VOL. 44 | NO. 2 | **MAY - AUGUST 2023** | PP 16-26



E-LOCATION ID: 1337

Electrochemical Studies of Magnesium Coated with Modified Chitosan and Electrosprayed as an Anticorrosive Protection Method in Bone Repair

Estudios Electroquímicos del Magnesio Recubierto con Quitosano Modificado y Electrorociado como Proceso de Protección Anticorrosiva en la Reparación de Hueso

José Luis Ramírez-Reyes¹ (D) (E), Deni Esperanza Gaytán-Macías² (D), Harlem Quintana-Camacho² (D), Gonzalo Galicia Aguilar¹ (D) Guillermina González Mancera³ (D) ¹Instituto de Ingeniería-Universidad Veracruzana, Veracruz - México ²Programa MIC-Instituto de Ingeniería-Universidad Veracruzana, Veracruz - México ³ULab-MEB-Facultad de Química-Universidad Nacional Autónoma de México - México

ABSTRACT

Magnesium (Mg) is essential for the metabolic reactions of the human body and is known for its biocompatibility, its mechanical and physical properties are similar to human bone, which is why it is considered to have high potential in *biomedical applications such as temporary and resorbable implants*. Through surface modifications, the high tendency to corrosion of Mg could be controlled, such as *biodegradable membranes* that prevent the passage of chloride ions present in the human organism. To prepare the membrane, solutions of chitosan modified with gelatin and/or glutaraldehyde are used and by means *of the electrospray method* applied to protect the Mg. To simulate body fluid conditions a Kokubo saline solution (BFK) was prepared. The study focuses on evaluating the corrosion rate of Mg with a coating made of a *chitosan electrosprayed membrane*, applying *electrochemical measurements of electrochemical impedance spectroscopy and linear polarization resistance*.

The key additive to improve the behavior of the membranes was observed with the use of gelatin, where the membrane with the best results lowing corrosion rates is the Mg CH+GE+GL system, which it was observed with very good physical integrity in the images of morphological analyzes of the surface after 30 days of exposure.

KEYWORDS: Bone Repair, chitosan modified, electrochemical techniques, electrosprayed coatings, magnesium

RESUMEN

El magnesio (Mg) es esencial para las reacciones metabólicas del cuerpo humano y es conocido por subiocompatibilidad, sus propiedades mecánicas y físicas son similares a las del hueso humano, por lo que se considera que tiene un alto potencial en *aplicaciones biomédicas como implantes temporales y reabsorbibles*. Mediante modificaciones superficiales se podría controlar la alta tendencia a la corrosión del Mg, como por ejemplo *membranas biodegradables* que impidan el paso de iones cloruro presentes en el organismo humano. Para preparar la membrana se utilizan soluciones de quitosano modificado con grenetina y/o glutaraldehído y mediante el *método de electrorociado* se aplican para proteger el Mg. Para simular las condiciones de los fluidos corporales se preparó una solución salina de Kokubo. El estudio se enfoca en evaluar la *velocidad de corrosión* del Mg con un recubrimiento hecho de una *membrana electrorociada con quitosano, aplicando técnicas electroquímicas de espectroscopia de impedancia electroquímica y resistencia de polarización lineal*. El aditivo clave para mejorar el comportamiento de las membranas se observó con el uso de gelatina, donde la membrana con mejores resultados bajando los índices de corrosión es el sistema Mg CH+GR+GL, el cual se observó con muy buena integridad física en las imágenes de análisis morfológicos de la superficie después de 30 días de exposición.

PALABRAS CLAVE: Magnesio, quitosano modificado, recubrimientos electro-rociados, reparación de hueso, técnicas electroquímicas

Corresponding author

TO: José Luis Ramírez-Reyes INSTITUTION: Instituto de Ingeniería-Universidad Veracruzana ADDRESS: SS Juan Pablo II, s/n, Zona Universitaria, Veracruz, Ver. Mex. C. P. 94294 CORREO ELECTRÓNICO: luiramirez@uv.mx Received: 2 February 2023 Accepted: 18 May 2023 REVISTA MEXICANA DE INGENIERÍA BIOMÉDICA | VOL. 44 | NO. 2 | MAY - AUGUST 2023

