

Oogenic development and gonotrophic cycle of *Aedes aegypti* and *Aedes albopictus* in laboratory

Mauricio Casas-Martínez, DSc,⁽¹⁾ Rodrigo Tamayo-Domínguez, BSc,⁽¹⁾ J Guillermo Bond-Compeán, DSc,⁽¹⁾ Julio C Rojas, DPhil,⁽²⁾ Manuel Weber, PhD,⁽³⁾ Armando Ulloa-García, DSc.⁽⁴⁾

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Desarrollo oogénico y ciclo gonotrófico de *Aedes aegypti* y *Aedes albopictus* en laboratorio.

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Abstract

Objective. To determine the time of oogenic development and the length of the gonotrophic cycle of *Ae. aegypti* and *Ae. albopictus* in laboratory. **Materials and methods.** Bloodfed females of *Ae. aegypti* and *Ae. albopictus* were dissected every 4 h to determine the development status of the follicles according to the Christophers' stages. **Results.** The minimum time of oocyte maturation in *Ae. aegypti* and *Ae. albopictus* was 64-82 h and 52-64 h post-feeding, respectively. We found that the gonotrophic cycle of *Ae. aegypti* (3.7-4.2 d) is longer than that of *Ae. albopictus* (3.2-3.7 d). The follicle length showed significant differences between species at Christophers' stages 2^o and 5, whereas follicle amplitude was different between the two mosquitoes at stages 2^o, 3 and 4. **Conclusions.** The study provided new evidence on the reproductive strategies of *Ae. aegypti* and *Ae. albopictus* females that coexist in the Neotropical region of Mexico.

Keywords: oogenic development; gonotrophic cycle; *Aedes*; vectors; dengue

Resumen

Objetivo. Determinar el tiempo de desarrollo oogénico y del ciclo gonotrófico de *Aedes aegypti* y *Aedes albopictus* en laboratorio. **Material y métodos.** Hembras de *Ae. aegypti* y *Ae. albopictus* alimentadas con sangre fueron disecadas cada cuatro horas para determinar el estado de desarrollo folicular, según los estadios de Christophers. **Resultados.** El tiempo mínimo de maduración del oocito en *Ae. aegypti* y *Ae. albopictus* fue de 64-82 h y 52-64 h post-alimentación, respectivamente. El ciclo gonotrófico de *Ae. aegypti* (3.7-4.2 d) fue mayor que el de *Ae. albopictus* (3.2-3.7 d). La longitud folicular presentó diferencias significativas entre las especies en los estadios de Christophers 2^o y 5, mientras que la amplitud folicular fue diferente entre ambos mosquitos en los estadios 2^o, 3 y 4. **Conclusiones.** El estudio proporcionó nueva evidencia sobre la estrategia reproductiva de las hembras de *Ae. aegypti* y *Ae. albopictus* que coexisten en la región neotropical de México.

Palabras clave: desarrollo oogénico; ciclo gonotrófico; *Aedes*; vectores; dengue

(1) Centro Regional de Investigación en Salud Pública, Instituto Nacional de Salud Pública. Tapachula, Chiapas, Mexico.

(2) Grupo de Ecología y Manejo de Artrópodos, El Colegio de la Frontera Sur Unidad Tapachula. Tapachula, Chiapas, Mexico.

(3) Departamento de Conservación de la Biodiversidad. El Colegio de la Frontera Sur Unidad Campeche. Campeche, Mexico.

(4) Facultad de Ciencias Químicas, Universidad Autónoma de Chiapas. Tapachula, Chiapas, Mexico.

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Corresponding author: Dr. Armando Ulloa García. Facultad de Ciencias Químicas, Universidad Autónoma de Chiapas. Carretera a Puerto Madero Km. 1.5. 30700, Tapachula, Chiapas, Mexico.
email: armando_ulloa@hotmail.com

