



MOLECULAR AND GENETIC MECHANISMS OF NEUROTOXICITY DURING ANTI-SEIZURE MEDICATIONS USE

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ABSTRACT

Epilepsy is a multifactorial pathology that has allowed the development of various drugs aiming to combat it. This effort was formally initiated in the 1940s when phenytoin began to be used. It eventually turned out to be a drug with great anticonvulsant efficacy. At present, several potentially good new generation anti-seizure medications (ASMs) have been developed. Most of them present more tolerability and less toxic effects. However, they continue to have adverse effects at different levels. In addition, some seizures are difficult to treat with ASMs, representing 30% of the total cases of people who suffer from epilepsy. This review aims to explore the genetic and molecular mechanisms of ASMs neurotoxicity, proposing the study of damage caused by epileptic seizures, in addition to the deterioration generated by anti-seizure drug administration within the central nervous system. It is beyond question that there is a need to develop drugs that lower the lower the risk of secondary and toxic effects of ASMs. Simultaneously, we must find strategies that produce fewer harmful interactions and more health benefits when taking anti-seizure drugs. (REV INVEST CLIN. 2023;75(1):1-12)

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INTRODUCTION

The high prevalence of epilepsy affects more than 70 million people worldwide¹. Thus, different anti-seizure medications (ASMs) – each with its molecular mechanism – have been developed since the 20th century. The International League Against Epilepsy (ILAE) defines epilepsy as a brain disease with any of

the following characteristics: (1) two unprovoked seizures separated by 24 h; (2) an unprovoked crisis with a risk of recurrence within 10 years similar to the risk of recurrence given by having two induced seizures (60%); and (3) a diagnosed epileptic syndrome². When patients have been diagnosed, the first line of control is the prescription of ASMs³. However, on many occasions, these drugs' use can generate alterations

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in their absorption, metabolism, and degradation pathway of the ASMs.

Seizures caused by abnormal, excessive, or concurrent electrical activity in the brain are known as epileptic seizures. Neuronal hyperexcitability in epilepsy is caused by an imbalance between excitatory and inhibitory ionic currents. As a result, most ASMs target Na^+ , Ca^{2+} , K^+ , and Cl^- ion channels, along with membrane receptors: primarily glutamate and gamma-aminobutyric acid (GABA)¹. Nevertheless, ASMs controlled seizures are not utterly safe for the body. These drugs are known to have serious side effects (Table 1). As a result, it is critical to evaluate the patient's clinical history and their response to treatment before initiating a pharmacological treatment to minimize the possible adverse effects. In some cases, the use of ASMs is debatable⁴. For example, the pharmacological treatment for benign epilepsy with centrotemporal spikes appears to be unnecessary if it is considered to come from a benign origin⁴. ASMs resistance affects up to one-third of patients⁵, posing a significant challenge in neurology to discover safer and more effective ASMs.

Some medications may predispose patients to epileptic seizures as a side effect of their therapeutic use⁶. This would add another pathway of neurotoxicity in both epileptic and non-epileptic people. In some cases, these epileptogenic pathways have been exploited as experimental models of epilepsy, which has aided in new treatment findings. Similarly, exposure to certain toxins can cause epileptic seizures. In this regard, certain toxins have been used as effective pharmacological compounds as experimental models of epilepsy. This review aims to analyze the molecular and genetic mechanisms of neurotoxicity involved in epilepsy, both damages caused by epileptic seizures and the therapeutic administration of ASMs targeting the central nervous system (CNS) neurotoxicity in epileptic seizures induced by other drugs and chemicals is reviewed.

MOLECULAR MECHANISMS OF ANTIEPILEPTIC DRUGS

To understand the ASMs molecular mechanisms of neurotoxicity, it is important to remember that these drugs act primarily by either inhibiting or exciting ion channels (Fig. 1). The resulting modification of

signaling pathways will be crucial in triggering a range of adverse effects as described below.

Sodium channels

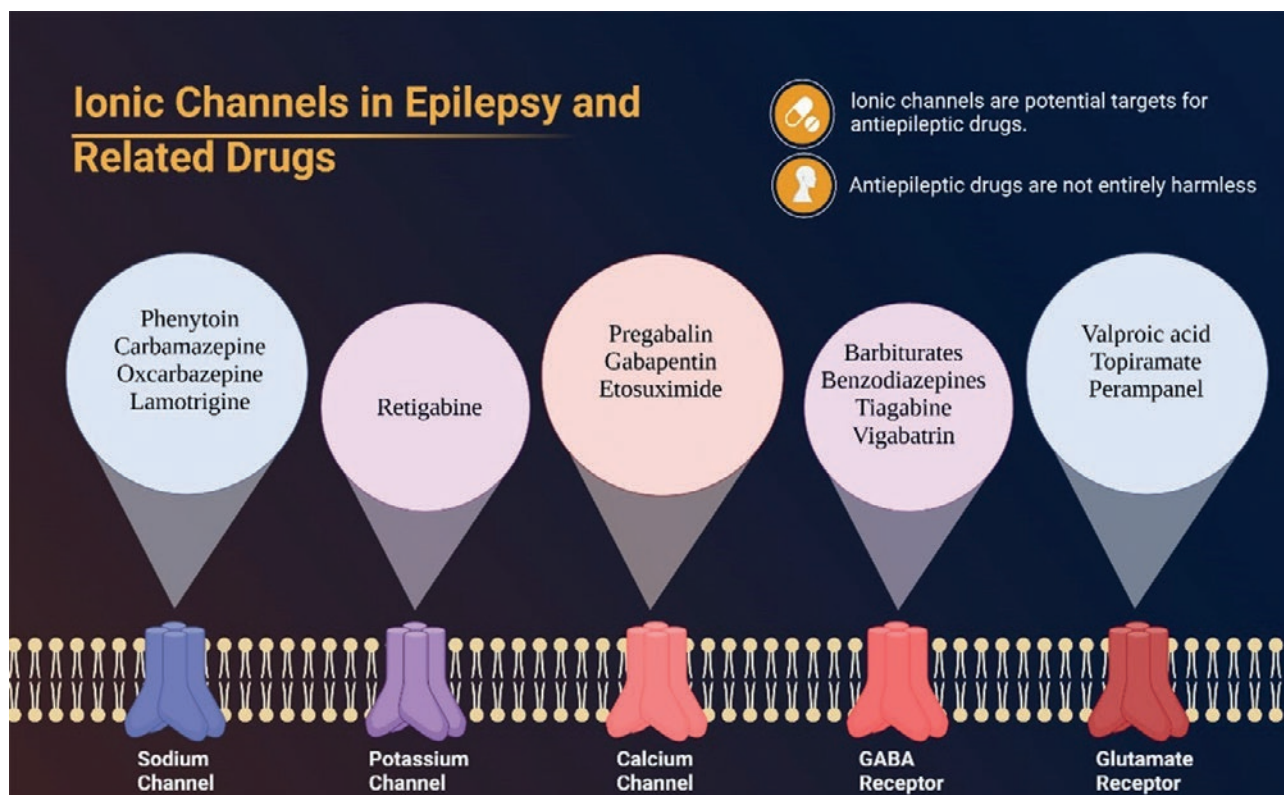
Gated-dependent sodium channels are heteromeric complexes, composed of α and one or two β subunits encoded by *SCN1A* and *SCN1B*, respectively. *SCN1A*, *SCN2A*, *SCN3A*, and *SCN8A* genes, code for Nav1.1, Nav1.2, Nav1.3, and Nav1.6 channel subtypes, the primary sodium channels in the CNS. The α subunit is made up of four homologous but distinct domains (DI-DIV), and each of them contains six transmembrane segments (S1-S6). A central pore is produced between the four domains, in which the segments S5, S6, and the connecting pore-loops (P-loops) form the channel; while segments S1-S4 form the voltage-sensing domain⁷. *SCN2A*, *SCN3A*, and *SCN8A* genes are expressed mainly in excitatory neurons, whereas inhibitory interneurons express predominantly *SCN1A* and a low expression of *SCN8A*⁸. The expression – as well as the function of each channel – varies despite the similarity that exists between each of them; therefore, the presence of mutations or polymorphisms results in the manifestation of different seizure patterns and syndromes. Most of the mutations are found in the *SCN1* gene. Mutation in *SCN1A* (Nav1.1) and *SCN2A* (Nav1.2) causes several subtypes of dominant idiopathic-generalized epilepsy (IGE)⁹. Particularly, *SCN1A* mutations are responsible for some types of epilepsy as Dravet syndrome and generalized epilepsy with febrile seizures (FS) plus. Some ASMs act on sodium channels, using their two states (closed and open state) to trigger their molecular mechanism¹⁰. The channel remains closed during the resting membrane potential, then quickly opens after depolarization and ultimately shuts to an inactive state. Depolarization is the physiological underpinning of epileptogenesis and the optimal target for ASMs since it is reliant on the sodium current (INa)¹¹. INa blockers stabilize deactivated Na^+ channels include phenytoin (PHT), carbamazepine, oxcarbazepine, and lamotrigine. Lacosamide (LCM) and eslicarbazepine are sodium channel blockers with a delayed inactivation rate¹². The drug rufinamide has important adverse effects that occur in up to 60% of patients, including headaches, vomiting, dizziness, vertigo, drowsiness, and anorexia among others. These effects may be transient and have been reported to improve over time¹³.