INTRODUCTION

Magnesium is an essential element for metabolic reactions in the human body and is known for its good biocompatibility, good mechanical properties (very close to human bone) and is considered a biodegradable material with potential in biomedical applications in the case of osteosynthesis, which is a process of intervention of fractures or fissures ^{[1][2]}. Its high electrochemical activity makes surface modification necessary in order to reduce corrosion rates. Thereby prolong its necessary mechanical properties throughout the recovery time of the bone tissue. Current investigations have focused on addressing this problem by implementing different strategies so that the rate of degradation of magnesium can be controlled within the physiological environment. Some of these are the use of magnesium alloys or protective techniques such as superficial modification treatments and protective recoveries [3]. However, in the surface modification of Mg, the required time for degradation depends on the targeted application, for instance in the design of coatings for tailor degradation is the unknown between the number of degradation test in the lab with the behavior of the coating in test in-vivo [4].

The main reactions reported for the Magnesium corrosion process are ^{[5][6]};

$$Mg \to Mg^{n+} + ne^{-} (\text{Anodic reacción}) \tag{1}$$
$$2H_2O + 2e^{-} \to H_2 + 2OH^{-} \tag{2}$$

 $2H_2O + 2e^- \rightarrow H_2 + 2OH^-$ (Cathodic reaction at acidic pH)

 $2H_2O + O_2 + 4e^- \rightarrow 4OH^-$ (3) (Cathodic reaction at neutral or alkaline pH)

$$Mg^{n+} + nOH^{-} \rightarrow Mg \ (OH)_{n}$$
(Global reaction products)
(4)

Chitosan membranes (CH) have excellent properties, such as antimicrobial activity, biocompatibility, good bio adhesiveness, it is non-toxic, it optimizes wound healing, bone formation, as well as a vehicle and/or drug releaser, and they are very efficient in guided bone regeneration ^{[7][8]}. Biodegradability is a property sought in these materials, however, pure chitosan has low mechanical properties due to its brittleness, it is soluble only in acidic media, and it loses its antibacterial activity at pH > 6.5.

To overcome these drawbacks, the most efficient method is to mix it with other polymers to provide it with fluidity such as gelatin which, in addition to fulfilling its function as an additive, enhances the antimicrobial property of chitosan in the form of a flexible film ^{[9][10][11][12][13]}.

There are many ways to produce these membranes; one example is to produce them using the electrospinning technique as electrospray, which generates micro-thin films with properties such as microporosity, large surface area, and biodegradable. Also, can be obtained by this technique as nanoparticles from a polymer in solution with a conductive solvent ^[14]

The first appearance of this technique was reported by Nollet from the 17th century, when he observed that *"a person, electrified by connection to a high-voltage generator, would not bleed normally if cut; the blood would come out of the wound in the form of drops"*. After that, Rayleigh defined a limit on the charge carried by a drop in 1882, Zeleny reported various modes of operation of electrospray during 1914, and Taylor determined the shape of the cone formed by the fluid at the capillary tip in an electric field in 1964. Widespread use of electrospray began after Dole introduced it as a method of generating gas-phase ions for mass spectrometry analysis in 1968 ^[17].

The present work focuses on evaluating the corrosion rate of structural Mg with a coating made of an electrosprayed chitosan membrane (CH) modified with gelatin (GE) and/or glutaraldehyde (GL). The electrochemical evaluation was carried out using the techniques of electrochemical impedance spectroscopy (EIS) and linear polarization resistance (LPR), applied José Luis Ramírez-Reyes et al. Electrochemical Studies of Magnesium Coated with Modified Chitosan and Electrosprayed as an Anticorrosive Protection Method in Bone Repair

by means of a potentiostat to the Mg samples bare and coated with the membranes, to evaluate the process of degradation of the membrane and the anticorrosive protection to the Mg. Also, by means of surface modification treatment and evaluating in a Kokubo solution (BFK) at 37 °C body temperature. In addition to the challenges in this job, still hard research is being carried out for the control of biodegradation of Mg alloys, and for developing new type of surface modification approaches ^[4].

MATERIALS AND METHODS

Mg discs of 1 cm in diameter and 3 mm thick were used. They were roughened up to 600 grit sandpaper and ultrasonic washed together with chemical roughening with ethanol and acetic acid, as shown in Figure 1.