The dengue fever is a disease caused by any one of the four related viruses distributed in more than 100 countries. Recently, the emergence and reemergence of other arboviruses such as Chikungunya and Zika have worsened the global epidemiological situation, mainly in the tropical and subtropical regions of the planet.^{1,2} These diseases are transmitted by mosquitoes, mainly by two species of the *Stegomyia* group: *Ae. aegypti* (Linnaeus, 1762) and *Ae. albopictus* (Skuse, 1894).^{3,4} Several studies have shown some differences between ecological aspects of *Ae. aegypti* and *Ae. albopictus*. For example, the former predominates in highly urbanized and suburban areas, whereas the latter is found in periurban and rural areas.^{5,6} Additionally, the former species depends mainly on human blood and tends to bite and rest indoors,⁷ whereas the latter feeds on a variety of vertebrate hosts outdoors.⁸ These ecological differences have been observed as well in local populations of *Ae. aegypti* and *Ae. albopictus* in villages located along the coastal plain of Chiapas, where the presence of both species was reported since 2002.^{9,10} From the perspective of human health, the gonotrophic cycle is one of the most important physiological processes in the life of the mosquito and an essential epidemiological component of the vectorial capacity model. This biological aspect is significant and decisive for the population dynamics of *Ae. aegypti* and *Ae. albopictus* in urban, suburban and rural ecological scenarios with outbreaks of endemic dengue and other arboviral diseases. Although there is information regarding the duration of the reproductive cycle of both species in several states of Mexico, few studies have generated information on the reproduction of both species occurring in the same geographical region and sharing identical climatic conditions and resources. Therefore, it is necessary to investigate whether these ecological differences induce changes in the length of gonotrophic cycle in both species. Several laboratory and field studies on their reproductive capacity have been reported separately for each one.^{11,12} The objective of this study was to determine the length of the gonotrophic cycles of two day-biting mosquitoes, *Ae. aegypti* and *Ae. albopictus*. An understanding of reproductive aspects of the natural populations of both species is valuable for assessing and predicting their roles in the ecological dynamics and transmission of arboviruses.¹³

Materials and methods

Collection of mosquitoes and rearing process. Females of *Ae. aegypti* and *Ae. albopictus* were collected with entomological nets and mouth aspirators in houses as well as in cemeteries of Tapachula (14° 54' 29" N, 92° 15' 38" W and 177 masl), Chiapas, Mexico. The collections were

performed from July to October 2010, during the rainy season. All collections were performed between 08:00 and 12:00 h. The captured mosquitoes were transported to the laboratory, where they were identified at species level based on their morphological characteristics.¹⁴ The mosquitoes were separated by species and kept in 45 cm³ metal cages covered with mesh gauze under laboratory conditions of 26.0±2.0°C, 75.0±5.0% RH, and 12:12 light/dark photoperiod. The field-collected females were fed with rabbit blood and submitted to a synchronized oviposition every three and four days throughout 10 gonotrophic cycles in order to obtain enough eggs of the same generation for the experiments. All *Ae. aegypti* and *Ae. albopictus* eggs were maintained on moist filter paper during three days to ensure the development of the embryos.¹⁵ The eggs were placed in white plastic containers (30 x 40 x 10 cm) with 2.5 l of filtered water at 30°C to induce egg hatching. Immature stages were fed with a powdered rodent diet (raw protein 23%, raw fat 4.5%, raw fiber 6%, ash 8%, and 12% humidity). The pupae were placed in round plastic containers (5.5 cm high x 10.5 cm diameter) with 300 ml of water; the containers were introduced into 45 cm³ metal cages until the adult emerged. All newly emerged mosquitoes were fed with a 10% sucrose solution during three days before the experiments under constant conditions of temperature and RH.

Vitellogenesis. Christophers' stages and duration of the oogenic development were determined for *Ae. aegypti* and *Ae. albopictus* based on the appearance of the follicles.¹⁶ Thirty unfed females (without traces of blood) of each species were dissected to determine the maturation status of their eggs. The remaining females were allowed to feed on a rabbit. Mosquitoes were supplied with cotton pads soaked with a 10% sucrose solution and maintained in the insectary at 26.5±1.1°C, 81.3±7.0% RH, and with a 12:12 h photoperiod. Starting at 4 h after bloodfeeding and continuing every 4 h up to 75 h, groups of 30 mosquitoes were dissected to determine their Christophers' stages. Females that did not develop beyond Christophers' stages 2 or 2' 18 h after feeding were reported as pre-gravid.¹⁷

Morphometry of the follicles of Ae. aegypti and Ae. albopictus. In order to document the maturation process of eggs, we photographed 10 follicles per female to compare the appearance of the yolk sac, oocyte and nurse cells, and the follicular size through time, according the scheme developed by Christophers.¹⁸ The size of the follicles was measured using an ocular micrometer (model Tokyo P7X) mounted in a stereo microscope (model SMZ645). The follicles were observed at 10 and 40 X. The length