Table 1. Pharmacodynamics and adverse effects of anti-seizure medications

Antiepileptic name	Mechanism of action	Adverse effects/toxicity
Acetazolamide	Inhibition of the carbonic anhydrase	Metabolic acidosis, nephrolithiasis, nausea, vomiting, paresthesia, and metallic taste
Benzodiazepines	Increase the frequency of Cl ⁻ -channel opening of GABA A receptor	Hypotension, respiratory depression, nystagmus, apnea, drowsiness, ataxia, sedation, and cardiac arrest
Brivaracetam	Binding to the SV2A protein	Dizziness, drowsiness, and flu symptoms
Carbamazepine	Reduction of neuronal action potential, inhibition of NMDA-activated ion current, blocking of adenosine receptor	SIADH, fatigue, diplopia, ataxia, leukopenia, and skin rash
Ethosuximide	Antagonism of the postsynaptic T-type voltage-gated calcium channel	Drowsiness, insomnia, headache, ataxia, nausea, vomiting, diarrhea, anorexia, and hiccups
Fenfluramine	Increased extracellular serotonin levels, serotonin reuptake inhibition	Anorexia, drowsiness, ataxia, sedation, serotonin syndrome, pulmonary arterial hypertension, and valvular heart disease
Gabapentin	Inhibition of the $\alpha\delta$ calcium channel subunit	Dizziness, ataxia, edema, nystagmus, weight gain, and respiratory depression
Lacosamide	Slow inactivation of voltage-gated sodium channels	Drowsiness, dizziness, ataxia, diplopia, nystagmus, tremor, and headache
Lamotrigine	Sodium channel blocker, decrease extracellular glutamate and aspartate	Skin rash, fatigue, drowsiness, dizziness, diplopia, leukopenia, and insomnia
Levetiracetam	Binding to the SV2A protein, inhibition of presynaptic calcium channels	Drowsiness, dizziness, emotional lability, and headache
Oxcarbazepine	Block of voltage-gated sodium and calcium channels	Headache, dizziness, drowsiness, diplopia, nausea, dyspepsia, and hyponatremia
Phenobarbital (Barbiturates)	Increase the time of Cl ⁻ -channel opening of GABA A receptor	Dizziness, nystagmus, ataxia, bradycardia, bradypnea, hypothermia, and hypotension
Phenytoin	Block of voltage-gated sodium channels	Nystagmus, diplopia, sedation, ataxia, hypotension, nausea, vomiting, folate deficiency, gingival enlargement, hypertrichosis, drug-induced lupus, and anorexia
Piracetam	AMPA receptor modulator, improves acetylcholine function	Insomnia, irritability, headache, tremor, weight gain, agitation, and bleeding
Pregabalin	Inhibition of the $\alpha\delta$ calcium channel subunit	Dizziness, drowsiness, weight gain, blurred vision, diplopia, edema, dysarthria, and myalgia
Tiagabine	GABA reuptake inhibition	Dizziness, asthenia, tremor, headache, memory impairment, and psychosis
Topiramate	Block of voltage-gated sodium and calcium channels, inhibition of the carbonic anhydrase, block of AMPA receptor	Dizziness, weight loss, nausea, diarrhea, rhinitis, asthenia, myopia, and glaucoma
Valproate	Block of voltage-gated sodium channels, increase GABA levels, inhibition of histone deacetylases	Drowsiness, anorexia, nausea, dizziness, alopecia, encephalopathy, hepatotoxicity, pancreatitis, and polycystic ovary syndrome
Vigabatrin	Inhibition of the GABA aminotransferase	Headache, insomnia, vision loss, diplopia, drowsiness, memory impairment, psychosis, and fatigue
Zonisamide	Block of sodium and T-type calcium channels, inhibition of the carbonic anhydrase	Dizziness, irritability, diplopia, memory impairment, depression, insomnia, paresthesia, and metabolic acidosis

GABA: gamma-aminobutyric acid; SV2A: synaptic vesicle protein 2A.

Figure 1. Ionic channels involved in epilepsy's pathogenesis and related drugs that act over them. Notice that some channels may be excitatory or inhibitory.



Potassium channels

Voltage-gated potassium (Kv) channels are integral membrane proteins, belonging to the largest ion channel family in the human, encoded by 40 genes, and divided into 12 subfamilies (Kv1-Kv12). They play an important role in the action potential generation and propagation. Besides, Kv channels regulate the threshold potential for firing, the duration of action potentials, and the firing rates. Kvs' physiological role is defined by their localization within neuronal compartments¹⁴. The Kv channel is a homotetramer composed of four identical alpha (α) subunits that generate a potassium ion-selective pore in cell membranes¹⁴. Each α subunit has six α -transmembrane domains (TMD) (S1–S6), a membrane-entering P-loop located between the S5 and S6 domains, and cytosolic N- and C-terminus. The pore is formed by the segments S5–P–S6, while the sequences S1–S4 are essential for voltage sensing and channel activation¹⁵. These types of channels increase their functional

diversity by assembling with other types of auxiliary subunits (β -subunit, Kv β), which by themselves do not have a functional activity; but alterations in them can affect Kv channel function and network excitability, leading to increased seizure susceptibility¹³⁻¹⁵.