FIGURE 1. Ultrasonic bath and morphological surface of washed Mg before the phosphate treatment.

Kokubo Saline Solution (BFK)

All the used reactives were grade ACS and prepared for one L of solution following the methodology of Tadashi Kokubo ^{[7] [18] [19]} as shown in Table 1.

Order	Reactive	Quantity, g	Conc., mM
#1	NaCl	7.996	136.7
#2	NaHCO ₃	0.350	4.16
#3	KCl	0.224	3
#4	K ₂ HPO ₄ ·3H ₂ O	0.1764	0.7737
#5	MgCl ₂ ·6H ₂ O	0.305	1.5
#6	1 kmol/m ³ HCl	40 cm ³	40
#7	CaCl ₂	0.278	2.5
#8	Na ₂ SO ₄	0.071	0.5
#9	(CH ₂ OH)3CNH ₂	6.057	48.46
#10	1 kmol/m ³ HCl	pН	
		Adjusting	

TABLE 1. Chemical composition of the Kokubo solution (BFK) and the sequence order for the preparation. For the formation of the membrane, chitosan solutions were prepared at 1.5 % diluted concentration of a mixture of acetic acid and distilled water in a ratio of 9:1, mixing everything at a temperature of 60 ± 5 °C until dissolved. To compare with chitosan modified with gelatin and/or glutaraldehyde, it was carried out by adding 2 % gelatin in the solution together with chitosan, the modification with glutaraldehyde is done after the formation of the membrane, placing a drop of it on the coating, letting it absorb and evaporate.

The membranes were made with the Fluidnatek LE-100 used as electro spraying equipment, placing 3 ml of the chitosan solution in a syringe, as shown in Figure 2. The images show the membranes a) Chitosan, b) Chitosan modified with gelatin, c) Chitosan modified with gelatin + glutaraldehyde and d) Chitosan modified with gelatin + glutaraldehyde. The Figure 3 show an image SEM with the dispersion of the microparticles contained in drops of chitosan at the contact with the Mg substrate. The processing parameters were as follows: applied voltage, -30 kV; feed solution flow rate, 0.3 ml/hr; distance between the nozzle and the substrate, 10 cm; deposit time, 30 min.



FIGURE 2. Scheme of the spray process and samples with the sprayed membranes a) MgCH, b) MgCH+GE, c) MgCH+GL and d) MgCH+GE+GL over discs of Mg.



FIGURE 3. Image SEM dispersion of the microparticles contained in drops of chitosan at Mg substrate.

Once the samples with the membranes were ready, an electrochemical cell was prepared in a simulated Kokubo physiological body solution (BFK) and using a temperature of 37 °C as shown in the cell arrangement in Figure 4. The impedance spectroscopy technique (EIS) was carried out, using a frequency sweep from 10kHz to 0.01 Hz, with an amplitude of ± 10 mV peakto-peak and 7 points per decade of frequency and the linear polarization resistance (LPR) where a bias of \pm 20 mV/Ecorr was used. The corrosion potential (Ecorr), wasmonitored with respect to time. The electrochemical tests were carried out with a Bio-Logic brand potentiostat model SP-150, where the electrochemical monitoring was carried out from the first hour of immersion and subsequently measured every 24 hours for a period of 30 days.



FIGURE 4. Scheme and configuration of the electrochemical cell for the electrochemical evaluation.

Some of the main chloride reactions expected in the Mg -BFK solution are described below;

$$Mg_{(s)} + 2H_2O_{(l)} \rightarrow Mg(OH)_{2(s)} + H_{2(g)}$$

$$\tag{5}$$

$$2Mg_{(s)} + O_{2(g)} + 2H_2O_{(l)} \rightarrow 2Mg(OH)_{2(s)}$$

$$\tag{6}$$

$$Mg(OH)_{2(s)} \rightarrow MgO_{(s)} + H_2O_{(l)}$$
(7)

$$MgO_{(s)} + H_2O_{(l)} \to Mg(OH)_{2(s)}$$
(8)

$$Mg(OH)_{2(s)} + 2NaCl_{(s)} \rightarrow MgCl_{2(s)} + 2NaOH_{(s)}$$
⁽⁹⁾

For the analysis of the surface morphology of the samples before and after exposure, a high-low vacuum scanning electron microscope, model SEM 6000 NEOSCOPE with a voltage of 150 keV, was used to perform the elemental chemical analysis of the membranes, the X-ray scattering technique known as EDS was used.