Ae. aegypti and 76-88 h (3.2-3.7 d) for *Ae. albopictus*. Ninety percent (135/150) of those *Aedes aegypti* females that were maintained up to 80 h to complete vitellogenesis developed to Christophers' 5, whereas only 84.1% (101/120) of the *Ae. albopictus* females completed vitellogenesis. We observed that 23.3% (35/150) of the *Ae. aegypti* females did not develop beyond Christophers' stage 2 or 2'', i.e., remained pre-gravid, compared to a mere 3.3% (1/30) of *Ae. albopictus* females (table I).

Morphometry of the follicles of Ae. aegypti and Ae. albopictus. We observed that the follicles had a semispherical shape throughout the development of the oocyte of both species (figure 1).

Their length and width increased gradually between Christophers' stages 1 and 3. From Christophers' stage 4, the follicles of both species grew in length until they reached Christophers' stage 4-5 (figure 2A and 2B).

Discussion

Aedes aegypti and *Ae. albopictus*, two important vectors of several arboviruses that affect humans worldwide, often interact in their invasive ranges. In these circumstances, a number of factors are thought to influence their population dynamics. The distribution and abundance of both species, which have profound epidemiological implications, are often governed by competitive interactions between sympatric mosquito populations.²⁰

The gonotrophic cycle stands out among the most widely researched reproductive aspects of the physiology of vectors. A number of authors argue that the

period from blood ingestion to egg deposition is one of the most important factors in the population dynamics of *Ae. aegypti* and *Ae. albopictus*. The duration of the gonotrophic cycle varies between mosquito species because multiple factors are involved.^{21,22}

The results reported here correspond to the time of the first gonotrophic cycle of F₁ females of each species, under laboratory conditions. *Aedes aegypti* showed a gonotrophic cycle length of 3.66 d, similar to that reported by MacDonald,²⁰ who performed laboratory experiments with mosquitoes from the Malayan Peninsula (3 d), while *Ae. albopictus* exhibited a gonotrophic cycle of 3.16 d. The same trends were also observed with Gubler's and Bhattacharya's method.²¹ We may argue that the observed differences could be due to characteristics intrinsic of each species, such as the temporal genetic divergence of the individuals, the nutritional reserves stored during the larval stages, and the hormonal modulation of the physiological processes,²²⁻²⁶ rather than the effect of the experimental factors, because our study was performed under controlled conditions of temperature and relative humidity. However, it is known that climatic factors can affect the duration of the gonotrophic cycle of both mosquitoes. For instance, a laboratory experiment showed that wide fluctuations in the daytime and nighttime environmental temperatures ($\pm 7^\circ\text{C}$) had an important effect on the gonotrophic cycle of *Ae. albopictus* that resulted in an average duration of more than a week, due to changes in the rates of blood digestion and vitellogenesis. Recently, a field study reported that the duration of the gonotrophic cycle of *Ae. aegypti* was affected by the climatic seasonality.