Voltage-gated dependent potassium channels are also involved in several processes such as cellular proliferation, migration, neurotoxicity, neuroprotection, and neuroregulation. More than 300 *KCNQ1*, *KCNQ2*, and *KCNQ3* mutations have been identified to cause epilepsy including epileptic encephalopathy (EE) and benign familial neonatal epilepsy¹⁶. Due to their repolarizing actions, these channels have been explored pharmacologically to treat epilepsy. One example is retigabine, an activator of potassium channels Kv 7.2/7.3.¹⁷ Retigabine is indicated for resistant partial seizure treatment, with or without secondary generalization. Patients receiving retigabine as treatment should be under strict medical supervision, because this drug is capable of producing changes in vision or

retinal pigmentation¹⁸. If this adverse effect occurs, retigabine should be discontinued¹⁸.

Calcium channels

Calcium is essential for cell membrane excitability, because it is a cation that depolarizes the membrane, allowing neurotransmitters to be released from synaptic vesicles to the synapse. Voltage-gated calcium channels (CaV or VGCC) are transmembrane proteins that act to regulate cellular functions related to calcium. Depending on the sensitivity to membrane depolarization, two CaVs have been described: High voltage-activated (HVA) and low voltage-activated (LVA) calcium channels¹⁹. HVA channels are heteromultimeric proteins that consist of a Cav α 1 subunit, plus ancillary Cav β , and Cav α 2 δ subunits in a 1:1:1 ratio. LVA channels may consist of only the Cav α 1 subunit. The Cav α 1 subunit is the essential component of the pore channel, and ten variants have been identified and divided into three subfamilies according to their pharmacological properties and pore-opening kinetic: CaV1 and CaV2 for HVA and CaV3 for LVA. CaV1 contains four members, all of them giving rise to L-type currents. CaV2 contains three members, with P/Q-, N-, and R-type currents. Finally, CaV3 has three members, all producing type T currents²⁰. Cav α 1 subunits all have similar structures, separated into four domains (I-IV) each of which is composed of S1-S6²¹.

Recently, 12 mutations in the *CACNA1A* gene, which codes for the transmembrane pore-forming subunit of CaV2.1, have been reported in 10 unrelated cases of epilepsy. The results showed that *CACNA1A* mutations were associated with pure epilepsy and the spectrum of epileptic phenotypes. In the same way, the Leu226Trp *CACNA1A* variant was associated with juvenile myoclonic epilepsy (JME), in three members of an Iranian family affected by IGE²².

Pregabalin and gabapentin block HVA calcium channels through the Cav2 subunit, inhibiting neurotransmitter release at the synapse and lowering neuronal excitability. Ethosuximide (ESM), which inhibits calcium (ICa⁺) currents in thalamic neurons through the T-type channel, is the medication of choice in the typical absence seizures²³. At present, Ca²⁺ blockers are used to control epileptic seizures, within the ASMs that have their mechanism of action blocking

presynaptic Ca²⁺ channels. In the L- and N-type calcium channel blockers group, we can find topiramate, lamotrigine, zonisamide, and valproate. On the other hand, drugs such as gabapentin and lamotrigine are known to be H current modulators. It is suggested that PHT inhibits Ca²⁺ flux. Among its most important adverse effects are blood dyscrasias, such as aplastic anemia, Stevens-Johnson syndrome, fever, mouth and throat pain, fatigue, and burning eyes²³.

Chloride channels-GABA receptors

Chloride is an essential anion for maintaining electro-negativity in neurons. Cl⁻'s principal hyperpolarizing effects on neuronal transmission are mediated through the GABA_A receptor (GABA_AR). The most prevalent inhibitor neurotransmitter in the CNS is GABA. GABA can work through ionotropic (GABA_AR) and metabotropic (GABA_BR) receptors. The first of these has been one of the most crucial therapeutic targets in ASMs²⁴.

GABA_AR are heteropentameric structures made up of five subunits. A total of 19 GABA subunits have been described belonging to different families: α (1–6), β (1–3), γ (1–3), δ , ϵ , π , θ , and ρ (1–3). The receptor consists of two α subunits, two β subunits, and one γ subunit, arranged around a central pore permeable to Cl⁻. Each subunit shares a common structure and is composed of four TMD (TMD: TM1–TM4)²⁵. It has been proposed that the TM2 domain forms the channel pore, while the intracellular domain between TM3 and TM4 is the site that modulates receptor activity. Some agents act on GABA_AR, an important receptor targeted by multiple ASMs. Benzodiazepines (BDZ), barbiturates, and loreclezole, actions on the GABA_AR are the main or only known mechanism of anticonvulsant action. For topiramate, felbamate, retigabine, losigamone, and stiripentol, GABA_AR modulation is one of several potential anticonvulsant mechanisms²⁶. Barbiturates, such as phenobarbital, and BDZ, such as diazepam, are two types of medications commonly used as ASMs. Barbiturates, on the one hand, prolong the opening of the chloride channel in the GABA_AR, whereas BDZs enhance the frequency with which it opens. Mutations in the GABA_AR subunit are related to several types of idiopathic epilepsy, associated with FS²⁷. Other mutations identified in patients with epilepsy are those found in the genes encoding the α 1, α 6, β 2, β 3, γ 2, or δ subunits of GABA_ARs. Many of

these mutations affect the normal traffic of the receptor generating partially or completely impairing their expression on the synaptic plasma membrane²⁸.

In addition, GABA function has been aided by the development of new antiepileptic medicines. Tiagabine was created to inhibit presynaptic GABA uptake by neurons and glial cells. Vigabatrin is a non-competitive GABA transaminase inhibitor that enhances GABA's impact on synapses²⁹.

Glutamate receptors

Glutamate is the primary excitatory amino acid in the CNS, and it is produced in neurons from L-glutamine. Glutamate affects neurons through ionotropic and metabotropic receptors. N-methyl-D-aspartate (NDMA), α -amino-3-hydroxy-5-methyl-4-isoxazole propionic acid (AMPA), and kainic acid (kainate) receptors are examples of ionotropic receptors³⁰. Metabotropic receptors are made up of eight receptors (mGluR1-mGluR8) classified into three classes. Group I receptors (mGluR1 and mGluR5) are connected to Gq proteins at both the presynaptic and postsynaptic levels. In Groups II and III, receptors are primarily bound to Gi proteins at the presynaptic level³¹.

It has recently been described that those mutations of *GRIN1*, *GRIN2A*, *GRIN2B*, and *GRIN2D* genes, which encode the GluN1, GluN2A, GluN2B, and GluN2D subunits, respectively, have all been associated with different epileptic phenotypes, such as EE, Rolandic epilepsy, Landau-Kleffner syndrome, and pike-and-waves during slow-wave sleep syndrome. The most frequent mutations are located in the *GRIN2A* gene and are present in the most severe phenotypes of epilepsy³². Perampanel is a selective AMPA-receptor antagonist. On the other hand, felbamate is an NMDA-receptor antagonist, which reduces excitatory neurotransmission by preventing glutamate binding³³.