RESULTS AND DISCUSSION

The corrosion potential (Ecorr) and corrosion rates (CR) results are shown in Figure 5 and Figure 6 respectively, where we can see the same trend for bare Mg, Mg with chitosan membrane (Mg CH) and Mg with gelatin-modified chitosan (Mg CH+GE) and glutaraldehyde (Mg CH+GL), however, for Mg with the membrane modified with gelatin and glutaraldehyde (Mg CH+GE+GL) was observed that the Ecorr tends to rise more positive values, which implies an energy gain. This translates into thermodynamic stability for the reactivity of the species involved in the interface of the evaluated systems and the tendency of CR to decrease, implying that the metal-coating systems tend to improve the resistance against the corrosiveness of the environment.

Corrosion mechanism of Mg alloys in simulated body fluid always has been complicated because of many factors like the complex chemical composition influence. However, the obtained CR values between 0.1 to 2 mm/y are in coincidence for in vitro conditions reported values ^[4].



FIGURE 5. Ecorr of bare and coated Mg at the immersion time measurements in the BFK solution at 37 °C.



The Figure 7 is the Nyquist diagram of bare Mg and clearly shows the corrosion process suffered by magnesium with resistance up to 1.4 k Ω -cm2 before the presence of the coating. In the case of Mg coated with chitosan, we can see an increase in the corrosion resistance except for day 30, where it decreased due to the total degradation of the membrane and that exposed to the metal. For the chitosan membrane shown in the Nyquist of Figure 8, the resistance was

increased by twice and no degradation is observed in the membrane.



FIGURE 7. Nyquist diagrams of the bare Mg in the BFK solution at 37 °C the immersion time.



FIGURE 8. Nyquist diagrams of the Mg coated with chitosan and immersed in BFK at 37ºC.

For the gelatin-modified chitosan membrane shown in the Nyquist of Figure 9, the corrosion resistance was 3 times higher, and less degradation after 30 days as observed in the membrane. The Mg with chitosan modified with glutaraldehyde which is shown in the Nyquist of Figure 10 shows that the corrosion resistance is similar to the bare Mg and if a degradation of the membrane is observed on day 30.



FIGURE 9. Nyquist diagrams of the Mg coated with chitosan and modified with gelatin and immersed in BFK solution.



FIGURE 10. Nyquist diagrams of the Mg coated with chitosan and modified with glutaraldehyde and immersed in BFK solution.

The Nyquist diagram for the gelatin and glutaraldehyde modified chitosan coated Mg is shown in the Nyquist of Figure 11, it can be clearly seen that the corrosion resistance is extremely large compared to the other membranes, the corrosion process becomes slower and the membrane makes it difficult for chloride ions to reach Mg. In the Figure 12 the spectra obtained from Raman Spectroscopy of bare Mg, chitosan, and the different Mg+ membrane systems are shown, and observing the covering capacity of the additives CH, GE and GL over the Mg substrate. The diagram shows the peaks of 1050 cm-1 for Mg and 2885 cm-1 reported in the literature as characteristic of chitosan ^[20].



FIGURE 11. Nyquist diagrams of the Mg coated with chitosan modified with gelatin plus glutaraldehyde and immersed in BFK.



FIGURE 12. Raman Spectroscopy response obtained from the bare and coated Mg with membranes of chitosan modified with gelatin and glutaraldehyde.



FIGURE 13. SEM microscopy of samples after exposition to the BFK solution a) Bare Mg, b) Mg with chitosan, c) Mg with chitosan modified with gelatin and d) Mg with chitosan modified with grenetine plus glutaraldehyde.

The Figure 13 show the SEM images of (a) corrosion products on the bare Mg, (b) coated Mg with chitosan, showing morphology with less corrosion than the bare Mg, (c) Mg coated with chitosan plus the gelatin, showing the dispersion of microspheres of the membrane, controlling the embrittlement of the coating and reducing the corrosion rate on the substrate and (d), the formed membrane practically intact, well formed and smooth, over the entire surface due to the cross-linking reaction produced by glutaraldehyde with chitosan, in addition to increasing mechanical resistance and flexibility given by the gelatin, showing the best anticorrosive effect over the Mg substrate.

