recent study by Atri et al⁸⁵ using a real-time mapping of DF allows targeting specifically HDF sites and further demonstrates that abolishing by ablation a preexisting LA-to-RA DF gradients predicted long term freedom of AF in both paroxysmal and persistent AF patients. In persistent AF patients, Yoshida et al⁸⁶ recently demonstrated that isolation of the PV region eliminated high DF components and transform the fibrillation into tachycardia with a slower frequency. Interestingly, for reasons which are not entirely clear the frequency of the emerging tachycardia waves had a significantly high power during the preceding fibrillation.⁸⁶ The aforementioned data, together with those of previous studies by Atri et al³² and Lazar et al,⁸⁷ clearly indicate that the HDF sites play a role in the maintenance of AF in a significant number of patients.

Activation frequency and driver mechanisms in patients

Identifying rotors as drivers of AF directly in patients is very difficult with the existing mapping techniques.^{88,89} Yet, the ACh dose dependent acceleration of rotor frequency²⁹ shown in Figure 3 offered an idea that enabled translating animal experiments to the patient and gave us the opportunity to obtain evidence, albeit indirect, for the presence of rotors as drivers through pharmacologic means. Translation was made possible also by the

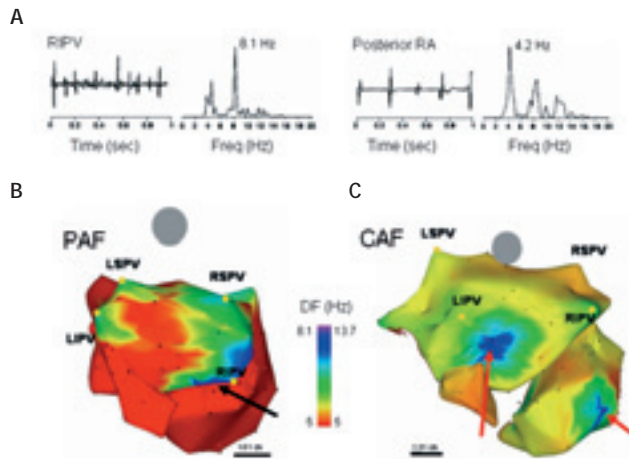


Figure 7. DF analysis in AF patients. A, Bipolar electrograms and corresponding power spectra obtained from the RIPv (left) and posterior RA in a patient with spontaneous paroxysmal AF. Each site shows distinct DF (8.1 and 4.2 Hz in RIPv and RA, respectively) demonstrating the utility of spectral analysis. B, DF map in a patient with paroxysmal AF (Posterior-anterior view; 6 hours). Note HDF sites in each of the PVs. Ablation sequence in this patient was LSPV, LIPV RSPV and RIPv (site of AF termination, black arrow); AFCL increased by 10 ms, 25 ms, 9 ms and 75 ms, respectively, before termination. C, DF map in a patient with permanent AF (24 months). The maximal DF and atrial frequency are higher than the patient in panel A. In addition, HDF sites are located outside the PVs (red arrows). Ablation sequence in this patient was RIPv, RSPV, LSPV and LIPV; AFCL increased by 5 ms, 2 ms, 0 ms and 5 ms respectively. Color-bar, DF scale in Hz. PAF: paroxysmal AF, CAF: permanent AF, LSPV, LIPV, RSPV, RIPv: Left/right superior/inferior pulmonary veins (PVs). Reproduced from Sanders et al, *Circulation* 2005;112:789-797.⁷³

fact that adenosine, which is widely used in the clinic, is known to activate the same Kir3.x subfamily of inward rectifier potassium channels as ACh.⁹⁰⁻⁹² By increasing K⁺ conductance in the atrium, both ACh and adenosine hyperpolarize the cell membrane, abbreviate the action potential duration and the refractory period, and inhibit spontaneous pacemaker discharge as well as early and delayed depolarizations.^{90,91} On the other hand they both accelerate reentrant activity.²⁹ Thus, in a recent study³² we used adenosine to test the hypothesis that localized reentry maintains AF also in humans. We determined the effects of adenosine infusion on DF at varying locations of both atria with the idea that adenosine-induced acceleration reveals reentry as the mechanism of AF maintenance and rules out an automatic or triggered mechanism. We generated baseline DF maps of the LA using novel real-time spectral analysis software that allowed determination of the specific high DF (HDF) sites likely to harbor the AF drivers⁷³ in AF patients. Then the adenosine effect was measured at the primary and secondary HDF sites in the LA. Figure 8 shows a representative example in PAF where the AF frequency at baseline was relatively slow (<5 Hz) and 3 HDF sites were identified with the primary HDF site being located near the RIPv (red arrow). Panels B and C show that while the adenosine infusion practically abolished the ventricular activity as detected by V₅, the DF at the primary HDF site accelerated from 4.64 at