Valproic acid (VPA) has numerous targets in its mode of action; it can improve GABAergic activities, inhibit the excitatory action of glutamate in the NDMA receptor, block voltage-dependent sodium currents, and has additional anti-epileptogenic properties that are not fully understood³⁴. Topiramate, a sulfamate-substituted monosaccharide with comparable mechanisms of action, has a beneficial antiepileptic effect

by raising the seizure threshold³⁵. Other molecular targets, such as levetiracetam and brivaracetam, are synaptic vesicle protein 2A (SV2A) modulator ligands that can limit neurotransmitter release³⁶. Acetazolamide and other carbonic anhydrase inhibitors have also been authorized to treat epilepsy.

NEUROTOXICITY OF ANTI-SEIZURE MEDICATIONS

ASMs are the first-line treatment for practically every kind of epilepsy; nonetheless, these medications are not devoid of producing neurotoxicity or inducing epileptic episodes. While an ASMs may be the gold standard of therapy for one kind of epilepsy, it might exacerbate other types of epilepsy if overused. Wrong diagnoses, pharmacological tolerance, inverse pharmacodynamic effects, incorrect dosage by overdosing, and irregular medication consumption are all common reasons for this paradoxical result.

The medications of choice for childhood absence epilepsy include ESM and VPA, whereas carbamazepine, PHT, phenobarbital (at high dosages), and tiagabine might aggravate seizures²³. In Lennox-Gastaut syndrome, the recommended medication is VPA, rufinamide, lamotrigine, or topiramate; however, gabapentin or BDZ aggravate seizures. JME can be treated with VPA, levetiracetam, and lamotrigine. However, it is exacerbated with carbamazepine (CBZ) and PHT³⁷. PHT can also worsen myoclonus in symptomatic epilepsies. Infantile spasms traditionally treated with VPA and vigabatrin (or adrenocorticotropin in the setting of West syndrome) may be exacerbated by CBZ and BDZ³⁸. The VPA is the ASM with the most therapeutic indications in epilepsy, due to the broad range of its mechanism of action; nevertheless, this suggests that enough side effects have emerged. Dose-dependent side effects include gastrointestinal abnormalities such as anorexia, nausea, and vomiting, which usually subside at low doses, there may be increased appetite with subsequent weight gain; the phenomenon of hypersensitivity is occasionally expressed by a skin rash; and up to 50% of patients may notice hair thinning and fragility, with alopecia occurring in some cases³⁹. Menstrual irregularities and polycystic ovarian syndrome have been linked to VPA intake, according to case studies⁴⁰. In terms of neurotoxicity, it should be noted that the appearance of distal fine

tremor, while not significant enough to withdraw the drug. In addition, it should be noted that hyperammonemia encephalopathy may occur in severe cases, so VPA should not be administered in patients with liver disease or mitochondrial diseases⁴¹. Pseudoatrophy of the brain has been observed in several cases.

PHT is an antiarrhythmic medication. Chronic usage causes gingival hyperplasia, hirsutism, coarse facial features, hyperpigmentation, and acne, along with changes in bone metabolism and folate deficiencies at the systemic level⁴². Serum levels > 30 µg/mL are associated with vestibular system dysfunction, which manifests as nystagmus, ataxia, dysarthria, and, lastly, consciousness level abnormalities. Personality changes, peripheral neuropathy, and extrapyramidal symptoms have also been observed on a rarer basis. Chronic PHT treatment can result in permanent cerebellar atrophy⁴³.

CBZ is a medication that has a similar structure to TCA. Despite being a highly effective treatment for focal onset seizures, it should be used with extreme caution in generalized seizures due to the paradoxical consequences. Most of its toxic effects are manifested at the CNS level and include somnolence, tiredness, dizziness, diplopia, ataxia, and asterixis. Other systemic side effects include secondary hyponatremia, SIADH, agranulocytosis, leukopenia, and decreased cardiac conduction⁴⁴.

Phenobarbital (PHB) is a barbiturate having hypnotic and sedative characteristics. Its neurotoxic effects are dose-dependent and quite like those seen with other ASMs, including ataxia, tiredness, drowsiness, behavioral abnormalities, mood disorders (depression), libido issues, and sexual impotence. The above is traditionally related to Dupuytren contracture, along with folate, cobalamin, and thyroxine insufficiency⁴⁵.

BDZ (diazepam, clonazepam, clobazam, midazolam, and lorazepam) has different pharmacological interactions based on their half-life. However, they share neurological dose-dependent side effects that can be mild, such as fatigue, dizziness, somnolence, ataxia, and irritability. At higher doses, it can cause life-threatening situations such as arterial hypotension and respiratory depression⁴⁶.

Most of the other ASMs have the same neurological side effects as the medications listed above.

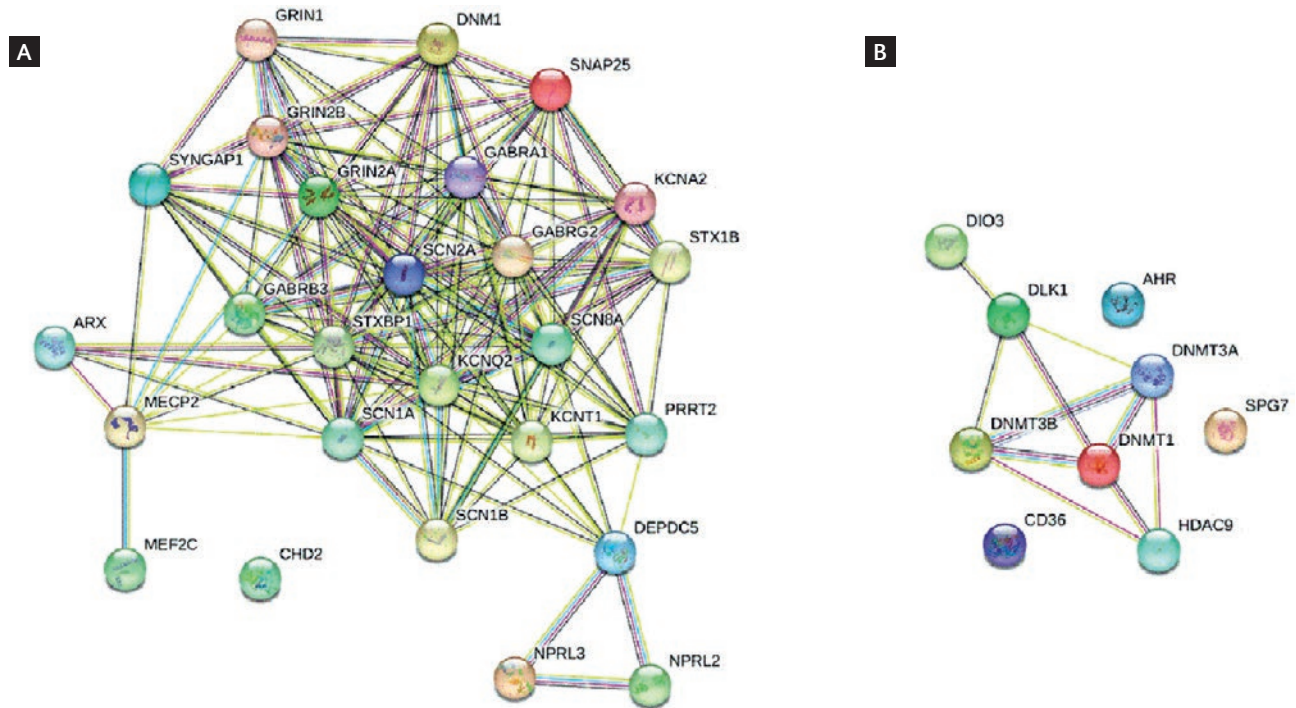
Nonetheless, many significant side effects should be noted. Vigabatrin (VGB) promotes GABAergic cell malfunction in the retina cone cells, resulting in visual field changes and irreversible blindness. Suicidal thoughts may also be increased with vigabatrin⁴⁷. When it comes to topiramate (TPM), almost half of the individuals using this treatment have paresthesia and weight loss. Verbal language impairments, on the other hand, are rather prevalent. Psychotic symptoms have been reported in individuals with a psychiatric history at high doses of TPM, perhaps due to the drug's suppression of the prefrontal cortex⁴⁸. Although no therapy is completely unassailable for the body, the tolerance to new antiepileptics has increased the prescription for epilepsy treatments. Levetiracetam is one of these treatments, with modest and potentially reversible neurotoxic effects and reduced pharmacological interaction⁴⁹. Under the right treatment regimens, both lamotrigine and LCM are typically well-tolerated.

