



COMPLETE SCREENING OF EXONS 2, 3, AND 4 OF *KRAS* AND *NRAS* GENES REVEALS A HIGHER NUMBER OF CLINICALLY RELEVANT MUTATIONS THAN FOOD AND DRUG ADMINISTRATION QUANTITATIVE POLYMERASE CHAIN REACTION-BASED COMMERCIAL KITS

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Table S1. Primer sequences and PCR product lengths

Primer name	Primer sequences	Genetic region	Fragment length
K2F	TGTGTGACATGTTCTAATATAGTCAC	KRAS Exon 2 (NC_000012.12)	216 bp
K2R	AGAATGGCCTGACCAGTAA		
K3F	ACTGTGTTCTCCCTCTCAG	KRAS Exon 3 (NC_000012.12)	282 bp
K3R	TGCATGGCATTAGCAAAGAC		
K4F	TCCCAGAGAACAAATTAAAAGAG	KRAS Exon 4 (NC_000012.12)	355 bp
K4R	AACCAAAGCCAAAGCAGTACC		
N2F	CTCGCCAATTAAACCTGATT	NRAS Exon 2 (NC_000001.11)	213 bp
N2R	CCGACAAGTGAGAGACAGGA		
N3F	ACACCCCCAGGATTCTTACAGA	NRAS Exon 3 (NC_000001.11)	126 bp
N3R	GACTTGCTATTATTGATGGC		
N4F	CCACTGTACCCAGCCTAATCTTG	NRAS Exon 4 (NC_000001.11)	257 bp
N4R	GCAAACCTTGCACAAATGCTGA		

PCR: polymerase chain reaction.

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ANALYSIS OF THE ENDOMETRIAL TRANSCRIPTOME AT THE TIME OF IMPLANTATION IN WOMEN RECEIVING A SINGLE POST-OVULATORY DOSE OF LEVONORGESTREL OR MIFEPRISTONE

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Table S1. Comparison of gene directionality of 60 common differentially expressed genes in endometrial biopsies between T-LNG and ERA