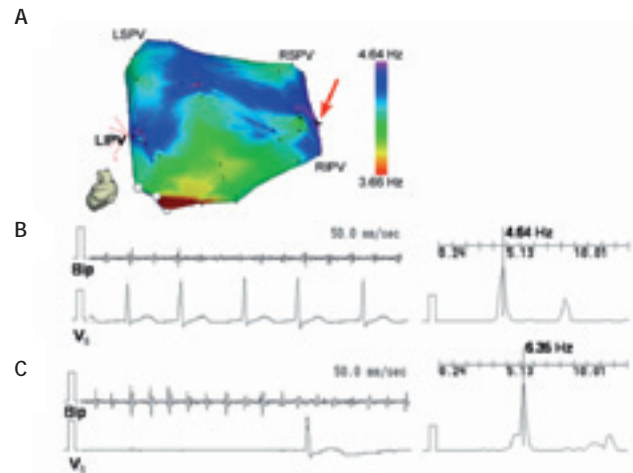


Figure 8. Accelerating effect of adenosine on HDF site. A, LA posterior view DF map from a paroxysmal AF patient. The DF map was produced by the real-time frequency-mapping CARTO system before infusion of adenosine. Red arrow indicates primary HDF site near the RIPv. B, Baseline recording at the primary HDF site with its power spectrum and simultaneous V₅ reference. C, Recording at the primary HDF site with power spectrum and simultaneous V₅ reference during peak adenosine effect showing increase of DF. LSPV, LIPV, RSPV, RIPv: Left/right superior/inferior pulmonary veins (PVs); Bip, bipolar catheter. Reproduced from Atienza et al, *Circulation* 2006;114:2434-2442.³²

baseline to 6.35 Hz at the peak of the adenosine effect. An additional adenosine infusion performed while measuring activity at a secondary HDF site also showed an increase in DF, but to a lesser extent. Interestingly, in this patient the arrhythmia terminated during post-mapping ablation at the primary HDF site, supporting again the critical role of such sites as AF drivers.⁷³ Compared to baseline, adenosine significantly accelerated the primary and secondary HDF sites in PAF patients from about 5 to 6.7 Hz,³² demonstrating that the sites involved in the maintenance of AF are affected by adenosine.

In a larger cohort of paroxysmal and persistent AF patients, Atienza et al³² analyzed the effect of adenosine on the activation rate in specific regions at the junction of the PV and the LA (PV-LAJ), the roof of the RA (HRA) and the CS. In general, patients with persistent AF demonstrated significantly higher maximal baseline DFs than paroxysmal AF patients ($p < 0.001$). However, adenosine infusion in persistent AF patients increased local DFs only in the HRA. The increase in the PV-LAJ was not statistically significant and there was no change in the CS DF. In sum, adenosine infusion increased frequency primarily at sites that activated at the highest rate at baseline. In paroxysmal AF patients, adenosine increased activation frequency in the PV-LAJ. In persistent AF patients, the highest-frequency sources accelerated by adenosine were located in either atria, but not at PV sites. Thus, the DF

increase in response to adenosine is consistent with reentrant drivers maintaining AF that have different locations in paroxysmal compared with persistent AF patients.³²

AF and fibrosis in post-operative patients

As much as 30% of patients undergoing cardiac surgery develop post-operative AF, which leads to higher morbidity, mortality, and prolonged hospitalization.^{93,94} While several factors likely contribute to postsurgical AF (e.g., stretch, inflammation, pericarditis, ischemia, autonomic influences), the exact mechanisms of initiation and maintenance are unknown. Structural remodeling of the atria by fatty infiltration, fibrosis, and iron has been implicated in the pathogenesis of chronic AF⁶⁷ and postsurgical AF.⁹⁵ Fatty infiltrates and fibrosis form barriers around myocytes creating conditions favorable for reentry.^{60,67} However, in the literature, biopsies were obtained mostly from the RA of individuals undergoing cardiac surgery^{95,96} and have demonstrated mixed results regarding the importance of preoperative structural and electrophysiological changes in patients who develop postoperative AF.⁹⁷

We recently studied 44 patients recovering from coronary artery bypass grafts (CABG) and/or valve replacement/repair surgery.⁹⁸ Of those, 31 remained in normal sinus rhythm (SR), 12 developed new-onset AF, and one developed new-onset atrial flutter. Of the 44 patients, a subset of 27 was instrumented with epicardial wires for evaluation of LA and RA DFs should they develop AF. Two temporary cardiac pacing wires were sutured onto the LA surface, and the opposing end was brought through the skin. The first recording wire was placed posteriorly at the junction of the LA and LA appendage (LAA). The second wire was placed on the posterior aspect of the LAA approximately 2 cm away from the first one. Two RA leads were placed similarly on the RA appendage and RA free wall, approximately 3 cm apart.⁹⁸ At the onset, we investigated whether the spatial organization of AF that develops spontaneously after the cardiac surgery is associated with stable LA versus RA differences in DF that persist over a 1-hour period. On average, DF values from 5-second long unipolar recordings (following removal of ventricular activity) were consistent with the DF distribution across the atria in paroxysmal AF patients.⁷³ LA DFs were significantly greater than RA DFs in all AF patients regardless of the individual LA or RA leads considered (LA: 6.29 ± 0.6 and 6.23 ± 0.6 , RA: 4.58 ± 0.7 and 4.55 ± 0.63 Hz. $p < 0.01$ LA vs. RA). The time course of the mean DF values at 10-minute intervals from the LA and RA in five patients who remained in AF for >1 hour shows the LA-to-RA DF difference throughout the 60-minute period despite a slight deceleration in the LA DF.⁹⁸