EPIGENETIC TOXICITY DUE TO ANTI-SEIZURE MEDICATIONS

As mentioned above, ASMs are directed to different molecular targets to exert their effects as regulators of voltage-dependent ion channels or synaptic excitability. The above implies that these functions also consequently modify gene expression, functioning as epigenetic modifiers. Epigenetics refers to all those gene-dependent changes that are heritable but lack alterations in deoxyribonucleic acid (DNA) sequencing. The point of association between drugs and their gene effects is found in epigenetic markers (Fig. 2A and B). The modification of histone proteins, DNA methylation, and untranslated RNA functions are key markers⁵⁰.

Histone modification is primarily involved in transcription manipulation through acetylation. This process is regulated by groups of specific enzymes that cause metabolites to serve as gene activators or inhibitors, functioning as a balance between histone acetyltransferases and histone deacetylases. The latter is classified into four classes, and it has been shown that drugs such as VPA, CBZ, TPM, and LCM can inhibit them, mainly class I. An inhibition of this enzyme group provokes the accumulation of acetylated protein forms, regulating in turn cell proliferation and cell death⁵¹.

Figure 2. Interaction networks in epilepsy. Nodes represent proteins and edges represent protein-protein associations (i.e., proteins jointly contribute to a shared function). Filled nodes represent some 3D structure is known or predicted. **(A)** The most significant genes implicated in epilepsy. The genes mainly involved encode subunits of ion channels: sodium (*SCN1A*, *SCN2A*, *SCN1B*, and *SCN8A*), potassium (*KCNQ2*, *KCNA2*, and *KCNT1*), and GABA-chloride (*GABRA1*, *GABRG2*, and *GABRB3*). *SCN2A* stands out for its multiple associations with almost all the genes. Analysis: average node degree: 12.5, average local clustering coefficient: 0.792. **(B)** The most significant genes altered with antiepileptic treatment. It can be seen that the three genes with greater importance in these associations are *DNMT3A*, *DNMT3B*, and *DNMT1*. These genes encode DNA (cytosine-5)-methyltransferases. *DNMT3A* and *DNMT3B* methylate genome-wide *de novo* and establish methylation patterns during development. *DNMT1* methylates hemimethylated DNA. Analysis: average node degree: 2.22, average local clustering coefficient: 0.556 (Retrieved from: <https://string-db.org/cgi/network.pl>).



The DNA methylation process – the second epigenetic biomarker – is interfered with by *de novo* or maintenance methyltransferases (DNMTs), which are responsible for guiding a stable, inherited gene regulation essential in epigenetic regulation. These enzymes add a methyl group to DNA during or after DNA replication. Three groups of DNA methyltransferases (DNMT1, DNMT3a, and DNMT3b) have been identified in mammals. Drug exposure affects methylation, inducing a state of hypermethylation or hypomethylation in the promoter regions of genes. VPA is a known inhibitor of all three enzymatic forms of DNMTs, which influences neuronal migration and brain development⁵². Other drugs such as ESM have a potentiating effect by elevating DNMT1 and DNMT3a expression when administered chronically³⁰.

Untranslated RNA lacks protein-coding potential on its own, but its incidence when expressing other genes

is key in this type of epigenetic biomarker³¹. PHB has also been associated with alterations by this mechanism, specifically in abnormal expression at delta-like homolog 1 gene (*Dlk1*) and the iodothyronine deiodinase type III gene (*Dio3*) levels. The expressivity modifications in these genes are related to hepatocytes' hypertrophy with oncogenic capacity⁵³. The intervention of ASMs at the three epigenetic mechanisms discussed above has negative health consequences, which the researchers call "epigenetic toxicity." When we talk about genetic alterations, we immediately think of oncogenes. In this sense, ASM has been associated with carcinogenic promoters, for example, it has been seen that PHB by altering the expression of non-coding RNA by activating carcinogenic induces the transformation of hepatocytes into neoplastic cells⁵⁴.

ASMs also have neurotoxic potential, as they have been linked to alterations in genes key to neuronal

development in the fetal stage. The genes primarily affected are those that code for placental transporters and folate metabolism. Of all the drugs, the one with the highest teratogenic risk is VPA since studies have shown that its involvement in acetylation and histone methylation affects the neuroepithelium in rats. Mice with administration schedules of this drug have shown elevated levels of epigenetic markers recognized in this pathology such as miR-132 and miR-134-5p in their nerve tissue. Furthermore, autism spectrum disorder has even been associated with the use of VPA⁵⁵. An inflammatory effect of genetic origin is another of the mechanisms of toxicity that has been found in epilepsy treatments. VPA decreases the methylation levels of the PPAR γ , PPAR α , CD36, and AHR genes, which causes an inhibition of beta-oxidation and an increase in the hepatic entry of fatty acids. The result will be an inflammatory chain that starts from the consequent hepatic steatosis. VPA is also a known drug-causing agent of pancreatitis, interfering with the proliferation of acinar cells and reprogramming their embryological configuration by inhibiting HDAC⁵⁶. Another drug associated with activation of the inflammation cascade is CBZ, which can activate CD4+ T lymphocytes by epigenetic factors such as miR-18a and miR-155. This lymphocyte activation will favor immune hypersensitivity processes such as Stevens-Johnson syndrome⁵⁷.

DRUG-INDUCED EPILEPTIC SEIZURES

The fact that certain drugs used for different purposes can trigger epileptic seizures has allowed to deepen the research both of epileptogenesis and adverse effects of ASMs, since some of these drugs are used as experimental models of epilepsy⁵⁸. Penicillin administration is one of the chemical models of epilepsy most used in experiments⁵⁹. The drugs that can cause epileptic seizures and their respective mechanisms of ictogenesis are described, which allows us to have more caution when selecting a drug for a patient with comorbidities related to epilepsy.

Antibiotics

Antibiotics, especially beta-lactams (penicillin, cephalosporins, monobactams, and carbapenems), are a class of medications that have been extensively

explored for these effects. The GABA $_A$ R is the most prevalent toxicity target for these medicines (Fig. 3). Penicillin, cephalosporins, imipenem, and fluoroquinolones, among others, may function as direct antagonists of the GABA $_A$ R in the CNS in susceptible individuals. Isoniazid, on the other hand, an antitubercular drug, has two identified mechanisms: competitive inhibition of pyridoxine kinase and blockage of glutamate decarboxylase, in which both actions impede GABA production⁶⁰.

