



Novel information about the bioactive compound profile, antioxidant and antiproliferative potential of *Parkinsonia praecox* (Fabaceae)

Nueva información del perfil de compuestos bioactivos, potencial antioxidante y antiproliferativo de *Parkinsonia praecox* (Fabaceae)

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Abstract:

Background and Aims: *Parkinsonia praecox*, “palo brea”, is a medicinal plant distributed in the Mexican Sonoran Desert. However, there is little information about the chemical composition and biological potential of this plant. As a consequence, the objective of the present study was to determine the chemical composition, as well as the antioxidant and antiproliferative activity, of *P. praecox*.

Methods: Methanolic extracts of stems (PPS), berries (PPB) and flowers (PPF) were performed. Chemical composition was determined by phytochemical screening, Folin Ciocalteu and UPLC-DAD methods. Antioxidant activity was determined by DPPH, ABTS, ORAC and FRAP methods. Antiproliferative activity was evaluated by MTT against A549 (non-small-cell lung cancer cells), MDA-MB-231 (triple negative breast cancer), PC-3 (adenocarcinoma prostate cancer grade IV), HeLa (human cervical cancer) and L929 (non-cancerous subcutaneous connective tissue) cell lines.

Results: Phytochemical screening showed the presence of terpenes, phenolic compounds, flavonoids, tannins and sugars in the extracts. PPS showed the highest ($p<0.05$) concentration of phenolic compounds (65.5 mg GAE/g), identifying and quantifying quercetin (218.86 μ g/g). Additionally, PPS exhibited the highest capacity ($p<0.05$) to stabilize the DPPH (IC_{50} : 137 μ g/ml), ABTS (39.56 μ M TE/g), hydroxyl radicals (ORAC: 1777.78 μ M TE/g), and to reduce metals (FRAP: 935.6 μ M Fe(II)/g). Similar behavior was observed in antiproliferative activity, since PPS presented the highest cytotoxicity ($p<0.05$): A549 (IC_{50} : 341.3 μ g/ml), MDA-MB-231 (IC_{50} : 147.3 μ g/ml), PC-3 (IC_{50} : 78.8 μ g/ml), HeLa (IC_{50} : 121.6 μ g/ml) and L929 (IC_{50} : 93.29 μ g/ml).

Conclusion: This is the first research where the bioactive compound profile and the biological potential of *P. praecox* are reported. The results show the strong association between the antioxidant and the antiproliferative activities with the presence of phenolic compounds. This represents a potential support for the development of pharmacological therapies.

Key words: biological activities, bioactive compounds, Sonoran desert, medicinal plants.

Resumen:

Antecedentes y Objetivos: *Parkinsonia praecox*, “palo brea”, es una planta medicinal distribuida en el Desierto de Sonora en México. Sin embargo, existe poca información acerca de la composición química y del potencial biológico de esta planta. Basado en lo anterior, el objetivo del presente estudio fue determinar la composición química y el potencial antioxidante y antiproliferativo de *P. praecox*.

Métodos: Se prepararon extractos metanólicos de tallos (PPS), frutos (PPB) y flores (PPF). La composición química se determinó a través de los métodos de perfil fitoquímico, Folin-Ciocalteu y UPLC-DAD. La actividad antioxidante se evaluó por los métodos DPPH, ABTS, ORAC y FRAP. La actividad antiproliferativa se determinó por el ensayo MTT contra las líneas celulares A549 (cáncer de pulmón de células no pequeñas), MDA-MB-231 (cáncer de mama triple negativo), PC-3 (cáncer de próstata grado IV), HeLa (cáncer de cervix) y L929 (tejido conectivo subcutáneo no canceroso).

Resultados: El perfil fitoquímico mostró la presencia de terpenos, compuestos fenólicos, flavonoides, taninos y azúcares en los extractos. PPS presentó la concentración más elevada ($p<0.05$) de compuestos fenólicos (65.5 mg GAE/g), identificando y cuantificando quercetina (218.86 μ g/g). Además, PPS mostró la capacidad más elevada ($p<0.05$) para estabilizar a los radicales DPPH (IC_{50} : 137 μ g/ml), ABTS (39.56 μ M TE/g), hidroxilo (ORAC: 1777.78 μ M TE/g), y reducir metales (FRAP: 935.6 μ M Fe(II)/g). Un comportamiento similar se observó en la actividad antiproliferativa, ya que PPS mostró la citotoxicidad más elevada ($p<0.05$): A549 (IC_{50} : 341.3 μ g/ml), MDA-MB-231 (IC_{50} : 147.3 μ g/ml), PC-3 (IC_{50} : 78.8 μ g/ml), HeLa (IC_{50} : 121.6 μ g/ml) y L929 (IC_{50} : 93.29 μ g/ml).

Conclusión: Este es el primer estudio donde se reporta el perfil de compuestos bioactivos y el potencial biológico de *P. praecox*. Los resultados muestran una fuerte asociación entre la actividad antioxidante y antiproliferativa con la presencia de compuestos fenólicos. Esto representa un potencial soporte para el desarrollo de terapias farmacológicas.

Palabras clave: actividades biológicas, compuestos bioactivos, Desierto de Sonora, plantas medicinales.