Three- to five-millimeter biopsies were obtained from both LA and RA appendages in all 44 patients during the surgery.⁹⁸ Panels A-D in **Figure 9** show micrographs of $6 \mu\text{m}$ sections from those specimen stained with picrosirius red. Those examples demonstrate the range of fibrosis seen in our samples. There was no significant difference between the LA and RA biopsies in the area of tissue available for analysis. However, the percent area that was occupied by fibrosis was higher in the LA of patients who developed AF. A statistically significant difference in LA fibrosis was found

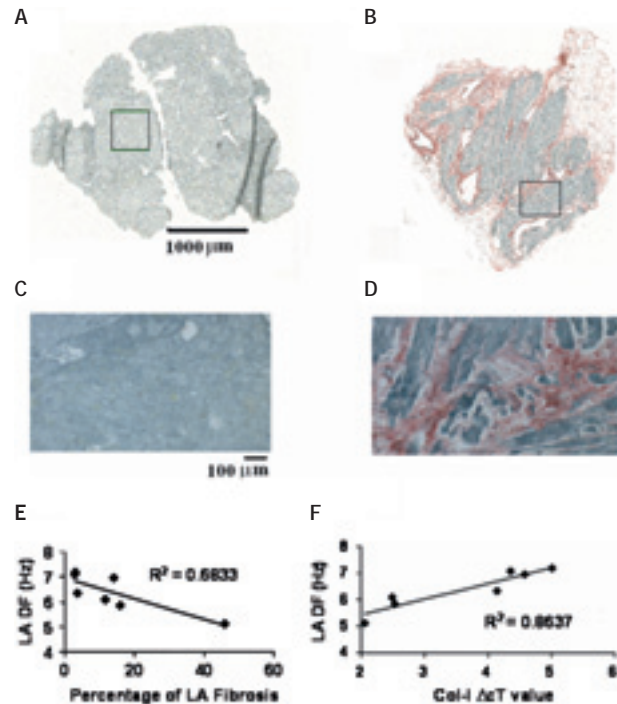


Figure 9. Fibrosis and frequency of excitation during AF in post heart surgery patients. A and B, Montage images of atrial tissue after staining with picrosirius red (fibrosis-red, muscle-green). A, LA biopsy from a 30-year-old patient who remained in SR shows no evidence of fibrosis. B, LA biopsy from a 74-year-old patient who developed AF shows 16% atrial fibrosis. C, Enlarged (x10) boxed region in A demonstrates a lack of fibrosis. D, Enlarged (x10) boxed region in B demonstrates the highest amount of fibrosis. E, Relationship between LA fibrosis and LA DF in seven postoperative AF patients. F, Relationship between LA collagen-I expression (where lower ΔcT values represent increased collagen-I expression) and LA DF in seven patients who developed postoperative AF. Reproduced from Swartz et al, Heart Rhythm 2009;6:1415-1422.⁹⁸

between those patients who developed AF and those who remained in SR. In contrast, there was no difference in RA fibrosis between these two groups as reported previously.⁹⁶ Using quantitative reverse transcription-polymerase chain reaction (RT-PCR) analysis we determined LA vs. RA differences in mRNA levels for genes coding for fibrosis.⁹⁸ We asked whether preoperative differences in atrial structure correlate with the development of postoperative AF. We found that collagen-I (a marker for fibrosis⁹⁹) expression was similar in patients who developed AF and those who remained in SR. However, in **Figure 9E-F**, examination of the impact of atrial fibrosis and collagen-I expression on the AF DF in the seven patients who developed AF and had intracardiac recordings demonstrated that an increasing percentage of LA fibrosis resulted in slower LA frequencies. This resulted in an R^2 value of 0.68 ($P = 0.02$). However, this did not remain significant after Bonferroni correction. Nevertheless, as **Figure 9F** demonstrates, when examining collagen-I expression in LA tissue there was a strong correlation between collagen-I expression and LA DF (lower ΔcT values represent increased collagen-I expression), where the R^2 value was 0.86 ($p = 0.002$), which remained significant after Bonferroni correction.

In sum, increasing amounts of LA fibrosis, particularly from collagen-I, are associated with a lower LA DF during AF after cardiac surgery. The accumulation of fibrosis has been found to modify the electrophysiologic properties of the atrium by reducing the velocity of the impulse and providing a substrate for reentry and AF initiation.^{60,99,100} The finding of reduced DF with increased presence of fibrosis is similar to the reduced frequency of reentry in monolayers with increasing myofibroblasts/myocytes area ratio. Thus, one may speculate that fibrosis in post-operative patients is sufficient to alter the dynamics of an ensuing AF independently of other remodeled factors associated, for example, with age or heart failure.