Macrolides may cause seizures, according to limited research. Clarithromycin, which is routinely used to treat upper respiratory tract infections, has been examined as a stimulant, specifically for the treatment of hypersomnia. Clarithromycin-induced reversible cellular hyperexcitability was reported by Bichler et al., and it is suggested that this action is mediated through the GABA receptor⁶¹. There have been reports of epileptic seizures induced by the therapeutic administration of metronidazole, polymyxin B, and colistin, but no convincing evidence on the action mechanism is available⁶².

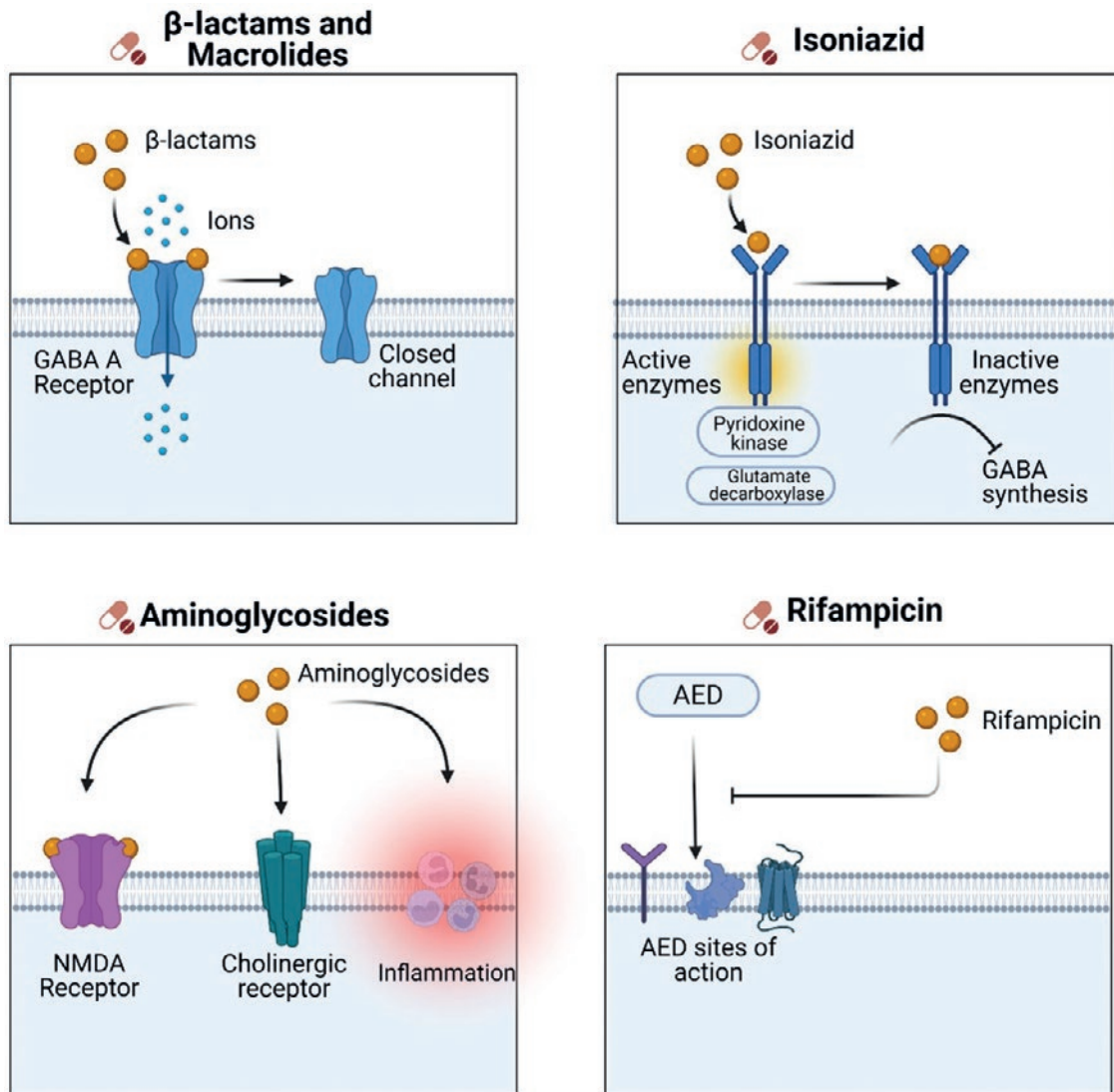
Increased permeability of the blood-brain barrier in CNS disorders and prematurity, as well as impaired drug metabolism in kidney and liver failure are risk factors for developing drug-induced seizures⁶⁰. Another notable neurotoxic impact of antibiotics is that of aminoglycosides, which produce ototoxicity, peripheral neuropathy, and possibly encephalopathy by activating NMDA receptors, interacting with cholinergic receptors, and causing an inflammatory response in the axon. Gentamicin is the primary cause of adverse symptoms⁶³.

Furthermore, we must examine the pharmacological interaction under polypharmacy situations. For example, rifampicin can lower blood levels of CBZ, VPA, ESM, and PHT. On the other hand, trimethoprim-sulfamethoxazole enhances PHT toxicity⁶⁰.

Stimulants and sympathomimetics

For several years, the role of noradrenaline in epileptic seizures has been demonstrated. Experimental models of epilepsy based on ligands of adrenergic receptors have been developed; therefore, drugs with adrenergic agonist action could be expected to cause epileptic seizures as side effects, as with

Figure 3. Drug-induced epileptic seizures. Beta-lactams and macrolides have effects on suppressing the activity of GABA_A receptor. Isoniazid has two identified mechanisms: Competitive inhibition of pyridoxine kinase and blockage of glutamate decarboxylase, which both actions impede GABA production. Aminoglycosides may interact with NMDA receptors, cholinergic receptors, and may cause an inflammatory response in the axon. And lastly, rifampicin can lower blood levels of carbamazepine, valproic acid, ethosuximide, and phenytoin.



phenylephrine and pseudoephedrine. A class of powerful CNS stimulant medications known as amphetamines is synthetic adrenergic agents. Methylphenidate, an amphetamine structural analog, is a medication licensed for the treatment of attention deficit hyperactivity disorder. It works by boosting dopamine and noradrenaline levels in the CSN by inhibiting monoamine carrier recapture, resulting in dangerous levels capable of inducing epileptic seizures⁶⁴.

Certain psychopharmaceuticals can cause epileptic seizures through a similar mechanism, including monoamine oxidase inhibitors (isocarboxazid, phenelzine, and selegiline), selective inhibitors of serotonin reuptake (fluoxetine, sertraline, and paroxetine), serotonin-norepinephrine reuptake inhibitors (atomoxetine and desvenlafaxine), and tricyclic antidepressants (amitriptyline, imipramine, and nortriptyline). Bupropion is a dopamine and noradrenaline reuptake inhibitor intended to help people quit smoking,

according to Yan⁶⁵. Despite being well-tolerated in his early clinical trials, it has subsequently been linked to seizures following an accidental or purposeful overdose. This dose-dependent impact is directly tied to the history of epilepsy. Nevertheless, the mechanism of action by which seizures occur has not yet been properly defined.