Gene symbol	Gene name	ERA (FC)	T-LNG (FC)
GPX3	Glutathione peroxidase 3 (plasma)	35.49	3.66
LIF	Leukemia inhibitory factor (cholinergic differentiation factor)	15.03	2.14
CXCL14	Chemokine (C-X-C motif) ligand	14.02	2.23
C4BPA	Complement component 4 binding protein, alpha	13.14	2.64
TSPAN8	Tetraspanin 8	12.90	1.55
GADD45A	Growth arrest and DNA damage-inducible, alpha	8.25	1.69
NNMT	Nicotinamide N-methyltransferase	7.74	1.62
DPP4	Dipeptidylpeptidase 4 (CD26, adenosine deaminase complexing protein 2)	7.72	2.57
S100P	S100 calcium-binding protein P	6.95	1.30
SNX10	Sorting nexin 10	6.56	1.93
CP	Ceruloplasmin (ferroxidase)	6.34	1.64
ADRA2A	Adrenergic, alpha-2A, receptor	5.78	2.50
TAGLN	Transgelin	5.71	1.61
THBS2	Thrombospondin 2	5.17	1.60
SERPING1	Serpin peptidase inhibitor, clade G (C1 inhibitor), member 1, (angioedema, hereditary), transcript variant 1	5.16	1.72
AOX1	Aldehyde oxidase 1	4.82	2.69
CYP3A5	Cytochrome P450, family 3, subfamily A, polypeptide 5	4.82	2.58
DHRS3	Dehydrogenase/reductase (SDR family) member 3	4.72	2.08
MYL9	Myosin, light polypeptide 9, regulatory, transcript variant 2	4.72	1.42
HABP2	Hyaluronan-binding protein 2	4.09	5.20
TBC1D2	TBC1 domain family, member 2	4.07	1.26
CDA	Homo sapiens cytidine deaminase (CDA), mRNA [NM_001785]	4.00	1.94
IMPA2	Inositol(myo)-1(or 4)-monophosphatase 2 3.91 2	3.91	1.78
XCL2	Chemokine (C motif) ligand 2	3.73	1.43
CLDN10	Claudin 10, transcript variant 1	3.68	2.06
COTL1	Coactosin-like 1 (Dictyostelium)	3.58	1.52
SYNE2	Spectrin repeat containing, nuclear envelope 2, transcript variant 5	3.52	1.88
PROS1	Homo sapiens protein S (alpha)	3.28	1.45
HLA-DOB	Major histocompatibility complex, class II, DO	-11.06	-2.35
CTNNA2	Catenin (cadherin-associated protein), alpha 2	-8.02	-2.00
NR4A2	Nuclear receptor subfamily 4, group A, member 2, transcript variant 1	-7.05	-2.00
DUOX1	Dual oxidase 1, transcript variant 1	-6.61	-3.05
LRP4	Low-density lipoprotein receptor-related protein 4	-5.87	-1.37
SLC16A6	Solute carrier family 16 (monocarboxylic acid transporters), member 6	-5.61	-2.14
EDN3	Endothelin 3, transcript variant 3	-5.30	-1.45
ANK3	Ankyrin 3, node of Ranvier (ankyrin G), transcript variant 1	-4.79	-2.73
CAPN6	Calpain 6 (CAPN6)	-4.76	-5.17
HPGD	Hydroxyprostaglandin dehydrogenase 15-(NAD)	-4.72	-2.00
SLC7A1	Solute carrier family 7 (cationic amino acid transporter, y β system), member 1	-4.72	-1.65
MMP26	Matrix metallopeptidase 26	-4.66	-5.90
LRRC17	Leucine-rich repeat-containing 17, transcript variant 2	-4.64	-1.67
ALPL	Alkaline phosphatase, liver/bone/kidney	-4.44	-2.14
HEY2	Hairy/enhancer-of-split related with YRPW motif 2	-4.07	-1.97
COBL	Cordon-bleu homolog (mouse)	-3.99	-1.79
SERPINA5	Serpin peptidase inhibitor, clade A (alpha-1 antiproteinase, antitrypsin), member 5	-3.84	-2.73
CREB3L1	cAMP responsive element-binding protein 3-like 1	-3.77	-1.55
IDH1	Isocitrate dehydrogenase 1 (NADP β), soluble	-3.68	-1.61
CBR3	Carbonyl reductase 3	-3.66	-1.72
ADAMTS8	A disintegrin-like and metalloprotease (reprolysin type) with thrombospondin type 1 motif, 8	-3.59	-2.48
BIRC3	Baculoviral IAP repeat-containing 3 (BIRC3), transcript variant 1	-3.58	-1.75
DFNB31	Deafness, autosomal recessive 31	-3.54	-2.19
KCNJ2	Potassium inwardly-rectifying channel, subfamily J, member 2	-3.47	-1.60
BARD1	BRCA1-associated RING domain 1	-3.47	-1.78
MAP2K6	Mitogen-activated protein kinase 6, transcript variant 1	-3.45	-2.58
NDRG2	NDRG family member 2, transcript variant 1	-3.40	-1.35
LRRC1	Leucine-rich repeat-containing 1	-3.23	-3.16
RASSF2	Ras association (RalGDS/AF-6) domain family 2, transcript variant 1	-3.19	-1.43
OPRK1	Opioid receptor, kappa 1	-3.17	-1.66
STEAP4	STEAP family member 4	-3.04	-1.79
HEY1	Hairy/enhancer-of-split related with YRPW motif 1	-3.03	-1.95

FC: fold change; ERA: endometrial receptivity array (22).

Negative values mean down-regulation.

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Table S2. Gene directionality of differentially expressed common genes in endometrial biopsies between T-MFP and ERA