Closing remarks

The experimental and numerical studies on AF described here were conducted using sheep and simplified computer models. Obviously, results obtained from animal hearts should be cautiously extrapolated to humans. Similarly, one must be extremely cautious when attempting to generalize the applicability of theoretical concepts derived from numerical experiments. Nevertheless, as discussed above, knowledge derived from spectral analysis in the sheep atria and cell cultures may be used to link ionic and structural mechanisms of AF in experiments and patients. In general, the evidence that rotor-like activity is the driving force that maintains both atrial and ventricular fibrillation is compelling.^{17,29,31,32} and in that regard the analysis of excitation frequency arguably contributed to the recognition that time independent rectifying K^+ currents play an important role in the dynamics of rotors and AF.

Indeed, in agreement with our animal and numerical studies on rotor frequency in cholinergic AF in isolated sheep hearts presented in Figures 3-5,^{17,29} as well as our human studies,^{32,98} a recent publication describes measurement of I_{K1} and $I_{K,ACH}$ in cardiomyocytes from RA and LA atrial appendages of SR- and AF-patients undergoing cardiac surgery.¹⁰¹ In that recent study basal inward-rectifier background current was ~2-fold larger in chronic AF vs. SR patients, without RA-LA differences. In PAF, basal current was ~2-fold larger in LA vs. RA, indicating a left-to-right atrial gradient. These results support the hypothesis that a left-to-right gradient in inward-rectifier background current contributes to high-frequency reentrant sources in LA that maintain PAF.¹⁰¹ The findings on K^+ currents may allow investigators and physicians to devise new antiarrhythmic therapies that are more effective than currently available means in either preventing rotor formation, or terminating rotors after they have had time to stabilize. For instance, our numerical simulations on AF in humans demonstrate that sustained rotors in the atria depend on the conductance level the inward-rectifiers I_{K1} and $I_{K,ACH}$.^{32,102} At normal atrial condition compatible with PAF a presence of $I_{K,ACH}$ is a major player in rotor acceleration and stabilization.^{29,30,32} However, in persistent AF the increase in I_{K1} plays a further major role in the dynamics of driving rotors. By altering numerically the rectification profile of I_{K1} reentrant activity was either slowed down or abolished. Hence, one could envision that, in the foreseeable future, development of a new generation of safe and effective antifibrillatory K^+ channels modifying drugs could lead to protection against AF in patients.

An alternative approach for drug-refractory AF in patients is ablation. However this approach can be challenging in certain groups of patients.⁵ In the future, the combined use of time and frequency domain measures, including electrogram fractionation,¹⁰³ principal value decomposition and DF mapping,¹⁰⁴ should help elucidate mechanism of impulse propagation. The ability to determine whether AF is maintained by discrete sources and to identify their location may help improve the efficacy of ablation procedures. Indeed, DF analysis of bipolar recordings obtained during AF ablation is strongly suggestive of specific high DF sites representing the drivers that maintain AF in certain groups of patients.^{73,85} Thus, the identification of the drivers and characterization of their dynamics may provide an efficient mean of developing more patient-specific ablation procedures with well defined end-points^{85,105} potentially leading to increased efficacy and safety.^{106,107}

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References

1. Chen LY, Shen WK. Epidemiology of atrial fibrillation: a current perspective. *Heart Rhythm* 2007;4:S1-S6.
2. Haissaguerre M, Jais P, Shah DC, et al. Spontaneous initiation of atrial fibrillation by ectopic beats originating in the pulmonary veins. *N Engl J Med* 1998;339:659-666.
3. Haissaguerre M, Shah DC, Jais P, et al. Mapping-guided ablation of pulmonary veins to cure atrial fibrillation. *American Journal of Cardiology* 2000;86:9K-19K.
4. Moe GK, Abildskov JA. Atrial fibrillation as a self-sustaining arrhythmia independent of focal discharges. *American Heart Journal* 1959;58:59-70.
5. Oral H, Pappone C, Chugh A, et al. Circumferential pulmonary-vein ablation for chronic atrial fibrillation. *N Engl J Med* 2006;354:934-941.
6. Moe GK. On the multiple wavelet hypothesis of atrial fibrillation. *Archives Internationales de Pharmacodynamie et de Therapie* 1962;CXL:183-188.
7. Allesie MA, Lammers WJEP, Bonke FIM, et al. Experimental evaluation of Moe's wavelet hypothesis of atrial fibrillation. In: Zipes DP, Jalife J, editors. *Cardiac Electrophysiology and Arrhythmias*. Orlando: Grune & Stratton, 1985: 265-275.
8. Lewis T. *The mechanism and graphic registration of the heart beat*. 3 ed. London: Shaw & Sons, 1925.
9. Krinskii VI. Excitation propagation in nonhomogenous medium (actions analogous to heart fibrillation). *Biofizika* 1966;11:676-683.
10. Winfree AT. Suppressing drosophila circadian rhythm with dim light. *Science* 1974;183:970-972.
11. Allesie MA, Bonke FI, Schopman FJ. Circus movement in rabbit atrial muscle as a mechanism of tachycardia. III. The «leading circle» concept: a new model of circus movement in cardiac tissue without the involvement of an anatomical obstacle. *Circ Res* 1977;41:9-18.
12. Jalife J, Berenfeld O, Skanes A, et al. Mechanisms of atrial fibrillation: mother rotors or multiple daughter wavelets, or both? *J Cardiovasc Electrophysiol* 1998;9:S2-S12.
13. Morillo CA, Klein GJ, Jones DL, et al. Chronic rapid atrial pacing: Structural, functional, and electrophysiological characteristics of a new model of sustained atrial fibrillation. *Circulation* 1995;91:1588-1595.
14. Harada A, Sasaki K, Fukushima T, et al. Atrial activation during chronic atrial fibrillation in patients with isolated mitral valve