Elem	Mg	Mg	Mg-	Mg-	Mg-
ent	without	attack	CH	CH+	CH+
	attack	ed		GE	GE+
					GL
С	25.17	27.05	31.03	44.52	43.8
0	1.6	36.08	42.2	30.55	32.2
Mg	73.23	32.2	12.1	4.9	5.3
Si		0.05	0.03	0.08	0.21
Р		1.08	6.2	4.15	4.5
S		0.28	0.08	0.2	0.07
Cl		0.06	0.05	6.85	0.03
K		0.13	0.17		0.69
Na		0.28	0.87	0.26	0.97
Ca		1.98	6.1	5.7	11.8
Al		0.10	0.04	0.05	0.19
Total	100%	100%	100%	100%	100%

TABLE 2. Elemental chemical composition detected in % mass by EDS on the surface of samples of coated Mg before and after the exposition to the BFK solution. Table 2 shows the results of the elemental chemical composition for the Mg samples and chitosan membranes, where it is observed that in the exposed Mg samples it registers the main elements (O, P and Ca) of the BFK solution, while the chitosan membranes raise the C, O, P and Ca content, lowering the detection of Mg due to the coating and the gelatin involves the presence of elements such as C, O, Cl, and Ca mainly, and when the coating system includes the glutaraldehyde the Si, Al and Ca increases such a bit, maybe due the trapping effect of such elements in the formed lattice. Then, as was observed in the EIS response, the best results were observed with the membrane made of CH+ GE+ GL.

In the literature there are many in-vitro studies that deal with a variety of topics, some evaluate and compare electrolytes, others evaluate the surface morphology of coatings such as chitosan ^[21]. others report the behavior of Mg alloys with other metals such as Zn, Ca, and others ^[4]. There are even several reviewtype papers on the applications of Mg and its alloys as a biomaterial ^[22], but in no work has the application of chitosan membranes modified with gelatin and glutaraldehyde been studied, and due to the results obtained, it is considered a contribution that deserves further studies with a view to its application at in-vivo conditions, even the membranes can also be used as a vehicle for the transport of drugs in post-operative treatments ^[23].

CONCLUSIONS

The membrane that degraded the least and had the best results with the lowest corrosion rates is that of the Mg-CH+GE+GL system and it is observed almost intact in the SEM results. The Raman information of the membranes, helped in the interpreting of the covering properties of the additives.

The corrosion protection also can be attributed to the synergistic effect of such additives; the gelatin serves

as a means for the chitosan to have greater resistance since it provides viscoelasticity, flexibility and mobility properties. Glutaraldehyde serves as a cross-linking agent between the chitosan polymer chains, creating greater cohesion in the membrane, improving its mechanical properties and its resistance to protect the Mg substrate against the corrosiveness of the medium.

With the results of the electrochemical evaluation, it was verified that the chitosan with the gelatin and glutaraldehyde acting independently, do not reach the anticorrosive protection properties and only by acting the whole system together is a synergistic effect achieved. The challenge in both strategies is to combine control of the corrosion behavior with the membrane biodegradation and desired biological performance for the in-vivo applications.

The electrospray process of the polymeric substances is another important factor due to the distribution and better coverage area provided by the microparticles that form the membrane. Also is recommended the chemical characterization of the membranes in the near future.

AKNOWLEGEMENTS

To the CONACYT for the scholarships for the Master in Corrosion Engineering students and their support in carrying out this research work.

CONFLICTS OF INTREST

The authors declare no conflict of interest.