Gene symbol	Gene name	ERA (FC)	T-MFP (FC)
PAEP	Progesterone-associated endometrial protein (placental protein 14, pregnancy-associated endometrial alpha-2-globulin, alpha uterine protein) (PAEP), transcript variant 2	31.43	-13.00
C4BPA	Complement component 4 binding protein, alpha	13.14	-5.43
MAOA	Monoamine oxidase A, nuclear gene encoding mitochondrial protein	9.39	-4.99
GADD45A	Growth arrest and DNA damage-inducible, alpha	8.25	-1.66
NNMT	Nicotinamide N-methyltransferase	7.74	-1.85
SCGB2A2	Secretoglobin, family 2A, member 2	7.43	-3.43
SNX10	Sorting nexin 10	6.56	-1.53
CP	Ceruloplasmin (ferroxidase)	6.34	-3.60
EFNA1	Ephrin-A1, transcript variant 1	5.77	-2.19
KLRC1	Killer cell lectin-like receptor subfamily C, member 1, transcript variant 2	5.75	-2.07
TAGLN	Transgelin	5.71	-1.64
SLC15A1	Solute carrier family 15 (oligopeptide transporter), member 1	5.59	-6.41
THBS2	Thrombospondin 2	5.17	-2.55
HPSE	Heparanase	5.17	-5.62
SERPING1	Serpin peptidase inhibitor, clade G (C1 inhibitor), member 1, (angioedema, hereditary), transcript variant 1	5.16	-2.38
CRISP3	Cysteine-rich secretory protein 3	5.09	-21.71
RPRM	Reprimo, TP53 dependent G2 arrest mediator candidate	5.03	-1.50
EDNRB	Endothelin receptor type B, transcript variant 2	4.89	-3.03
ABLIM3	Actin-binding LIM protein family, member 3	4.87	-1.61
GLIPR1	GLI pathogenesis-related 1 (glioma)	4.83	-2.00
CTSW	Cathepsin W	4.78	-2.03
AQP3	Aquaporin 3 (Gill blood group)	4.63	-1.68
S100A4	S100 calcium-binding protein A4, transcript variant 1	4.59	-1.30
NKG7	Natural killer cell group 7 sequence	4.42	-1.66
ARG2	Arginase, type II, nuclear gene encoding mitochondrial protein	4.35	-7.95
ANXA4	Annexin A4	4.27	-2.62
ARID5B	AT-rich interactive domain 5B (MRF1-like)	4.19	-1.56
SPP1	Secreted phosphoprotein 1 (osteopontin, bone sialoprotein I, early T-lymphocyte activation 1)	4.15	-11.00
DKK1	Dickkopf homolog 1 (<i>Xenopus laevis</i>)	4.10	-8.22
HABP2	Hyaluronan-binding protein 2	4.09	-1.72
GZMA	Granzyme A (granzyme 1, cytotoxic T-lymphocyte-associated serine esterase 3)	4.07	-2.75
C10orf10	Chromosome 10 open reading frame 10	3.98	-6.68
STAR	Steroidogenic acute regulator, nuclear gene encoding mitochondrial protein, transcript variant 1	3.93	-1.83
LMOD1	Leiomodin 1 (smooth muscle)	3.92	-2.01
HAND2	Heart and neural crest derivatives expressed 2	3.91	-2.06
CXCL13	Chemokine (C-X-C motif) ligand 13 (B-cell chemoattractant)	3.90	-9.19
RARRES3	Retinoic acid receptor responder (tazarotene induced) 3	3.79	-1.49
MT2A	Metallothionein 2A	3.72	-3.16
RRAS	Related RAS viral (r-ras) oncogene homolog	3.70	-1.80
COTL1	Coactosin-like 1 (Dictyostelium)	3.58	-1.42
ID4	Inhibitor of DNA-binding 4, dominant negative helix-loop-helix protein	3.56	-2.33
LMCD1	LIM and cysteine-rich domains 1	3.53	-1.61
MFAP5	Microfibrillar-associated protein 5	3.42	-3.86
MT1G	Metallothionein 1G	3.41	-71.01
CLDN4	Claudin 4	3.41	-2.11
PTPRR	Protein tyrosine phosphatase, receptor type, R, transcript variant 1	3.40	-2.04
DDX52	DEAD (Asp-Glu-Ala-Asp) box polypeptide 52, transcript variant 2	3.38	-2.43
HAL	Histidine ammonia-lyase	3.37	-2.01
ADAMTS1	A disintegrin-like and metalloprotease (reprolysin type) with thrombospondin type 1 motif, 1	3.32	-4.86