- disease. *Ann Thorac Surg*. 1996;61:104-112.
15. Skanes AC, Mandapati R, Berenfeld O, et al. Spatiotemporal periodicity during atrial fibrillation in the isolated sheep heart. *Circulation* 1998;98:1236-1248.
 16. Berenfeld O, Mandapati R, Dixit S, et al. Spatially distributed dominant excitation frequencies reveal hidden organization in atrial fibrillation in the Langendorff-perfused sheep heart. *J Cardiovasc Electrophysiol* 2000;11:869-879.
 17. Mandapati R, Skanes A, Chen J, Berenfeld O, Jalife J. Stable microreentrant sources as a mechanism of atrial fibrillation in the isolated sheep heart. *Circulation* 2000;101:194-199.
 18. Mansour M, Mandapati R, Berenfeld O, et al. Left-to-right gradient of atrial frequencies during acute atrial fibrillation in the isolated sheep heart. *Circulation* 2001;103:2631-2636.
 19. Pappone C, Rosanio S. Pulmonary Vein Isolation for Atrial Fibrillation. In: Zipes DP, Jalife J, editors. *Cardiac Electrophysiology - From Cell to Bedside*. Philadelphia: Saunders, 2004: 1039-1052.
 20. Haissaguerre M, Sanders P, Hocini M, et al. Changes in atrial fibrillation cycle length and inducibility during catheter ablation and their relation to outcome. *Circulation* 2004;109:3007-3013.
 21. Berenfeld O. Quantifying activation frequency in atrial fibrillation to establish underlying mechanisms and ablation guidance. *Heart Rhythm*. 2007;4:1225-1234.
 22. Schuessler RB, Kay MW, Melby SJ, et al. Spatial and temporal stability of the dominant frequency of activation in human atrial fibrillation. *J Electrocardiol*. 2006.
 23. Berenfeld O, Zaitsev AV, Mironov SF, et al. Frequency-dependent breakdown of wave propagation into fibrillatory conduction across the pectinate muscle network in the isolated sheep right atrium. *Circ Res* 2002;90:1173-1180.
 24. Scherf D. Studies on auricular tachycardia caused by aconitine administration. *Proc Soc Exp Biol Med* 1947;64:233-239.
 25. Kumagai K, Khrestian C, Waldo AL. Simultaneous multisite mapping studies during induced atrial fibrillation in the sterile pericarditis model. Insights into the mechanisms of its maintenance. *Circulation* 1997;95:511-521.
 26. Kumagai K, Uno K, Khrestian C, et al. Single site radiofrequency catheter ablation of atrial fibrillation: Studies guided by simultaneous multisite mapping in the canine sterile pericarditis model. *Journal of the American College of Cardiology*. 2000;36:917-923.
 27. Jais P, Haissaguerre M, Shah DC, et al. A focal source of atrial fibrillation treated by discrete radiofrequency ablation. *Circulation* 1997;95:572-576.
 28. Schuessler RB, Grayson TM, Bromberg BI, et al. Cholinergically mediated tachyarrhythmias induced by a single extrastimulus in the isolated canine right atrium. *Circ Res* 1992;71:1254-1267.
 29. Sarmast F, Kolli A, Zaitsev A, et al. Cholinergic atrial fibrillation: I-K, I-ACh gradients determine unequal left/right atrial frequencies and rotor dynamics. *Cardiovascular Research*. 2003;59:863-873.
 30. Kneller J, Zou R, Vigmond EJ, et al. Cholinergic atrial fibrillation in a computer model of a two-dimensional sheet of canine atrial cells with realistic ionic properties. *Circ Res* 2002;90:E73-E87.
 31. Samie FH, Berenfeld O, Anumonwo J, et al. Rectification of the Background Potassium Current: A Determinant of Rotor Dynamics in Ventricular Fibrillation. *Circ Res* 2001;89:1216-1223.
 32. Atienza F, Almendral J, Moreno J, et al. Activation of inward rectifier potassium channels accelerates atrial fibrillation in humans: evidence for a reentrant mechanism. *Circulation* 2006;114:2434-2442.
 33. Courtemanche M, Ramirez RJ, Nattel S. Ionic mechanisms underlying human atrial action potential properties: insights from a mathematical model. *American Journal of Physiology-Heart and Circulatory Physiology* 1998;44:H301-H321.
 34. Li D, Zhang L, Kneller J, et al. Potential ionic mechanism for repolarization differences between canine right and left atrium. *Circ Res* 2001;88:1168-1175.
 35. Beaumont J, Jalife J. Rotors and spiral waves in two dimensions. In: Zipes DP, Jalife J, editors. *Cardiac Electrophysiology From Cell to Bedside*. Philadelphia, PA: W.B. Saunders, 2000:327-335.
 36. Lubitz SA, Benjamin EJ, Ellinor PT. Atrial fibrillation in congestive heart failure. *Heart Fail Clin* 2010;6:187-200.
 37. Shelton RJ, Clark AL, Kaye GC, et al. The atrial fibrillation paradox of heart failure. *Congest Heart Fail* 2010;16:3-9.
 38. Nattel S. Driver regions in atrial fibrillation associated with congestive heart failure: where are they, and what are they telling us? *J Cardiovasc Electrophysiol* 2005;16:1359-1361.
 39. Manabe I, Shindo T, Nagai R. Gene expression in fibroblasts and fibrosis: involvement in cardiac hypertrophy. *Circ Res* 2002;91:1103-1113.
 40. Brown RD, Ambler SK, Mitchell MD, et al. The cardiac fibroblast: therapeutic target in myocardial remodeling and failure. *Annu Rev Pharmacol Toxicol* 2005;45:657-687.
 41. Basso C, Ronco F, Marcus F, et al. Quantitative assessment of endomyocardial biopsy in arrhythmogenic right ventricular cardiomyopathy/dysplasia: an in vitro validation of diagnostic criteria. *Eur Heart J* 2008;29:2760-2771.
 42. Rohr S. Myofibroblasts in diseased hearts: new players in cardiac arrhythmias? *Heart Rhythm* 2009;6:848-856.
 43. de Bakker JM, van Capelle FJ, Janse MJ, et al. Slow conduction in the infarcted human heart. 'Zigzag' course of activation. *Circulation* 1993;88:915-926.
 44. Bian W, Tung L. Structure-related initiation of reentry by rapid pacing in monolayers of cardiac cells. *Circ Res* 2006;98:e29-e38.
 45. Spach MS, Boineau JP. Microfibrosis produces electrical load variations due to loss of side-to-side cell connections: a major mechanism of structural heart disease arrhythmias. *Pacing Clin Electrophysiol* 1997;20:397-413.
 46. de Bakker JM, Stein M, van Rijen HV. Three-dimensional anatomic structure as substrate for ventricular tachycardia/ventricular fibrillation. *Heart Rhythm*. 2005;2:777-779.
 47. Valderrabano M, Kim YH, Yashima M, et al. Obstacle-induced transition from ventricular fibrillation to tachycardia in isolated swine right ventricles: insights into the transition dynamics and implications for the critical mass. *J Am Coll Cardiol* 2000;36:2000-2008.
 48. Jacquemet V, Henriquez CS. Genesis of complex fractionated atrial electrograms in zones of slow conduction: a computer model of microfibrosis. *Heart Rhythm* 2009;6:803-810.
 49. Jacquemet V, Henriquez CS. Modulation of conduction velocity by nonmyocytes in the low coupling regime. *IEEE Trans Biomed Eng* 2009;56:893-896.
 50. Camelliti P, Green CR, LeGrice I, et al. Fibroblast network in rabbit sinoatrial node: structural and functional identification of homogeneous and heterogeneous cell coupling. *Circ Res* 2004;94:828-835.
 51. Weber KT. The Dead Sea lives! Someone's rockin' my dreamboat. *Cardiovasc Res* 1995;29:604-610.
 52. Camelliti P, Borg TK, Kohl P. Structural and functional characterization of cardiac fibroblasts. *Cardiovasc Res* 2005;65:40-51.
 53. Gaudesius G, Miragoli M, Thomas SP, et al. Coupling of cardiac electrical activity over extended distances by fibroblasts of cardiac origin. *Circ Res* 2003;93:421-428.
 54. Miragoli M, Gaudesius G, Rohr S. Electrotonic modulation of cardiac impulse conduction by myofibroblasts. *Circ Res* 2006;98:801-810.
 55. Rook MB, van Ginneken AC, de Jonge B, et al. Differences in gap junction channels between cardiac myocytes, fibroblasts, and heterologous pairs. *Am J Physiol* 1992;263:C959-C977.
 56. Goshima K. Formation of nexuses and electrotonic transmission between myocardial and FL cells in monolayer culture.