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Introduction

For thousands of years, plants from the Sonoran Desert in Northwestern Mexico have adapted to survive the hostile climatic conditions, including long droughts (Sánchez-Escalante, 2007b). The number of plant species in this place is higher than in temperate ecosystems (Sánchez-Escalante, 2007b), being considered a subtropical desert that promotes the development of a great number of plants with pharmacological potential, as evidenced by research carried out in the last years (Torres-Moreno et al., 2015; Vidal Gutiérrez et al., 2020). Among the great diversity of plants in the Sonoran Desert, we also find some species of the genus *Parkinsonia* L. such as *P. microphylla* Torr. (palo verde), *P. florida* (Benth. ex A. Gray) S. Watson (palo verde, palo azul), *P. aculeata* L. (bagote, bacaporo or guaco) and *P. praecox* (Ruiz & Pav.) Hawkins (brea, palo brea o árbol del manteco) are extensively distributed in this area (Sánchez-Escalante, 2007a).

The genus *Parkinsonia* has been studied for its biological properties. *Parkinsonia aculeata* has been recognized for its antioxidant, antimutagenic, antibacterial, antifungal, anti-inflammatory and antitumor effects (Saha et al., 2010a; Singh et al., 2013; Al-Youssef and Hassan, 2015; Sharma et al., 2016). Chemical studies on *P. aculeata* showed that this plant is constituted by essential oils (Al-Youssef and Hassan, 2015), flavonoids and polyphenolic compounds (Abdelaziz et al., 2020), with pharmacological potential. On the other hand, gold and silver nanoparticles made from leaves extracts of *P. florida* have shown antibacterial activity against *Staphylococcus aureus* Rosenbach 1884 and *Escherichia coli* (Escherich, 1885) (López-Millán et al., 2019). This suggests that *Parkinsonia* spp. are a cost-effective option for the development of new pharmaceutical formulations, due to their wide distribution throughout the American continent, their accelerated growth, as well as the large variety of bioactive compounds associated with several biological activities found in representatives of this

genus (Sánchez-Escalante, 2007a; Al-Youssef and Hassan, 2015; Abdelaziz et al., 2020; Romão and Mansano, 2021).

Parkinsonia praecox (Fabaceae) is a plant of a great interest for the variety of uses and goods it provides (Fig. 1). Traditionally, it is used as forage, construction basis and fuel. Furthermore, its vascular exudates called “gums” are commonly utilized as a substitute of arabic gum, glue, as well as in traditional medicine for treatment of bronchial conditions (Labrada-Aranda, 2005). In Argentina, *P. praecox* brea gum commonly named “woodland candy, dulce del bosque” has been used by villagers since pre-Colombian time without any harmful consequences produced in health (von Müller et al., 2009).

Studies on *P. praecox* showed that this plant has antibacterial, antifungal, and cytotoxic effect on *Artemia salina* (Linneaus 1758) (Torrealba, 2008). The toxicological evaluation showed that brea gum does not induce toxicological effects in mice when it is administered as a food additive (von Müller et al., 2009). Phytochemical analysis demonstrated that *P. praecox* is constituted by alkaloids, triterpenes, sterols, saponins, flavonoids and polyphenolics (Torrealba, 2008). However, its antiproliferative and antioxidant effect remains unexplored. In this study, with the aim to generate new information about the chemical composition and the biological potential of *P. praecox*, phytochemical screening, phenolic profile, and the antioxidant and antiproliferative effect of its stems, berries and flowers were investigated. The results reported here position *P. praecox* as a new option for the search of new compounds with antiproliferative and antioxidant potential.

Materials and Methods

Plant and extract elaboration

Parkinsonia praecox stems, berries and flowers were collected in ejido San Pablo, Sonora, Mexico, in April 2019 (28°48'43.4"N 110°57'36.8"W). Samples were identified (voucher number 102769) by Dr. José Angel Villareal Quin-

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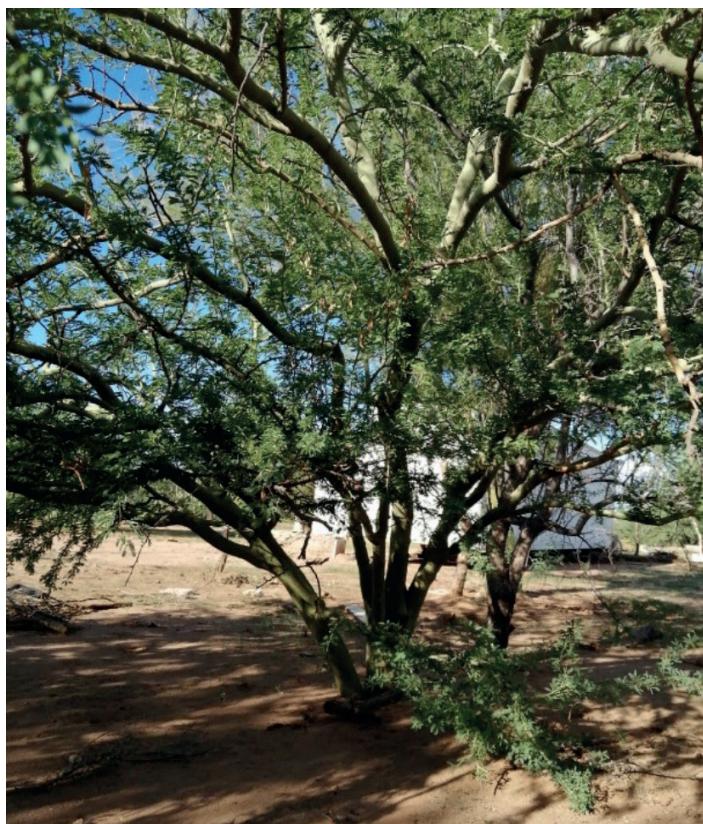


Figure 1: *Parkinsonia praecox* (Ruiz & Pav.) Hawkins.

tana (specialist on plant taxonomy) and deposited in the Herbarium of the Universidad Autónoma Agraria Antonio Narro (ANSM), Saltillo, Coahuila, Mexico. The samples were dried at room temperature and extracted with methanol (1:10 w/v) during four days with occasional stirring. Obtained extracts were filtered (Whatman No. 4) and the solvent was removed under vacuum (IKA, RV 10 digital, Wilmington, NC, USA) at 40 °C, followed by storage at -20 °C.