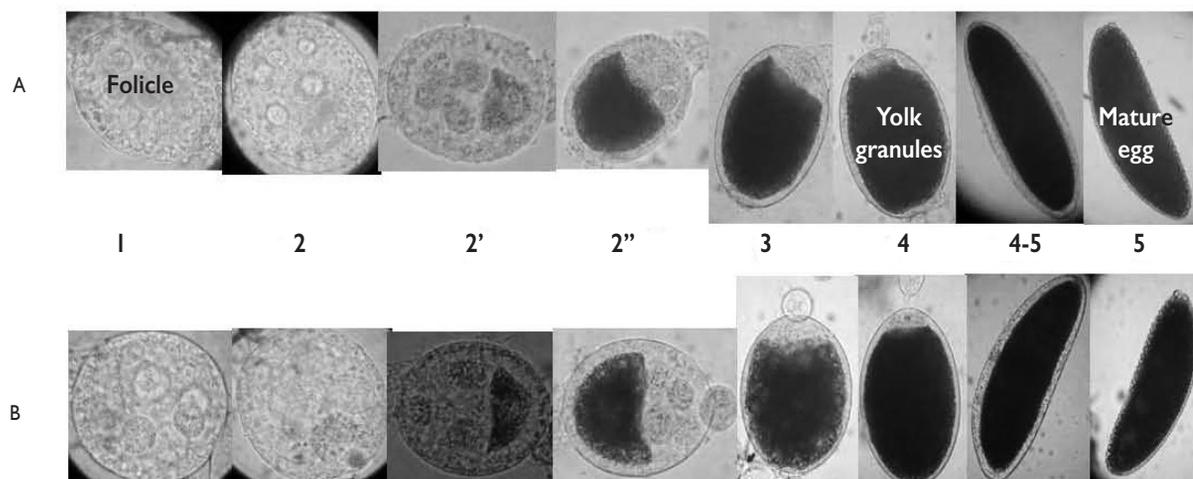


FIGURE 1. STRUCTURAL COMPARISON OF THE FOLLICLES OF *AE. AEGYPTI*: (A) AND *AE. ALBOPICTUS* (B) IN DIFFERENT CHRISTOPHERS' STAGES. TAPACHULA, CHIAPAS, MÉXICO, 2010

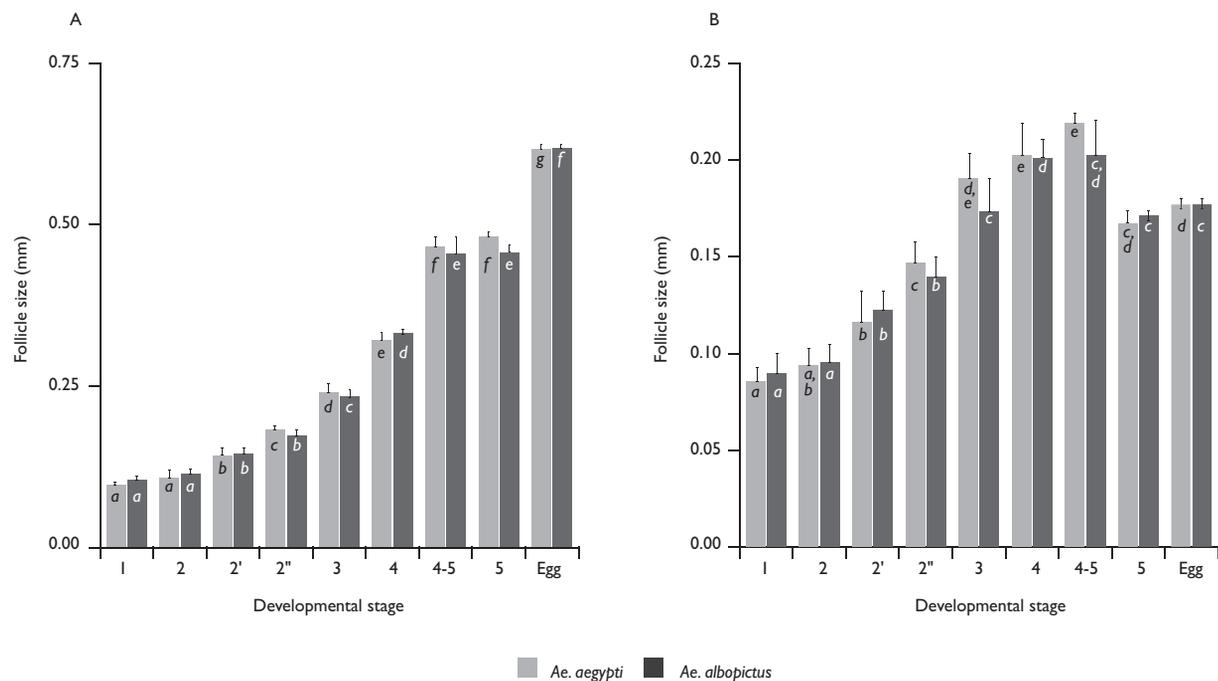


FIGURE 2. SIZE OF *AE. AEGYPTI* AND *AE. ALBOPICTUS* FOLLICLES AND EGGS: A) LENGTH (MEAN \pm SD) AND B) WIDTH (MEAN \pm SD). DIFFERENT LETTERS IN THE SAME COLOR BARS INDICATE SIGNIFICANT DIFFERENCES ($P \leq 0.05$). TAPACHULA, CHIAPAS, MÉXICO, 2010