Other drugs associated with seizures

However, research on the capacity of non-steroidal anti-inflammatory drugs and opioids to cause epileptic seizures is yet inconclusive. Both acetaminophen and ibuprofen appear to be medicines that, at therapeutic levels, would not lower the seizure threshold. Due to adenosine A1-receptor antagonism, methylxanthines (theophylline and aminophylline) can produce seizures⁷. Antidiabetic medications, especially those with a strong hypoglycemia impact, might cause seizures. Seizures develop at blood glucose levels of 16–19 mg/100 mL according to experimental research. Furthermore, it has been reported, that there are spontaneous sporadic occurrences of epileptic seizures following the administration of anti-cancer medicines and immunosuppressants⁶⁶.

CONCLUSION

As we have discussed, the evidence of the adverse effects prevalence due to antiepileptic drugs administration is significant. These findings are not surprising, since each individual response may vary depending on the molecular mechanisms involved with the epilepsy type, as well as the drug used. In this regard, epigenetic factors have a relevant function in the regulation of mechanisms involved in ASMs processing. Many ASMs generate epigenetic toxicity that can lead to gene dysregulation and thus to the possible cancer development by promoting neoplastic cells generation. In addition, the presence of genetic variants in the different ionic channels is relevant to defining the prognosis and type of treatment used. It is not surprising that the response to a drug can vary between individuals, who present the same phenotype. Hence, the importance of creating new strategies that mitigate the side effects of ASMs, considering the individual aspects of each patient and their comorbidities to develop personalized therapies.

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REFERENCES

1. Thijs RD, Surges R, O'Brien TJ, Sander JW. Epilepsy in adults. *Lancet*. 2019;393:689-701.
2. Fisher RS, Cross JH, French JA, Higurashi N, Hirsch E, Jansen FE, et al. Operational classification of seizure types by the international league against epilepsy: position paper of the ILAE commission for classification and terminology. *Epilepsia*. 2017; 58:522-30.
3. French JA, Perucca E. Time to start calling things by their own names? The case for antiseizure medicines. *Epilepsy Curr*. 2020; 20:69-72.
4. Dryżałowski P, Józwiak S, Franckiewicz M, Strzelecka J. Benign epilepsy with centrotemporal spikes-Current concepts of diagnosis and treatment. *Neurol Neurochir Pol*. 2018;52:677-89.
5. Löscher W, Potschka H, Sisodiya SM, Vezzani A. Drug resistance in epilepsy: clinical impact, potential mechanisms, and new innovative treatment options. *Pharmacol Rev*. 2020;72:606-38.
6. Hitchings AW. Drugs that lower the seizure threshold. *Adverse Drug React Bull*. 2016;298:1151-4.
7. Steinlein OK. Ion channels and epilepsy. *Am J Med Genet Semin Med Genet*. 2001;106:146-59.
8. Du J, Simmons S, Brunklaus A, Adiconis X, Hession CC, Fu Z, et al. Differential excitatory vs inhibitory SCN expression at single cell level regulates brain sodium channel function in neurodevelopmental disorders. *Eur J Paediatr Neurol*. 2020;24:129-33.
9. Escayg A, MacDonald BT, Meisler MH, Baulac S, Huberfeld G, An-Gourfinkel I, et al. Mutations of SCN1A, encoding a neuronal sodium channel, in two families with GEFS+2. *Nat Genet*. 2000; 24:343-5.
10. Goldin AL. Nomenclature of voltage-gated sodium channels. *Neuron*. 2001;28:365-8.
11. Yu FH, Catterall WA. Overview of the voltage-gated sodium channel family. *Genome Biol*. 2003;4:207.
12. Hebeisen S, Pires N, Loureiro AI, Jo M, Whyment A, Palma N, et al. Neuropharmacology Eslicarbazepine and the enhancement of slow inactivation of voltage-gated sodium channels : a comparison with carbamazepine, oxcarbazepine and lacosamide. *Neuropharmacol J*. 2015;89:122-35.
13. Panebianco M, Prabhakar H, Marson AG. Rufinamide add-on therapy for drug-resistant epilepsy. *Cochrane Database Syst Rev*. 2020;11:CD011772.
14. Trimmer JS. Subcellular localization of K⁺ channels in mammalian brain neurons: remarkable precision in the midst of extraordinary complexity. *Neuron*. 2016;85:238-56.
15. Villa C, Combi R. Potassium channels and human epileptic phenotypes : an updated overview. *Front Cell Neurosci*. 2016; 10:1-14.
16. Baculis BC, Zhang J, Chung HJ. The role of Kv7 channels in neural plasticity and behavior. *Front Physiol*. 2020;11:568667.
17. Wulff H, Castle NA, Pardo LA. Voltage-gated potassium channels as therapeutic drug targets. *Nat Rev Drug Discov*. 2010; 8:982-1001.
18. Czuczwar P, Wojtak A, Cioczek-czuczwar A, Parada-turska J, Maciejewski R, Czuczwar SJ. Retigabine : the newer potential antiepileptic drug. *Pharmacol Rep*. 2010;62:211-9.
19. Fedulova SA, Kostyuk PG, Veselovsky NS. Two types of calcium channels in the somatic membrane. *J Physiol*. 1985;359: 431-46.
20. Hering S, Zangerl-Plessl EM, Beyl S, Hohaus A, Andranovits S, Timin EN. Calcium channel gating. *Eur J Physiol*. 2018;470: 1291-309.
21. Dolphin AC. Functions of presynaptic voltage-gated calcium channels. *Function*. 2021;2:1-10.