Phytochemical screenings

Phytochemical screenings of *P. praecox* extracts were performed to determine the presence of terpenes, flavonoids, tannins, saponins, cardiac glycosides, sugars, free amino-acids, alkaloids, phenolic compounds, coumarin and quinones following published methods.

Terpene identification

Identification of these compounds was carried out through the Lieberman-Burchard test (Burke et al., 1974; Xiong et al., 2007; Adu et al., 2019; Wutsqa et al., 2021). Briefly, 0.05

g of *P. praecox* extracts were dissolved in 1 ml of chloroform with a further addition of 1 ml of acetic anhydride. Subsequently, two drops of concentrated H₂SO₄ (97%, MerckMillipore, Darmstadt, Germany) were added to the sample. A green color change indicated the presence of terpenes (Akter et al., 2016; Suurbaa et al., 2017).

Flavonoids identification

Flavonoids identification was carried out through the Shinoda reaction (Cheng-Yong et al., 2016; Ribeiro et al., 2017; Musman et al., 2019; Kumkhale et al., 2020). An aliquot (0.05 g) of *P. praecox* extracts was dissolved in ethanol (absolute, MerckMillipore, Darmstadt, Germany). Two drops of concentrated HCl (37%, MerckMillipore, Darmstadt, Germany) were added followed by a chip of magnesium metal. Appearance of a magenta color indicated the presence of flavonoids (Akter et al., 2016; Suurbaa et al., 2017; Gollo et al., 2020).

Tannins identification

An amount (0.05 g) of *P. praecox* extracts was dissolved in 2 ml of Milli-Q-water and filtered (Whatman No. 4). A couple of drops of 10% ferric chloride (Iron (III), chloride 97%, Sigma Aldrich, St. Louis, USA) solution were added to the sample and the formation of a blue-black color indicated the presence of tannins (Akter et al., 2016; Suurbaa et al., 2017; Gollo et al., 2020).

Saponins identification

Saponins were identified using the Rosenthaler's reaction (Borkar et al., 2016; Le et al., 2018). Briefly, 0.02 g of *P. praecox* extracts were placed in a watch glass and two drops of Rosenthaler's reagent and two drops of concentrated H₂SO₄ (97%, MerckMillipore, Darmstadt, Germany) were added. Formation of red to violet color in the sample indicated the presence of saponins (Suurbaa et al., 2017).

Cardiac glycosides identification

Identification of cardiac glycosides was performed following the Keller-Killiani reaction (DagerAlbalawi, 2016; Oladele et al., 2021). Briefly, 0.05 g of *P. praecox* extracts were dissolved in a solution formed by 1 ml of glacial acetic acid (>99%, Sigma Aldrich, St. Louis, USA) and two drops of 2% ferric chloride (Iron (III) chloride 97%, MerckMillipore, Darmstadt,

Germany). Afterwards, 1 ml of H_2SO_4 was slowly added and the formation of a red ring indicated the presence of cardiac glycosides (Suurbaa et al., 2017).

Alkaloids identification

An aliquot (0.05 g) of *P. praecox* extracts with 10 ml of 1% HCl (37%, MerckMillipore, Darmstadt, Germany) was heated to boil and then filtered. Then, two drops of Wagner's reagent were added to 2 ml of filtrate (Whatman No. 4). A brown precipitate indicated the presence of alkaloids. Afterwards, two drops of acidified Mayer's reagent were treated with 2 ml of filtrate and yellowish cloudy precipitate indicated the presence of alkaloids (Akter et al., 2016; Suurbaa et al., 2017).

Phenolic compounds identification

Briefly, 0.05 g of *P. praecox* extracts were dissolved in 1 ml of Milli-Q-water, followed by two drops of 2% ferric chloride (Iron (III) chloride 97%, Sigma Aldrich, St. Louis, USA) solution. Positive results were associated with blue, green, red or purple colorations (Suurbaa et al., 2017).

Coumarin identification

The identification of coumarins was carried out with a reaction of potassium hydroxide. Briefly, *P. praecox* extracts (0.05 g) were dissolved in 1 ml of boiling Milli-Q-water. Later, a portion of this solution was taken with a capillary and placed on a paper filter piece and allowed to dry. Once the sample was dried on the filter paper, one drop of potassium hydroxide (99.95%, Sigma Aldrich, St. Louis, USA) solution (0.5 M) was added and observed under a UV lamp at 366 nm (CAMAG UV 4 lamp, Muttenz, Switzerland). Yellow fluorescence coloration was considered positive for coumarine (Suurbaa et al., 2017).

Quinones identification

One drop of concentrated sulfuric acid (97%, MerckMillipore, Darmstadt, Germany) was added to 0.02 g of *P. praecox* extracts. The formation of red color indicated the presence of anthraquinones (Suurbaa et al., 2017).

Sugars identification

Identification of glycosides in the extract was carried out using a reaction with sodium hydroxide. For that, 0.05 g of *P.*

praecox extracts were dissolved in 1 ml of Milli-Q-water and two drops of the sodium hydroxide (98%, Sigma Aldrich, St. Louis, USA) solution were also added. A yellow coloration indicated the presence of glycosides in the sample (Suurbaa et al., 2017).