- Exp Cell Res 1970;63:124-130.
57. Kamkin A, Kiseleva I, Wagner KD, et al. Mechanically induced potentials in fibroblasts from human right atrium. *Exp Physiol* 1999;84:347-356.
 58. Kohl P, Kamkin AG, Kiseleva IS, et al. Mechanosensitive fibroblasts in the sino-atrial node region of rat heart: interaction with cardiomyocytes and possible role. *Exp Physiol* 1994;79:943-956.
 59. Kohl P, Camelliti P, Burton FL, et al. Electrical coupling of fibroblasts and myocytes: relevance for cardiac propagation. *J Electrocardiol.* 2005;38:45-50.
 60. Tanaka K, Zlochiver S, Vikstrom KL, et al. Spatial distribution of fibrosis governs fibrillation wave dynamics in the posterior left atrium during heart failure. *Circ Res* 2007;101:839-847.
 61. Fenelon G, Shepard RK, Stambler BS. Focal origin of atrial tachycardia in dogs with rapid ventricular pacing-induced heart failure. *J Cardiovasc Electrophysiol* 2003;14:1093-1102.
 62. Ryu K, Shroff SC, Sahadevan J, et al. Mapping of atrial activation during sustained atrial fibrillation in dogs with rapid ventricular pacing induced heart failure: evidence for a role of driver regions. *J Cardiovasc Electrophysiol* 2005;16:1348-1358.
 63. Okuyama Y, Miyachi Y, Park AM, et al. High resolution mapping of the pulmonary vein and the vein of Marshall during induced atrial fibrillation and atrial tachycardia in a canine model of pacing-induced congestive heart failure. *J Am Coll Cardiol* 2003;42:348-260.
 64. Stambler BS, Fenelon G, Shepard RK, et al. Characterization of sustained atrial tachycardia in dogs with rapid ventricular pacing-induced heart failure. *J Cardiovasc Electrophysiol* 2003;14:499-507.
 65. Rudolph V, Andrie RP, Rudolph TK, et al. Myeloperoxidase acts as a profibrotic mediator of atrial fibrillation. *Nat Med* 2010;16:470-474.
 66. Kalifa J, Tanaka K, Zaitsev AV, et al. Mechanisms of wave fractionation at boundaries of high-frequency excitation in the posterior left atrium of the isolated sheep heart during atrial fibrillation. *Circulation* 2006;113:626-633.
 67. Burstein B, Nattel S. Atrial fibrosis: mechanisms and clinical relevance in atrial fibrillation. *J Am Coll Cardiol* 2008;51:802-809.
 68. Zlochiver S, Munoz V, Vikstrom KL, et al. Electrotonic myofibroblast-to-myocyte coupling increases propensity to reentrant arrhythmias in two-dimensional cardiac monolayers. *Biophys J* 2008;95:4469-4480.
 69. Gray RA, Pertsov AM, Jalife J. Spatial and temporal organization during cardiac fibrillation. *Nature* 1998;392:75-78.
 70. Li D, Melnyk P, Feng J, et al. Effects of experimental heart failure on atrial cellular and ionic electrophysiology. *Circulation* 2000;101:2631-2638.
 71. Lazar S, Dixit S, Marchlinski FE, et al. Presence of left-to-right atrial frequency gradient in paroxysmal but not persistent atrial fibrillation in humans. *Circulation* 2004;110:3181-3186.
 72. Sahadevan J, Ryu K, Peltz L, et al. Epicardial Mapping of Chronic Atrial Fibrillation in Patients: Preliminary Observations. *Circulation* 2004;110:3293-3299.
 73. Sanders P, Berenfeld O, Hocini M, et al. Spectral analysis identifies sites of high-frequency activity maintaining atrial fibrillation in humans. *Circulation* 2005;112:789-797.
 74. Botteron GW, Smith JM. Quantitative assessment of the spatial organization of atrial fibrillation in the intact human heart. *Circulation* 1996;93:513-518.
 75. Fischer G, Stuhlinger MC, Nowak CN, et al. On computing dominant frequency from bipolar intracardiac electrograms. *IEEE Trans Biomed Eng* 2007;54:165-169.
 76. Wu TJ, Doshi RN, Huang HLA, et al. Simultaneous biatrial computerized mapping during permanent atrial fibrillation in patients with organic heart disease. *J Cardiovasc Electrophysiol* 2002;13:571-577.
 77. Sih HJ, Zipes DP, Barbari EJ, et al. Differences in organization between acute and chronic atrial fibrillation in dogs. *Journal of the American College of Cardiology* 2000;36:924-931.