Thus, in the rainy season at 26.7 °C it lasted four days, while in the dry season at 29.8 °C it lasted three days.²⁷ Consequently, it should be considered that the time of the oogenic development and, therefore, the duration of the gonotrophic cycle of both species in the field may vary due to the heterogeneity in the age structure of the female population, the influence of variations in the ambient temperature and relative humidity during the day and night,²⁸⁻³⁰ as well as to the source of blood supply and the presence of bacterial symbionts in the midgut of mosquitoes.³¹

In contrast to other studies,²⁷ the design of our experiment was more robust because we included a greater number of females dissected per time interval and three repetitions instead of one, and used F1 females of wild mosquitoes. In general, other studies have utilized females that have been kept in the laboratory for more than two years.²⁷

In epidemiological terms, the reduction of the time for completing the gonotrophic cycle has direct implications on the vectorial capacity of synanthropic mosquitoes because it increases the biting frequency of gonotrophic females on human hosts. Consequently, this physiological aspect constitutes an intrinsic factor that greatly affects the explosive nature of arboviral epidemics.^{19,32} In addition, the population structure of the

females may become an entomological parameter that might determine the transmission dynamics of arbovirus diseases because older females exhibit more partial feedings than younger females, and therefore older, infected females have more possibilities of contacting a larger number of hosts compared to younger females.³³ Climate change and environmental modifications may be important factors impacting the physiological capacities of females. Thus, it might be expected that the population dynamics of mosquitoes would be affected by the reduction of the duration of the biological cycles and the reproductive processes,³⁴ while the extrinsic incubation period of the etiological agents would decrease, changing the vectorial competence of the species and the condition of mosquitoes from uninfected to infective in a shorter time.^{28,29}

From an ecological point of view, the growth of the *Ae. aegypti* and *Ae. albopictus* populations will depend, in part, on the number of gonotrophic cycles and their length, as well as on their host preferences and fecundity.^{30,31} Thus, the physiological time of egg development and the gonotrophic cycle constitute a reproductive competence strategy for the establishment and spatial and temporal permanence of *Ae. aegypti* and *Ae. albopictus* populations in the community of urban, suburban and rural mosquitoes. At the same time, the

changes in temperature and humidity may affect the reproductive activity and influence the length of the gonotrophic cycle.²⁷ Moreover, it is likely that multiple oviposition with a single blood meal is the result of strategic behavior for the proliferation of mosquito populations in altered environments.³⁵ The pregravid stage in *Ae. aegypti* females indicates their need for more than one feeding to produce their first batch of eggs, which increases the contact and consequently their chances to become infected and later transmit diseases.³⁶

On the other hand, the morphometric dimensions and the structural appearance of the follicles of *Ae. aegypti* and *Ae. albopictus* were similar. However, comparison of the interstage longitudinal growth showed differences between the two species, while the interstage transversal growth only exhibited significant changes at the interval between Christophers' stages 4 and 5. In general, the step from one follicular stage to another was determined by the speed of digestion and assimilation of the blood.

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Declaration of conflict of interests. The authors declare that they have no conflict of interests.