Free aminoacids identification

Briefly, 0.05 g of *P. praecox* extracts were dissolved in 1 ml of ninhydrin reagent (Sigma Aldrich, St. Louis, USA), and the solution was boiled in a water bath for 10 min. Color-change expectations were according with the presence of certain type of amino group: bluish-violet color changes for those with primary amino group, yellow for proline and hydroxyproline, and brown for asparagine which has an amino group in the side chain (Suurbaa et al., 2017).

Folin Ciocalteu assay

Parkinsonia praecox extracts (10 μ l:1 mg/ml) were combined with distilled water (80 μ l), Folin-Ciocalteu reagent (2 N, Sigma-Aldrich, St. Louis, USA) 0.25 N (40 μ l), sodium carbonate (>99.5%, Sigma-Aldrich, St. Louis, USA) (60 μ l, 5% in distilled water), and distilled water (80 μ l). Samples were stored for 1 h (darkness) at 25 °C. Finally, absorbance was read on a microplate reader (750 nm) (ThermoScientific Multiskan GO, Osterode, Germany). Results were expressed as mg of gallic acid equivalent (GAE)/g dried weight (d.w.) (Velazquez et al., 2007).

UPLC-DAD analysis

Analysis of phenolic compounds of *P. praecox* extracts were determined by the UPLC™ H-class system (Acquity, Waters Co., Milford, USA) equipped with a photodiode array extended λ (PDA e λ) detector, an Acquity UPLC™ BEH C18 VanGuard precolumn (130 Å, 1.7 μ m, 2.1 mm × 5 mm), and an UPLC™ BEH C18 column (1.7 μ m, 3.0 × 100 mm) with a column temperature of 60 °C and an auto sampler set at 5 °C. The mobile phase was constituted by 0.5% formic acid in water (solvent A) (solution 50% in water, Sigma-Aldrich, St. Louis, USA) and methanol (solvent B) (HPLC grade, JT Baker Inc, Phillipsburg, USA). The elution was performed at different conditions: 0 min, flow 0.4 ml/min, 80% solvent A; 0.25 min, flow 0.150 ml/min, 80% solvent A; 5 min, flow 0.200 ml/min, 80% solvent A; 12 min, flow 0.180 ml/min,



55% solvent A; 25 min, flow 0.100 ml/min, 0% solvent A; 26 min, flow 0.200 ml/min, 60% solvent A; 27 min, flow 0.400 ml/min, 80% solvent A; 30 min, flow 0.400 ml/min, 80% solvent A. The detection of phenolic compounds was performed by the retention time of standards and their absorption spectrum (280 nm, 320 nm, 360 nm and 380 nm). Analyzed compounds were caffeic acid, cinnamic acid, chlorogenic acid, ellagic acid, ferulic acid, gallic acid, 2-hydroxy-cinnamic acid, *p*-coumaric acid, 3-hydroxybiphenyl, protocatechuic acid, rosmarinic acid, sinapic acid, syringic acid, vanillic acid, apigenin, catechol, catechin hydrate, epicatechin, epigallocatechin gallate, gallicatechin gallate, kaempferol, kaempferol 3- β -glucopyranose, luteolin, maclurin, mangiferin, myricetin, naringenin, naringin, neohesperidin, phloretin, phloridzin dyhydrate, pyrogallol, polydatin, quercetin-3- β -d-glucoside, quercetin dihydrate, resveratrol and rutin. Quantification of phenolic compounds was performed with standard calibration curves, and results were reported as μ g/g d.w. (Velderrain-Rodríguez et al., 2018). Identified compounds in the extracts were quercetin (retention time: 17.57-17.68 min; quantification range: 0.584 to 149.5 μ g/ml; regression type: lineal; r^2 value: 0.9997) (Sigma-Aldrich, St. Louis, USA) and *p*-coumaric acid (retention time: 10.13-10.42 min; quantification range: 0.756 to 96.75 μ g/ml; regression type: lineal; r^2 value: 0.9998) (Sigma-Aldrich, St. Louis, USA).

DPPH assay

Parkinsonia praecox extracts (100 μ l) were mixed with DPPH (Sigma-Aldrich, St. Louis, USA) solution at 300 μ M (100 μ l). After 30 min stored (in darkness) at 25 °C the absorbance was read on a microplate reader (517 nm) (ThermoScientific Multiskan GO, Osterode, Germany). Results were reported as IC_{50} (concentration to inhibit the 50% of DPPH radical, μ g/ml) and μ M Trolox Equivalent (TE)/g d.w. (Usia et al., 2002)

ABTS assay

ABTS + assay was performed by mixing 5 ml of ABTS (Sigma-Aldrich, St. Louis, USA) (7 mM) with 88 μ l of $K_2S_2O_8$ (>99%, Sigma-Aldrich, St. Louis, USA) (0.139 mM) for 16 h (in darkness). The absorbance of the radical was adjusted to 0.700±0.02 at 754 nm. *Parkinsonia praecox* extracts (5 μ l) were combined with adjusted radical (245 μ l) and then the

samples were incubated (5 min) in darkness. Finally, the absorbance was read at 754 nm (Fluostar Omega microplate reader, BMG Labtech, Ortenberg, Germany). Results were reported as μ M Trolox Equivalent (TE)/g d.w (Modified method of Re et al., 1999).

ORAC assay

One hundred and fifty μ l of fluorescein (Free acid, Sigma-Aldrich, St. Louis, USA) (10 nM), 25 μ l of phosphate buffer (Sodium phosphate dibasic heptahydrate and hydrated monobasic sodium phosphate; Fagalab, Mocorito, Sinaloa, Mexico) (75 mM, pH 7.4) as blank, and 25 μ l (50 μ g/ml) of *P. praecox* extracts were mixed. AAPH (97%, Sigma-Aldrich, St. Louis, USA) (240 mM) was also incorporated to the mixture and incubated at 37 °C (15 min). Fluorescence was read every 90 s for 1.5 h at an excitation wavelength of 485 nm and an emission wavelength of 530 nm on a microplate reader (Fluostar Omega microplate reader, BMG Labtech, Ortenberg, Germany). Results were reported as μ M Trolox Equivalent (TE)/g dried weight (d.w.) (modified method of Ou et al., 2001).