 78. Wu TJ, Ong JJC, Chang CM, et al. Pulmonary veins and ligament of Marshall as sources of rapid activations in a canine model of sustained atrial fibrillation. *Circulation* 2001;103:1157-1163.
 79. Arora R, Verheule S, Scott L, et al. Arrhythmogenic substrate of the pulmonary veins assessed by high-resolution optical mapping. *Circulation* 2003;107:1816-1821.
 80. Kalifa J, Jalife J, Zaitsev AV, et al. Intra-atrial pressure increases rate and organization of waves emanating from the superior pulmonary veins during atrial fibrillation. *Circulation* 2003;108:668-671.
 81. Vigmond EJ, Tsoi V, Kuo S, et al. The effect of vagally induced dispersion of action potential duration on atrial arrhythmogenesis. *Heart Rhythm* 2004;1:334-344.
 82. Kumagai K, Yasuda T, Tojo H, et al. Role of rapid focal activation in the maintenance of atrial fibrillation originating from the pulmonary veins. *Pacing Clin Electrophysiol* 2000;23:1823-1827.
 83. O'Donnell D, Furniss SS, Bourke JP. Paroxysmal cycle length shortening in the pulmonary veins during atrial fibrillation correlates with arrhythmogenic triggering foci in sinus rhythm. *J Cardiovasc Electrophysiol* 2002;13:124-128.
 84. Oral H, Ozaydin M, Tada H, et al. Mechanistic significance of intermittent pulmonary vein tachycardia in patients with atrial fibrillation. *J Cardiovasc Electrophysiol* 2002;13:645-650.
 85. Atienza F, Almendral J, Jalife J, et al. Real-time dominant frequency mapping and ablation of dominant frequency sites in atrial fibrillation with left-to-right frequency gradients predicts long-term maintenance of sinus rhythm. *Heart Rhythm.* 2009;6:33-40.
 86. Yoshida K, Chugh A, Ulfarsson M, et al. Relationship between the spectral characteristics of atrial fibrillation and atrial tachycardias that occur after catheter ablation of atrial fibrillation. *Heart Rhythm* 2009;6:11-17.
 87. Lazar S, Dixit S, Callans DJ, et al. Effect of pulmonary vein isolation on the left-to-right atrial dominant frequency gradient in human atrial fibrillation. *Heart Rhythm* 2006;3:889-895.
 88. Sanders P, Hocini M, Jais P, et al. Characterization of focal atrial tachycardia using high-density mapping. *J Am Coll Cardiol* 2005;46:2088-2099.
 89. Hsu LF, Jais P, Hocini M, et al. High-density circumferential pulmonary vein mapping with a 20-pole expandable circular mapping catheter. *Pacing Clin Electrophysiol* 2005;28 Suppl 1:S94-S98.
 90. Kabell G, Buchanan LV, Gibson JK, et al. Effects of adenosine on atrial refractoriness and arrhythmias. *Cardiovasc Res* 1994;28:1385-1389.
 91. Belardinelli L, Shryock JC, Song Y, et al. Ionic basis of the electrophysiological actions of adenosine on cardiomyocytes. *FASEB Journal* 1995;9:359-365.
 92. Khositseth A, Clapham DE, Ackerman MJ. Intracellular signaling and regulation of cardiac ion channels. In: Zipes DP, Jalife J, editors. *Cardiac Electrophysiology - From Cell to Bedside*. Philadelphia, PA: W.B. Saunders, 2004: 33-41.
 93. McKeown PP, Gutterman D. Executive summary: American College of Chest Physicians guidelines for the prevention and management of postoperative atrial fibrillation after cardiac surgery. *Chest* 2005;128:15-55.
 94. Mathew JP, Parks R, Savino JS, et al. Atrial fibrillation following coronary artery bypass graft surgery: predictors, outcomes, and resource utilization. MultiCenter Study of Perioperative Ischemia Research Group. *JAMA* 1996;276:300-306.
 95. Nakai T, Chandu J, Nakai K, et al. Histologic assessment of right atrial appendage myocardium in patients with atrial fibrillation after coronary artery bypass graft surgery. *Cardiology* 2007;108:90-96.
 96. Cosgrave J, Foley JB, Flavin R, et al. Preoperative atrial histological changes are not associated with postoperative atrial