References

- Weaver SC, Reisen WK. Present and future arboviral threats. *Antiviral Res.* 2010;85(2):328-45. <https://doi.org/10.1016/j.antiviral.2009.10.008>
- Ioos S, Mallet HP, Leparc Goffart I, Gauthier V, Cardoso T, Herida M. Current Zika virus epidemiology and recent epidemics. *Med Mal Infect.* 2014;44(7):302-7. <https://doi.org/10.1093/infdis/jix451>
- Defoliart GR, Watts DM, Grimstad PR. Changing patterns in mosquito-borne arboviruses. *J Am Mosq Control Assoc.* 1986;2(4):437-55.
- Halstead SB. Epidemiology. In: Halstead SB. *Dengue. Tropical Medicine: Science and Practice* Volume 5. London: Imperial Collage Press, 2008:75-122. https://doi.org/10.1142/9781848162297_0003
- Hawley WA. The biology of *Aedes albopictus*. *J Am Mosq Control Assoc.* 1988;1:1-39.
- Braks MAH, Honório NA, Lounibos LP, Lourenco-De-Oliveira R, Juliano SA. Interspecific competition between two invasive species of container mosquitoes, *Aedes aegypti* and *Aedes albopictus* (Diptera: Culicidae), in Brazil. *Ann Entomol Soc Am.* 2004;97(1):130-9. [https://doi.org/10.1603/0013-8746\(2004\)097\[0130:ICBTIS\]2.0.CO;2](https://doi.org/10.1603/0013-8746(2004)097[0130:ICBTIS]2.0.CO;2)
- Scott TW, Chow E, Strickman D, Kittayapong P, Wirtz RA, Lorenz LH, Edman JD. Blood-feeding patterns of *Aedes aegypti* (Diptera: Culicidae) collected in a rural Thai village. *J Med Entomol.* 1993;30(5):922-7. <https://doi.org/10.1093/jmedent/30.5.922>
- Niebylski ML, Savage HM, Nasci RS, Craig GB Jr. Blood hosts of *Aedes albopictus* in the United States. *J Am Mosq Control Assoc.* 1994;10(3):447-50.
- Casas-Martínez M, Torres-Estrada JL. First Evidence of *Aedes albopictus* (Skuse) in Southern Chiapas, Mexico. *Emerg Infect Dis.* 2003;9(5):606-7. <https://doi.org/10.3201/eid0905.020678>
- Casas-Martínez M, Orozco-Bonilla A, Muñoz-Reyes M, Ulloa-García A, Bond JG, Valle-Mora J, et al. A new tent trap for monitoring the daily activity of *Aedes aegypti* and *Aedes albopictus*. *J Vector Ecol.* 2013;38(2):277-88. <https://doi.org/10.1111/j.1948-7134.2013.12041.x>
- Pant CP, Yasuno M. Field studies on the gonotrophic cycle of *Aedes aegypti* in Bangkok, Thailand. *J Med Entomol.* 1973;25;10(2):219-23. <https://doi.org/10.1093/jmedent/10.2.219>
- Klowden MJ, Briegel H. Mosquito gonotrophic cycle and multiple feeding potential: contrasts between *Anopheles* and *Aedes* (Diptera: Culicidae). *J Med Entomol.* 1994;31(4):618-22. <https://doi.org/10.1093/jmedent/31.4.618>
- Lima-Camara TN, Honório NA, Lourenco-de-Oliveira R. Parity and ovarian development of *Aedes aegypti* and *Ae. albopictus* (Diptera: Culicidae) in metropolitan Rio de Janeiro. *J Vector Ecol.* 2007;32(1):34-40. [https://doi.org/10.3376/1081-1710\(2007\)32\[34:PAODOA\]2.0.CO;2](https://doi.org/10.3376/1081-1710(2007)32[34:PAODOA]2.0.CO;2)
- Savage HM, Smith GC. *Aedes albopictus* y *Aedes aegypti* en las Américas: implicaciones para la transmisión de arbovirus e identificación de hembras adultas dañadas. *Bol Oficina Sanit Panam.* 1995;118(6):473-7. <http://iris.paho.org/xmliui/handle/123456789/15585>
- Centers for Disease Control and Prevention. *Biología y control del Aedes aegypti*. Atlanta: CDC, 1980 [cited March 1, 2011] Available from: <https://stacks.cdc.gov/view/cdc/7579>
- Clements AN. *The biology of mosquitoes, Vol. 1: Development, nutrition and reproduction*. New York: CABI, 2000.
- Gillies MT. The recognition of age-groups within populations of *Anopheles gambiae* by the pre-gravid rate and the sporozoite rate. *Ann Trop Med Parasitol.* 1954;48(1):58-74. <https://doi.org/10.1080/00034983.1954.11685599>
- Christophers SR. The development of the egg follicle in Anophelines. *Paludism.* 1911;2:73-8.
- Mekuria Y, Granados R, Tidwell MA, Williams DC, Wirtz RA, Roberts DR. Malaria transmission potential by *Anopheles* mosquitoes of Dajabon, Dominican Republic. *J Am Mosq Control Assoc.* 1991;7(3):456-61.
- Bargielowski IE, Lounibos LP, Shin D, Smartt CT, Carrasquilla MC, Henry A, et al. Widespread evidence for interspecific mating between *Aedes aegypti* and *Aedes albopictus* (Diptera: Culicidae) in nature. *Infect Genet Evol.* 2015;36:456-61. <https://doi.org/10.1016/j.meegid.2015.08.016>
- Ulloa-García A, Gonzalez-Ceron L, Rodriguez MH. Host selection and gonotrophic cycle length of *Anopheles punctimacula* in southern Mexico. *J Am Mosq Control Assoc.* 2006;22:648-53. [https://doi.org/10.2987/8756-971X\(2006\)22\[648:HSAGCL\]2.0.CO;2](https://doi.org/10.2987/8756-971X(2006)22[648:HSAGCL]2.0.CO;2)
- Goindin D, Delannay C, Ramdini C, Gustave J, Fouque F. Parity and longevity of *Aedes aegypti* according to temperatures in controlled conditions and consequences on dengue transmission risks. *PLoS ONE.* 2015;10(8):e0135489. <https://doi.org/10.1371/journal.pone.0135489>
- Macdonald WW. *Aedes aegypti* in Malaya: II Larval and adult biology. *Ann Trop Med Parasitol.* 1956;50(4):399-414.