FRAP assay

FRAP reagent was elaborated with the combination of ten volumes of acetate buffer (300 mM, pH 3.6), 1 volume of TPTZ (>98%, Sigma-Aldrich, St. Louis, USA) (40 mM, dissolved in 40 mM HCl), and 1 volume of ferric chloride (97%, Sigma-Aldrich, St. Louis, USA) (20 mM, dissolved in water). *Parkinsonia praecox* extracts (20 μ l) were combined with FRAP reagent (280 μ l). After 30 min (in darkness) at 25 °C the samples were read using a microplate reader (630 nm) (ThermoScientific Multiskan GO, Osterode, Germany). Results were reported as μ M of Fe(II)/g d.w. (Benzie and Strain, 1996).

Cell culture

Cell lines were cultured in DMEM (Dulbecco's Modified Eagle Medium) (Sigma-Aldrich, St. Louis, USA) supplemented with 5% heat-inactivated serum feral bovine (ByProducts, Guadalajara, Mexico) and penicillin (Sigma-Aldrich, St. Louis, USA) (100 U/ml) in 25 cm² culture dishes. The cultures were maintained in an Isotherm incubator (Fisher Scientific, Waltham, USA) under the standard conditions (5% of CO₂ at 37 °C



and 95% of relative humidity). A549 (non-small-cell lung cancer cells), MDA-MB-231 (triple negative breast cancer), PC-3 (adenocarcinoma prostate cancer grade IV) and HeLa (human cervical cancer) and L929 (non-cancerous subcutaneous connective tissue) were obtained from the American Type Culture Collection (ATCC; Rockville, USA).

MTT assay

Fifty μ l (1×10^4 cells) were placed in each well of a flat 96-well plate and incubated (24 h, 37 °C, 5% of CO₂ atmosphere). Afterwards, the cells were treated with *P. praecox* extracts (50 μ l) and incubated during 48 h under the standard conditions. Previously, extracts were dissolved in DMSO and re-suspended in DMEM (Sigma-Aldrich, St. Louis, USA) to give a final DMSO (>99.9%, Sigma-Aldrich, St. Louis, USA) concentration of 0.25% (that showed no interference with the cell lines' growth). Then, DMEM (50 μ l) containing different concentrations of the extracts were added and the cell cultures were incubated for 48 h. Subsequently, MTT (Sigma-Aldrich, St. Louis, USA) solution (10 μ l, 5 mg/ml) was added to each well. Next, formazan crystals generated by metabolically active cells, were dissolved with acid isopropanol. The absorbance of microplates was read at 570 and 630 nm (iMark microplate absorbance reader, BioRad, Laboratories, Cd. Mx, Mexico). Results were reported as IC₅₀ values (IC₅₀ is defined as the required concentration to inhibit 50% of cell proliferation), and were calculated using nonlinear regression analysis in Microsoft Excel software (Hernandez et al., 2007; Torres-Moreno et al., 2015).

Statistical analysis

One-way analysis of variance was performed for the obtained data using the NCSS Statistical software v. 2007 (NCSS Statistical software, 2021). Tukey-Kramer tests were used to mean comparisons. Significance level was $p \leq 0.05$. Correlation analysis was performed using Pearson correlation.

Results

Phytochemical screenings

Phytochemicals screening of *P. praecox* extracts is shown in Table 1. The analysis consists of the determination of 11 groups of secondary metabolites, where PPB presented the highest quantity of metabolites, among them terpenes, phe-

Table 1: Phytochemical screening of *Parkinsonia praecox* (Ruiz & Pav.) Hawkins extracts. PPF: *Parkinsonia praecox* flower extract; PPB: *Parkinsonia praecox* berry extract; PPS: *Parkinsonia praecox* stem extract. - (absence); + (presence).

Secondary metabolite	PPF	PPB	PPS
Terpenes	-	+	-
Flavonoids	-	+	+
Tannins	+	+	+
Saponins	-	-	-
Cardiac glycosides	-	-	-
Sugars	+	+	+
Free amino acids	-	-	-
Alkaloids	-	-	-
Phenolic compounds	+	+	+
Coumarin	-	-	-
Quinones	-	-	-

nolic compounds, flavonoids, tannins and sugars. PPF and PPS exhibited similar bioactive compounds profile comparing with PPB, except for terpenes. It is important to highlight that these qualitative tests showed a pronounced intensity in the reaction corresponding to phenolic compounds for all cases, probably being the main group of compounds in the extracts.

Quantification of phenolic compounds

The phytochemical screening suggests that phenolic compounds are the most predominant bioactive compounds present in *P. praecox* extracts; phenolic compounds were quantified by the Folin-Ciocalteu method. Results evidenced that PPS presented higher amount of phenolic content ($p < 0.05$) compared to PPF and PPB, which presented 2.9 and 1.7 lower concentration, respectively (Fig. 2).