- fibrillation. *Cardiovasc Pathol.* 2006;15:213-217.
97. Workman AJ, Kane KA, Rankin AC. Cellular bases for human atrial fibrillation. *Heart Rhythm* 2008;5:S1-S6.
 98. Swartz MF, Fink GW, Lutz CJ, et al. Left versus right atrial difference in dominant frequency, K(+) channel transcripts, and fibrosis in patients developing atrial fibrillation after cardiac surgery. *Heart Rhythm* 2009;6:1415-1422.
 99. Weber KT, Pick R, Jalil JE, et al. Patterns of myocardial fibrosis. *J Mol Cell Cardiol.* 1989;21 Suppl 5:121-131.
 100. Anyukhovskiy EP, Sosunov EA, Plotnikov A, et al. Cellular electrophysiologic properties of old canine atria provide a substrate for arrhythmogenesis. *Cardiovasc Res* 2002;54:462-469.
 101. Voigt N, Trausch A, Knaut M, et al. Left-to-Right Atrial Inward-Rectifier Potassium Current Gradients in Patients with Paroxysmal Versus Chronic Atrial Fibrillation. *Circ Arrhythm Electrophysiol* 2010.
 102. Pandit SV, Berenfeld O, Anumonwo J, et al. Ionic Determinants of Functional Reentry in a 2-D Model of Human Atrial Cells During Simulated Chronic Atrial Fibrillation. *Biophys J* 2005.
 103. Nademanee K, McKenzie J, Kosar E, et al. A new approach for catheter ablation of atrial fibrillation: Mapping of the electrophysiologic substrate. *Journal of the American College of Cardiology* 2004;43:2044-2053.
 104. Zlochiver S, Yamazaki M, Kalifa J, et al. Rotor meandering contributes to irregularity in electrograms during atrial fibrillation. *Heart Rhythm* 2008;5:846-854.
 105. Yoshida K, Chugh A, Good E, et al. A critical decrease in dominant frequency and clinical outcome after catheter ablation of persistent atrial fibrillation. *Heart Rhythm* 2010;7:295-302.
 106. Bertaglia E, Zoppo F, Tondo C, et al. Early complications of pulmonary vein catheter ablation for atrial fibrillation: A multicenter prospective registry on procedural safety. *Heart Rhythm* 2007;4:1265-1271.
 107. Cappato R, Calkins H, Chen SA, et al. Worldwide survey on the methods, efficacy, and safety of catheter ablation for human atrial fibrillation. *Circulation* 2005;111:1100-1105.