24. Gubler DJ, Bhattacharya N. Observations on the reproductive history of *Aedes (Stegomyia) albopictus* in the laboratory. *Mosq News*. 1971;31(3):356-9.
25. Reuben R. Feeding and reproduction in vector mosquitoes. *Proc Indian Acad Sci*. 1987;96:275-80. <https://doi.org/10.1007/BF03180010>
26. Briegel H. Metabolic relationship between female body size, reserves, and fecundity of *Aedes aegypti*. *J Insect Physiol*. 1990;36(3):165-72. [https://doi.org/10.1016/0022-1910\(90\)90118-Y](https://doi.org/10.1016/0022-1910(90)90118-Y)
27. Baak-Baak CM, Ulloa-García A, Cigarroa-Toledo N, Tzuc Dzul JC, Machain-Williams C, Torres-Chable OM, et al. Blood feeding status, gonotrophic cycle and survivorship of *Aedes (Stegomyia) aegypti* (L) (Diptera: Culicidae) caught in churches from Merida, Yucatan, Mexico. *Neotrop Entomol*. 2017;46(6):622-30. <https://doi.org/10.1007/s13744-017-0499-x>
28. Nelson MJ. *Aedes aegypti*: Biology and Ecology. Pan American Health Organization: Washington DC, 1986.
29. Day JF, Edman JD, Scott TW. Reproductive fitness and survivorship of *Aedes aegypti* (Diptera: Culicidae) maintained on blood, with field observations from Thailand. *J Med Entomol*. 1994;31(4):611-7.
30. Carrington LB, Armijos MV, Lambrechts L, Barker CM, Scott TW. Effects of fluctuating daily temperatures at critical thermal extremes on *Aedes aegypti* life-history traits. *PLoS ONE*. 2013;8(3):e58824. <https://doi.org/10.1371/journal.pone.0058824>
31. Gaio AO, Gusmão DS, Santos AV, Berbert-Molina MA, Pimenta PF, Lemos FJ. Contribution of midgut bacteria to blood digestion and egg production in *Aedes aegypti* (diptera: culicidae) (L). *Parasit Vectors*. 2011;4:105. <https://doi.org/10.1186/1756-3305-4-105>
32. Klowden MJ. Endogenous regulation of the attraction of *Aedes aegypti* mosquitoes. *J Am Mosq Control Assoc*. 1994;10(2):326-32.
33. Telang A, Wells MA. The effect of larval and adult nutrition on successful autogenous egg production by a mosquito. *J Insect Physiol*. 2004;50(7):677-85. <https://doi.org/10.1016/j.jinsphys.2004.05.001>
34. Telang A, Li Y, Noriega FG, Brown MR. Effects of larval nutrition on the endocrinology of mosquito egg development. *J Exp Biol*. 2006;209:645-55. <https://doi.org/10.1242/jeb.02026>
35. Scott TW, Takken W. Feeding strategies of anthropophilic mosquitoes result in increased risk of pathogen transmission. *Trends Parasitol*. 2012;28(3):114-21. <https://doi.org/10.1016/j.pt.2012.01.001>
36. Xue RD, Edman JD, Scott TW. Age and body size effects on blood meal size and multiple blood feeding by *Aedes aegypti* (Diptera: Culicidae). *J Med Entomol*. 1995;32(4):471-4. <https://doi.org/10.1093/jmedent/32.4.471>