UPLC-DAD analysis

To identify the phenolic compounds presented in *P. praecox* extracts, UPLC-DAD analysis was carried out. For chromatogram interpretation, a contrasting analysis using a database of 37 phenolic compounds (phenolic acids and flavonoids) was performed. The analysis of PPS allowed to identify the presence of quercetin with a concentration of 218.8 μ g/g (Table 2), while in PPF, *p*-coumaric acid at concentration of 176.4 μ g/g was identified (Table 2). Regarding PPB, it was not possible to identify any flavonoid or phenolic acid. This



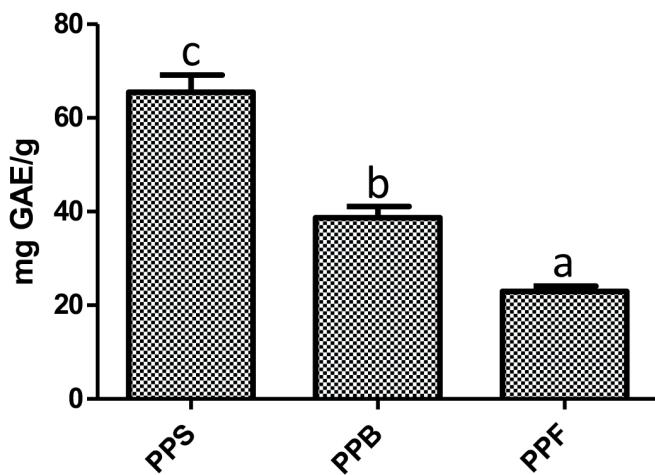


Figure 2: Phenolic quantification of *Parkinsonia praecox* (Ruiz & Pav.) Hawkins extract. PPS: *Parkinsonia praecox* stem extract; PPB: *Parkinsonia praecox* berry extract; PPF: *Parkinsonia praecox* flower extract. Data are presented as the mean \pm standard deviation of three independent experiments. ^{a-c}Bars with different letter in each graph represent significantly different values ($p<0.05$).

suggests that different phenolic compounds are present in PPB, which were not identified due to the limitations of the database.

Antioxidant activity

Antioxidant potential of evaluated extracts of *P. praecox* is shown in **Table 3**. Results obtained by the DPPH method evidenced that *P. praecox* extracts exhibited the capability to scavenge this free radical. PPS presented the highest ($p<0.05$) antioxidant potential comparing with other evaluated extracts. Additionally, PPS required the lowest ($p<0.05$) concentration (137 μ g/ml) to stabilize the 50% of DPPH radical (IC_{50}), while in case of PPB and PPF an increment of the concentration was required (1.4 and 2.2 time, respectively) to show a similar effect.

On the other hand, *P. praecox* extracts were able to stabilize the ABTS radical (**Table 3**). The PPS extract was the one with the highest ABTS radical stabilizing activity ($p<0.05$) compared to the PPB and PPF extracts, which exhibited 1.50 and 1.59 lower stabilizing capability, respectively.

Interestingly, *P. praecox* extracts, in addition to stabilizing synthetic radicals (DPPH and ABTS), were also able

Table 2: Concentration (μ g/g) of identified phenolic compounds in *Parkinsonia praecox* (Ruiz & Pav.) Hawkins extracts. PPS: *Parkinsonia praecox* stem extract; PPB: *Parkinsonia praecox* berry extract; PPF: *Parkinsonia praecox* flower extract. Data are presented as the mean \pm standard deviation of two independent experiments. ND: no detected.

Compound	PPS	PPB	PPF
Quercetin	218.8 \pm 1.5	ND	ND
<i>p</i> -coumaric acid	ND	ND	176.4 \pm 2.6

to inhibit biological radicals such as hydroxyl radical, which was evaluated through the ORAC method (**Table 3**). The results showed that PPS presented the highest ORAC values ($p<0.05$), which indicates greater effectiveness in the inhibition of the hydroxyl radical. The PPB and PPF extracts also showed antioxidant capacity by this method; however, it was 48.2 and 57.8% less effective than PPS.

Reducing antioxidant potential of *P. praecox* extracts was also demonstrated (**Table 3**). The evaluated extracts showed capability to reduce the ferric-TPTZ complex to ferrous-TPTZ complex. However, the PPS extract exhibited the highest ($p<0.05$) reducing power, presenting 200.04% and 304.9% greater antioxidant potential than PPB and PPF, respectively.

Correlation analysis was carried out between the antioxidant activity (DPPH, ABTS, ORAC and FRAP) and the content of phenolic compounds in *P. praecox* extracts (**Table 4**). Obtained results demonstrated that there is a strong correlation between the antioxidant activity of the *P. praecox* extracts and the content of total phenols, since the *r* values higher than 0.96 were obtained in all cases.

Antiproliferative activity

In this study, we evaluated by MTT assay the antiproliferative effect of *P. praecox* extracts against a panel of cancer cell lines and against the non-cancerous L929 cell line (**Table 5**). In general, prostate cancer PC-3 cells were the most sensitive for all the treatments; however, the highest activity was observed for PPF followed by PPS and PPB. L929 cells were most resistant to PPF and PPB, which suggests that these extracts are more selective than PPS. On the other hand, PPS showed moderate activity against MDA-MB-231 and HeLa cells; nevertheless, the proliferation of L929 cells was inhibited most efficiently by this extract.

Table 3: Antioxidant activity of *Parkinsonia praecox* (Ruiz & Pav.) Hawkins extracts. PPS: *Parkinsonia praecox* stem extract; PPB: *Parkinsonia praecox* berry extract; PPF: *Parkinsonia praecox* flower extract; DPPH: 2,2-diphenyl-1-picrylhydrazyl; TEAC: trolox equivalent antioxidant capacity; FRAP: ferric reducing antioxidant power assay. Data are presented as the mean \pm standard deviation of three independent experiments. ^{a-c}Different letters within the same column indicate significant differences ($p<0.05$).