AUTHOR CONTRIBUTIONS

D.E.G.M. conceptualized the project, participated in all the writing stages of the manuscript (preparation of the original draft, review and edition of the different versions and the final document), performed data curation, contributed to the research and oversaw the project. J.L.R.R. Designed and carried out experiments, performed statistical analyses, participated in all the writing stages of the manuscript, carried out analyses. H.Q.C. performed data curation, carried out and validated analyses, participated in the review of the final version of the manuscript. G.G.M. provided resources, designed and developed methodology and performed analyses SEM. G.G.A. carried out formal analyses, reviewed and edited the final version of the manuscript. All authors reviewed and approved the final version of the manuscript. José Luis Ramírez-Reyes et al. Electrochemical Studies of Magnesium Coated with Modified Chitosan and Electrosprayed as an Anticorrosive Protection Method in Bone Repair

REFERENCES

- N. T. Kirkland, N. Birbilis, M. Staiger, "Assessing the corrosion of biodegradable magnesium implants: a critical review of current methodologies and their limitations," Acta Biomater., vol. 8, no. 3, pp. 925-936, Mar. 2012. doi: https://doi.org/10.1016/j.actbio.2011.11.014
- N. Ardila, Z. Ajji, M.-C. Heuzey, A. Ajji, "Chitosan electrospraying: Mapping of process stability and micro and nanoparticle formation," J. Aerosol Sci., vol. 126, pp. 85-98, Dec. 2018, doi: <u>https://doi.org/10.1016/j.jaerosci.2018.08.010</u>
- M. Carboneras Chamorro, C. Iglesias Urraca, E. Onofre Bustamante, M. A. Alobera Gracia, C. Clemente de Arriba, M.C. García Alonso, M. L. Escudero Rincón, "Materiales metálicos biodegradables en el campo biomédico," Acta Cienc. Tecnol., vol. 19, pp. 30-34, 2011. [Online]. Available: https://core.ac.uk/reader/36133460
- M. Esmaily, J.E. Svensson, S. Fajardo, N. Birbilis, G.S. Frankel, S. Virtanen, R. Arrabal, S. Thomas, L.G. Johans, "Fundamentals and advances in magnesiun alloy corrosion," Prog. Mater. Sci., vol. 89, pp. 92-193, Aug. 2017. doi: https://doi.org/10.1016/j.pmatsci.2017.04.011
- [5] Y. Zheng, Magnesium Alloys as Degradable Biomaterials, 1st ed. Boca Ratón: CRC Press, 2019, pp. 600.
- [6] Z. Song, G. Yu, Z. Xie, B. Hu, X. He, X. Zhang (2014). "Performance of composite coating on AZ31B magnesium alloy prepared by anodic polarization and electroless electrophoresis coating," Surf. Coat. Technol. vol. 242, pp. 83-91, Mar. 2014. doi: https://doi. org/10.1016/j.surfcoat.2014.01.022
- [7] V. Valencia Goujon, "Estudio electroquímico de magnesio en medio fisiológico simulado (Kokubo) para su aplicación como biomaterial y uso de tratamientos de modificación superficial anticorrosivos," Bachelors dissertation, Univ. Nac. Aut. Mex., Mexico City, Mexico, 2013. [Online]. Available: <u>https://repositorio.unam.mx/contenidos/353329</u>
- [8] A.S. Paredes, O.S. Orteg, A. González, L. Bustillos, "Análisis comparativo de la regeneración ósea obtenida con quitosano y plasma rico en fibrina," Acta Odontol. Venez., vol. 52, no. 2, Art. no. 2, 2014.
 [Online]. Available: https://www.actaodontologica.com/ediciones/2014/2/art-2
- [9] C.A. Cárcamo Gatica, "Preparación de films de complejo polielectrolito quitosano-alginato y comparación de sus propiedades mecánicas y biológicas con films de quitosan," Bachelors dissertation, Univ. Chile, Santiago, Chile, 2005. [Online]. Available: <u>https://repositorio. uchile.cl/handle/2250/105463</u>
- [10] A.E. Mochalova, L.V. Nikishchenkova, N.N. Smirnova, L.A. Smirnova, "Thermodynamic properties of chitosan-based hydrogels in the range 0-350 K," Polym. Sci. Ser. B, vol. 49, pp. 42-46, Apr. 2007. doi: https://doi.org/10.1134/51560090407010101