22. Alehabib E, Kokotovic T, Ranji-Burachaloo S, Tafakhori A, Ramshe SM, Esmailizadeh Z, et al. Leu226Trp CACNA1A variant associated with juvenile myoclonic epilepsy with and without intellectual disability. *Clin Neurol Neurosurg.* 2022;213:107108.
23. Kilaru S, Emily K. A practical guide to treatment of childhood absence epilepsy. *Pediatr Drugs.* 2019;21:15-24.
24. Zheng W, Xie W, Zhang J, Strong JA, Wang L, Yu L, et al. Function of γ -aminobutyric acid receptor/channel 1 subunits in spinal cord. *J Biol Chem.* 2003;278:48321-9.
25. Brohan J, Goudra BG. The role of GABA receptor agonists in anesthesia and sedation. *CNS Drugs.* 2017;31:845-6.
26. Schipper S, Aalbers MW, Rijkers K, Swijssen A, Rigo JM. Tonic GABA A receptors as potential target for the treatment of temporal lobe epilepsy. *Mol Neurobiol.* 2016;53:5252-65.
27. Ghit A, Assal D, Al-Shami AS, Hussein DE. GABA A receptors : structure, function, pharmacology, and related disorders. *J Genet Eng Biotechnol.* 2021;19:123.
28. Mele M, Costa RO, Duarte CB. Alterations in GABA A-receptor trafficking and synaptic dysfunction in brain disorders. *Front Cell Neurosci.* 2019;13:77.
29. Wheless JW, Ramsay RE, Collins SD. Vigabatrin. *J Am Soc Exp Neurother.* 2007;4:163-72.
30. Hanada T. Ionotropic glutamate receptors in epilepsy : a review focusing on AMPA and NMDA receptors. *Biomolecules.* 2020;10:464.
31. Sengmany K, Gregory KJ. Metabotropic glutamate receptor subtype 5 : molecular pharmacology, allosteric modulation and stimulus bias. *Br J Pharmacol.* 2016;173:3001-17.
32. Perucca P, Perucca E. Identifying mutations in epilepsy genes : impact on treatment selection. *Epilepsy Res.* 2019;152:18-30.
33. Leppik IE, Wechsler RT, Williams B, Yang H, Zhou S, Laurenza A. Efficacy and safety of perampanel in the subgroup of elderly patients included in the phase III epilepsy clinical trials. *Epilepsy Res.* 2015;110:216-20.
34. Löscher W. Basic pharmacology of valproate a review after 35 years of clinical use for the treatment of epilepsy. *CNS Drugs.* 2002;16:669-94.
35. Khalil NY, Alrabiah HK, Rashoud SS, Bari A, Wani TA. Topiramate comprehensive profile. In: *Profiles of Drug Substances, Excipients, and Related Methodology.* 1st ed., Vol. 44. Netherlands: Elsevier Inc.; 2019. p. 333-78.
36. Tan J, Paquette V, Levine M, Ensom MH. Levetiracetam clinical pharmacokinetic monitoring in pediatric patients with epilepsy. *Clin Pharmacokinet.* 2017;56:1267-85.
37. Shnyder NA, Petrov KV. Juvenile myoclonic epilepsy: current state of the problem. *Pers Psychiatry Neurol.* 2021;1:2-20.
38. Moosa AN. Antiepileptic drug treatment of epilepsy in children. *Contin Lifelong Learn Neurol.* 2019;25:381-407.
39. Johannessen CU, Johannessen SI. Valproate: past, present, and future mechanisms of action. *CNS Drug Rev.* 2003;9:199-216.
40. Isoja JI, Tapanainen JS. Valproate, hyperandrogenism, and polycystic ovaries. *Dimens Contemp Ger Arts Lett.* 2000;57:1064-8.
41. Nanau RM, Neuman MG. Adverse drug reactions induced by valproic acid. *Clin Biochem.* 2013;46:1323-38.
42. Stinnett E, Rodu B, Grizzle WE. New developments in understanding phenytoin-induced gingival hyperplasia. *J Am Dent Assoc.* 1987;114:814-6.
43. Gallop K. Review article: phenytoin use and efficacy in the ED. *Emerg Med Australas.* 2010;22:108-18.
44. Fricke-Galindo I, Llerena A, Jung-Cook H, López-López M. Carbamazepine adverse drug reactions. *Expert Rev Clin Pharmacol.* 2018;11:705-18.
45. Si Y, Liu L, Tian L, Mu J, Chen D, Chen T, et al. A preliminary observation of the adverse effects of phenobarbital among patients with convulsive epilepsy in rural West China. *Epilepsy Behav.* 2016;54:65-70.
46. Geulayov G, Ferrey A, Casey D, Wells C, Fuller A, Bankhead C, et al. Relative toxicity of benzodiazepines and hypnotics commonly used for self-poisoning: an epidemiological study of fatal toxicity and case fatality. *J Psychopharmacol.* 2018;32:654-62.
47. Jastrzembski B, Locke J, Wan MJ. Clinical implications and cost of electroretinography screening for vigabatrin toxicity. *Can J Ophthalmol.* 2020;55:e98-100.
48. Besag FM, Vasey MJ. Neurocognitive effects of antiseizure medications in children and adolescents with epilepsy. *Pediatric Drugs.* 2021;23:253-86.
49. Morgan O, Medenwald B. Safety and tolerability of rapid administration undiluted levetiracetam. *Neurocrit Care.* 2020;32:131-4.
50. Kong FC, Ma CL, Zhong MK. Epigenetic effects mediated by antiepileptic drugs and their potential application. *Curr Neuropharmacol.* 2019;18:153-66.
51. Pierre JJ, Kantarjian HM. Targeting DNA methylation. *Mol Cell Biochem.* 2012;23:1-7.
52. Marx M, Billups D, Billups B. Maintaining the presynaptic glutamate supply for excitatory neurotransmission. *J Neurosci Res.* 2015;1044:1031-44.
53. Burnashev N, Szepietowski P. NMDA receptor subunit mutations in neurodevelopmental disorders. *Curr Opin Pharmacol.* 2014;20:73-82.
54. Holsapple MP, Pitot HC, Cohen SH, Boobis AR, Klaunig JE, Pastoor T, et al. Mode of action in relevance of rodent liver tumors to human cancer risk. *Toxicol Sci.* 2006;89:51-6.
55. Hirsch MM, Deckmann I, Fontes-Dutra M, Bauer-Negrini G, Della-Flora Nunes G, Nunes W, et al. Behavioral alterations in autism model induced by valproic acid and translational analysis of circulating microRNA. *Food Chem Toxicol.* 2018;115:336-43.
56. Eisses JF, Criscimanna A, Dionise ZR, Orabi AI, Javed TA, Sarwar S, et al. Valproic acid limits pancreatic recovery after pancreatitis by inhibiting histone deacetylases and preventing acinar re-differentiation programs. *Am J Pathol.* 2015;185:3304-15.
57. Monroy-Arreola A, Durán-Figueroa NV, Méndez-Flores S, Domínguez-Cherit J, Watkinson J, Badillo-Corona JA, et al. Up-Regulation of T-Cell activation MicroRNAs in drug-specific CD4 + T-cells from hypersensitive patients. *Chem Res Toxicol.* 2018;31:454-61.
58. Rubio C, Rubio-Osornio M, Retana-Márquez S, Verónica Custodio ML, Paz C. In vivo experimental models of epilepsy. *Cent Nerv Syst Agents Med Chem.* 2010;10:298-309.
59. Chen RC, Huang YH, How SW. Systemic penicillin as an experimental model of epilepsy. *Exp Neurol.* 1986;92:533-40.
60. Wanleenuwat P, Suntharampillai N, Iwanowski P. Antibiotic-induced epileptic seizures: mechanisms of action and clinical considerations. *Seizure.* 2020;81:167-74.
61. Bichler E, Elder C, García P. Clarithromycin increases neuronal excitability in CA3 pyramidal neurons through a reduction in GABAergic signaling. *J Neurophysiol.* 2017;117:93-103.
62. Nigam A, Kumari A, Jain R, Batra S. Colistin neurotoxicity: revisited. *BMJ Case Rep.* 2015;2015:bcr2015210787.
63. Grill MF, Maganti RK. Neurotoxic effects associated with antibiotic use: management considerations. *Br J Clin Pharmacol.* 2011;72:381-93.
64. Fitzgerald KT, Bronstein AC. Adderall® (Amphetamine-dextroamphetamine) toxicity. *Top Companion Anim Med.* 2013;28:2-7.
65. Yan T, Goldman RD. Bupropion for smoking cessation in adolescents. *Can Fam Physician.* 2021;67:743-5.
66. Rubio C, Rosiles-Abonce A, Taddei E, Rubio-Osornio M. Neurotoxicity and epileptogenesis. In: *Neurotox New Advances.* London: IntechOpen; 2021.