Treatment	Antioxidant assay				
	DPPH		TEAC	ORAC	FRAP
	$\mu\text{M ET/g}$	$\text{IC}_{50} (\mu\text{g/ml})$	$\mu\text{M ET/g}$	$\mu\text{M ET/g}$	$\mu\text{M Fe(II)/g}$
PPS	262.3 \pm 30.7 ^c	137 \pm 12.5a	39.5 \pm 0.2 ^c	1777.7 \pm 45.5 ^c	935.6 \pm 52.7 ^c
PPB	179.8 \pm 2.8 ^b	190.9 \pm 6.6 ^b	26.3 \pm 0.1 ^b	920.6 \pm 33.9 ^b	467.7 \pm 17.8 ^b
PPF	112.7 \pm 3.8 ^a	302.3 \pm 7.3 ^c	24.7 \pm 0.1 ^a	750.3 \pm 41.3 ^a	306.9 \pm 14.8 ^a

Table 4: Correlation (r) between total phenolic content (TPC) and antioxidant activity of *Parkinsonia praecox* (Ruiz & Pav.) Hawkins. DPPH: 2,2-diphenyl-1-picrylhydrazyl; ABTS: 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid; ORAC: assay measures antioxidant capacity; FRAP: ferric reducing antioxidant power assay.

Antioxidant method				
DPPH	ABTS	ORAC	FRAP	TPC
0.96	0.96	0.97	0.99	

Discussion

Natural products have been used for the treatment of different illnesses worldwide. This trend is still moving on and currently, 80% of the human population uses them as primary treatment for different health conditions (Sharifi-Rad et al., 2020). Effectiveness of some of these products has been demonstrated for the development of drugs, due to the presence of highly effective chemical compounds (Newman and Cragg, 2020; Zhang et al., 2020).

A previous study demonstrated the presence of several chemical compounds in *P. praecox*, such as alkaloids, triterpenes, sterols, saponins, flavonoids and polyphenolics (Torrealba, 2008). Additionally, other *Parkinsonia* species such as *P. aculeata* presented a wide variety of bioactive compounds, such as terpenes, saponins, steroids, phenolic compounds, flavonoids, tannins, alkaloids, proteins, amino acids, glucosides, and carbohydrates (Leite et al., 2010; Saha et al., 2010b; Sharma and Vig, 2014; Qureshi, 2017). Our study demonstrated that *P. praecox* presented minor groups of secondary metabolites, which could be associated

Table 5: Antiproliferative activity of *Parkinsonia praecox* (Ruiz & Pav.) Hawkins. PPS: *Parkinsonia praecox* stem extract; PPB: *Parkinsonia praecox* berry extract; PPF: *Parkinsonia praecox* flower extract. Data are presented as the mean \pm standard deviation of three independent experiments. ^{a-b}Different letters within the same column indicate significant differences ($p<0.05$). IC_{50} not determined at the maximum tested concentration (400 $\mu\text{g/ml}$).

Treatments	Cell lines $\text{IC}_{50} (\mu\text{g/ml})$				
	A549	MDA-MB-231	PC-3	HeLa	L929
PPS	341.3	147.3	78.87	121.65	93.29
	\pm	\pm	\pm	\pm	\pm
	13.6	8.9	0.5a	7.7a	5.9
PPB			94.21	220.20	
	>400		>400	\pm	\pm
				0.3b	13.2b
PPF			68.38	222.45	
	>400		>400	\pm	\pm
				9.1a	26.6b

with the analyzed plant genus, metabolism, biotic and abiotic plant conditions, type of extraction method and solvents used. These factors determine the bioactive compounds profile that will be present in natural extracts (Azmir et al., 2013; Ochoa-Velasco et al., 2017).

Phytochemical analysis showed that phenolic compounds are the most predominant bioactive compounds present in *P. praecox*. The obtained results in this study are consistent with previously reported for other analyzed species of the genus, such as *P. aculeata*, which presented phenolic content values between 10 to 288 mg GAE/g (Aranda-Souza et al., 2012; Sharma and Vig, 2013, 2014; Mud-

dathir et al., 2017; Qasim et al., 2017). In addition, this study shows for the first time the presence of specific compounds in *P. praecox*, which could support the biological potential of this plant.

The present study provides novel information about the antioxidant potential of *P. praecox*, being the first report about its biological activity. In addition, it is important to highlight that PPS could be classified as an extract with medium antioxidant potential because it presented an IC_{50} value lower than 150 μ g/ml, while the other extracts exhibited low antioxidant potential (Fidrianny et al., 2015). Additionally, PPS is a natural extract with high reducing potential because it presented FRAP value >500 μ M Fe(II)/g, which is the limit value to consider it a natural source with this characteristic (Wong et al., 2006).

On the other hand, the obtained antioxidant results in this study agree with the previously reported ones for *P. aculeata*. Aranda-Souza et al. (2012), Sharma and Vig (2013; 2014), and Sharma et al. (2017) reported that *P. aculeata* extracts obtained with different solvents (methanol, butanol, acetone, chloroform and hexane) exhibited capability to stabilize the DPPH radical (IC_{50} : 10.8 to <1000 μ g/ml), hydroxyl radical (200 μ g/ml of extracts inhibited the hydroxyl radical formation between 20.3 to 80%) and reduce metals (461 to <498 μ M Fe(II)/g; IC_{50} : 162.6 to <200.7 μ g/ml).

Elevated rates of free radicals and transition metals in biological systems cause indiscriminate damage of biomolecules such as DNA, leading to the development of several pathologies like neurodegenerative diseases, atherosclerosis, cancer and others (Collin, 2019). In this regard, *P. praecox* (especially PPS) showed the capability to stabilize synthetic and biological radicals, as well as to reduce metals. These extracts can represent a natural alternative that could be used to inhibit the formation of free radicals and interrupt their propagation.