- [11] Y. Liu, S. Wang, W. Lan, "Fabrication of antibacterial chitosan-PVA blended film using electrospray technique for food packaging applications," Int. J. Biol. Macromol., vol. 107, no. Pt A, pp. 848-854, Feb. 2018. doi: <u>https://doi.org/10.1016/j.ijbiomac.2017.09.044</u>
- [12] N.T.-P. Nguyen, L.V.-H. Nguyen, N.T. Thanh, V.V. Toi, T. Ngoc Quyen, P.A. Tran, H.-M. David Wang, T.-H. Nguyen, "Stabilization of silver nanoparticles in chitosan and gelatin hydrogel and its applications," Mater. Lett., vol. 248, pp. 241-245, Aug. 2019. doi: <u>https://doi.org/10.1016/j.matlet.2019.03.103</u>
- [13] H. Quintana-Camacho, J.L. Ramírez-Reyes, A.L. Medina-Almazán, N. García-Navarro, G. Galicia-Aguilar, "Investigation of the corrosion behavior of phophate coating magnesium in a kokubo solution," Int. J. Electrochem. Sci., vol. 13, pp. 6072-6082, 2018. doi: <u>https://doi.org/10.20964/2018.06.53</u>
- [14] J.A.S. Moreno, A.C. Mendes, K. Stephansen, C. Engwer, F.M. Goycoolea, A. Boisen, L.H. Nielsen, I.S. Chronakis, "Development of electrosprayed mucoadhesive chitosan microparticles," Carbohydr. Polym., vol. 190, pp. 240-247, Jun. 2018. doi: <u>https://doi. org/10.1016/j.carbpol.2018.02.062</u>
- [15] R.M.D. Soares, N.M. Siqueira, M.P. Prabhakaram, S. Ramakrishna, "Electrospinning and electrospray of bio-based and natural polymers for biomaterials development," Mater. Sci. Eng. C., vol. 92, pp. 969-982, Nov. 2018. doi: https://doi.org/10.1016/j.msec.2018.08.004
- [16] A. Jaworek, A.T. Sobczyk, A. Krupa, "Electrospray application to powder production and surface coating," J. Aerosol Sci., vol. 125, pp. 57-92, Nov. 2018. doi: <u>https://doi.org/10.1016/j.jaerosci.2018.04.006</u>
- [17] S. Kavadiya, P. Biswas, "Electrospray deposition of biomolecules: Applications, challenges, and recommendations," J. Aerosol Sci., vol. 125, pp. 182-207, Nov. 2018. doi: <u>https://doi.org/10.1016/j.jaero-sci.2018.04.009</u>
- [18] T. Kokubo, "Bioactive glass ceramics: properties and applications," Biomaterials, vol. 12, no. 2, pp. 155-163, Mar. 1991. doi: <u>https://doi.org/10.1016/0142-9612(91)90194-F</u>
- [19] J.Y. Wong, J.D. Bronzino, D.R. Peterson, Biomaterials: Principles and Practices, 1st Ed. Boca Ratón: CRC Press, 2012, pp. 288.
- [20] A. Zaja c, J. Hanuza, M. Wandas, L. Dymińska, "Determination of N-acetylation degree in chitosan using Raman spectroscopy," Spectrochim. Acta A Mol. Biomol. Spectrosc., vol. 134, pp. 114-120, Jan. 2015. doi: https://doi.org/10.1016/j.saa.2014.06.071
- [21] G.A.S. Kazi, T. Yamanaka, Y. Osamu, "Chitosan coating an efficient approach to improve the substrate surface for in vitro cultura System," J. Electrochem. Soc., vol. 166, no. 9, pp. B3025-B3030, 2019. doi: http://dx.doi.org/10.1149/2.0051909jes

- [22] S. Sobieszcyk, A. Zieliński, "Coatings in Arthoplastiy: Review Paper," Adv. Mater. Sci., vol. 8, no. 4, Dec. 2008. [Online]. Available <u>https://www.proquest.com/openview/e71b8f58cff352959f14f9b5dc1f-870d/1?pq-origsite=gscholar&cbl=2016339</u>
- [23] B. Pérez-Artacho, E. Sáez-Fernández, G.I. Martínez-Soler, V. Gallardo Lara, J.L. Arias Mediano, "Polímeros biodegradables en el transporte selectivo de moléculas antitumorales," ARS Pharm. vol. 51, s. 3, pp. 171-176, 2010. [Online]. Available: <u>http://hdl.handle.</u> <u>net/10481/26416</u>