Antioxidant activity of natural products is usually related to the presence of chemical compounds in plants, specifically with phenolic compounds (Vidal Gutiérrez et al., 2020). This suggests that the antioxidant activity of *P. praecox* is associated with the presence of these types of compounds. The highest antioxidant activity of PPS can be associated with the presence of phenolic compounds, mainly

quercetin. Different studies have demonstrated that this flavonoid is one of the most potent antioxidant agents in nature (Sharma et al., 2018). The antioxidant mechanism of quercetin is related with the presence of hydroxyl groups in the catechol ring (substitution in position 3' and 4) and chromane ring (substitution in position 3). In addition, the C ring plays an important role, especially the double bond between C2-C3. This characteristic promotes the electron or proton donation to exert antioxidant effect, and then confers stability by resonance effect (Ulusoy and Sanlier, 2020). Likewise, the antioxidant effect of PPF could be associated with the presence of *p*-coumaric acid, whose mechanism of action is due to the presence of the hydroxyl group in C3 of the phenolic ring and to the length of its hydrocarbon chain (Jing et al., 2012). These characteristics allow to donate protons or electrons to stabilize free radicals and reduce metals, as well as to stabilize the produced phenolic radicals (Jing et al., 2012). In the case of PPB, its antioxidant effect can be related with the presence of phenolic compounds which were not identified in this study. In addition to phenolic compounds, terpenes are also known for their antioxidant activity. Therefore, the antioxidant activity of PPB can be related with the presence of these types of compounds, whose mechanism of action is associated with the presence of hydroxyl or methoxyl groups in the phenolic ring (Gonzalez-Burgos and Gómez-Serranillos, 2012).

On the other hand, nowadays the search of new selective and effective treatments against cancer continues to be a priority for public health worldwide (Zhong et al., 2021). In this context, medicinal plants provide a good option for the search of new anticancer drugs (Block et al., 2015). According to the National Cancer Institute (NCI, USA) guidelines (Grever et al., 1992), a crude extract could be considered cytotoxic when it inhibits the proliferation of a cell line with a IC_{50} value <30 μ g/ml (Grever et al., 1992). However, even though *P. praecox* extracts inhibited cell proliferation with IC_{50} values >30 μ g/ml, the study of the antiproliferative effect of its fractions or isolated compounds cannot be ruled out. Previous research about plants has shown the possibility of compound isolation with great antiproliferative effect from methanolic extracts that inhibit a particular cancer cell line proliferation with IC_{50} values around 130 μ g/ml (Tor-

res-Moreno et al., 2015). Hence, *P. praecox* extracts could be considered to obtain antiproliferative compounds through a bioguided separation.

In several studies, it has been shown that there is a direct correlation between the phenolic composition and the antiproliferative activity of plants (Maya-Cano et al., 2021). The phytochemical screening showed that *P. praecox* extracts are constituted by phenolic compounds such as quercetin and *p*-coumaric acid that have been identified in PPS and PPF, respectively. Antiproliferative, apoptotic and cell cycle arrest capability of quercetin and *p*-coumaric acid on cancer cells have been demonstrated (Wang et al., 2012; Abotaleb et al., 2020). Wang et al. (2012) reported that quercetin reduces the proliferation of the prostate cancer cell lines PC-3 ($\approx 10\%$) and LNCaP (in ≈ 20 and 30%) at 5 and 10 μM and induces apoptosis and cell cycle arrest in S and G2 M phases. *P*-coumaric acid reduces the proliferation of the ovarian cancer cells A-2780 with IC_{50} of 3.64×10^{-2} mg/ml (Shahwar et al., 2015). The proliferation of the colon cancer cells HT-29 was reduced by *p*-coumaric acid with IC_{50} values of 1600 $\mu\text{mol/l}$ and 1000 $\mu\text{mol/l}$ at 48 and 72 h, respectively, whereas the proliferation of SW480 colon cancer cells was inhibited with IC_{50} of 1400 $\mu\text{mol/l}$ and 1000 $\mu\text{mol/l}$ at 48 and 72 h. The inhibition of HT-29 and SW480 cell proliferation was associated with the capability of *p*-coumaric acid to induce apoptosis through the inhibition of Gr78 via activation of PERK-eIF2 α -ATF-4-CHOP pathway (Abotaleb et al., 2020).

Hence, these compounds, synergistically or additively with other unidentified metabolites, could be contributing to the antiproliferative effect of PPF and PPS. On the other hand, terpenes are a large class of natural compounds with anticancer effect (Huang et al., 2012). Phytochemical analysis showed that terpenes are only present in PPB (Table 1); this suggests that the antiproliferative effect of PPB is due to the combined action of terpenes and phenolic compounds (Wright, 2001; Gul et al., 2016).

Conclusions

The present study demonstrated that *P. praecox* could be a source of several chemical compounds with biological potential. Additionally, a possible relation between antioxidant and antiproliferative activity is suggested as a conse-

quence of the presence of phenolic compounds. Therefore, despite showing moderate potential, this plant could be used to obtain fractions or to purify compounds with this capability, representing an alternative for different pharmaceutical applications views.

Author contributions

JCLR and HTM conceived and designed the study, wrote, reviewed, and approved the final manuscript. KLRM and RERZ performed experiments, analyzed and interpreted the results, and revised the manuscript. DEVR, MALP, and RELS performed experiments, analyzed results, and revised the manuscript. MAVO, NJS, GGA, performed experiments, analyzed results, and revised the manuscript.

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