PROS1	Homo sapiens protein S (alpha)	3.28	-1.40
FOSL2	FOS-like antigen 2	3.26	-2.27
AMIGO2	Adhesion molecule with Ig-like domain 2	3.20	-1.85
POLD4	Polymerase (DNA-directed), delta 4	3.16	-1.38
GBP2	Guanylate-binding protein 2, interferon-inducible	3.14	-4.08
GABARAPL1	GABA(A) receptor-associated protein-like 1	3.13	-1.89
MT1H	Metallothionein 1H	3.12	-48.84
ATP6V1A	ATPase, H ₀ transporting, lysosomal 70 kDa, V1 subunit A	3.04	-1.49
IL15	Interleukin 15, transcript variant 1	3.03	-4.26
BCL6	B-cell CLL/lymphoma 6 (zinc finger protein 51), transcript variant 2	3.02	-2.27
TRH	Thyrotropin-releasing hormone	-21.69	7.31
HLA-DOB	Major histocompatibility complex, class II, DO	-11.06	-2.55
CSRP2	Cysteine and glycine-rich protein 2	-9.62	1.80
SLC15A2	Solute carrier family 15 (H ₀ /peptide transporter), member 2	-8.44	2.62
SFRP4	Secreted frizzled-related protein 4	-8.33	3.76
NR4A2	Nuclear receptor subfamily 4, group A, member 2, transcript variant 1	-7.05	-2.17
DUOX1	Dual oxidase 1, transcript variant 1	-6.61	-3.51
KIF20A	Kinesin family member 20A	-6.33	2.11
PENK	Proenkephalin	-6.16	1.68
EDN3	Endothelin 3, transcript variant 3	-5.30	1.78
GALNT12	UDP-N-acetyl-alpha-D-galactosamine:polypeptide N-acetylgalactosaminyltransferase 12 (GalNAc-T12)	-4.85	3.23
CAPN6	Calpain 6 (CAPN6)	-4.76	-2.53
MMP26	Matrix metallopeptidase 26	-4.66	-8.40
LRRC17	Leucine-rich repeat-containing 17, transcript variant 2	-4.64	1.66
KCNG1	Potassium voltage-gated channel, subfamily G, member 1, transcript variant 1	-4.63	1.48
SOX17	SRY (sex determining region Y)-box 17	-4.24	1.81
TOP2A	Topoisomerase (DNA) II alpha 170 kDa	-3.98	2.51
SPDEF	SAM pointed domain-containing ets transcription factor	-3.78	5.54
KMO	Kynurenine 3-monooxygenase (kynurenine 3-hydroxylase)	-3.78	3.14
ATP1B1	ATPase, Na ₊ /K ₊ transporting, beta 1 polypeptide	-3.73	1.84
ADAMTS8	A disintegrin-like and metalloprotease (reprolysin type) with thrombospondin type 1 motif, 8	-3.59	-3.78
BIRC3	Baculoviral IAP repeat-containing 3 (BIRC3), transcript variant 1	-3.58	-2.50
PLA1A	Phospholipase A1 member A	-3.42	1.61
PBK	PDZ-binding kinase	-3.42	2.08
PRC1	Protein regulator of cytokinesis 1 (PRC1), transcript variant 1	-3.29	1.65
MSX1	Msh homeobox homolog 1 (<i>Drosophila</i>)	-3.26	1.93
LRRC1	Leucine-rich repeat-containing 1	-3.23	-3.60
EPHB3	EPH receptor B3	-3.20	1.52

FC: fold Change; ERA: endometrial receptivity array (22). Negative values mean down-regulation.



PROGNOSTIC SIGNIFICANCE OF THE MAD1L1 1673 G:A POLYMORPHISM IN OVARIAN ADENOCARCINOMAS

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Table S1. Clinical and histopathological variables of patients with ovarian adenocarcinoma treated at the National Cancer Institute – Mexico from 2005 to 2018 (n = 118)

Variable	Mean	SD
Age, years	52.12	11.78
BMI, kg/m ²	27.18	5.27
Variable	Median	Q1-Q3
Initial CA 125, U/mL	935.50	159.50-2770.20
Initial HE4, U/mL	50.40	36.23-82.58
Tumor Size, cm	11.50	5.87-18.00
Variable	n	%
BMI, kg/m²		
Underweight (<18.5)	3	2.54
Normal weight (18.5-24.9)	41	34.75
Overweight (25-29.9)	45	38.14
Obesity (≥30)	29	24.58
Initial ECOG performance status		
0	89	75.42
1	21	17.80
2	7	5.93
3	1	0.85
Histological type		
High-grade papillary serous	58	49.15
Low-grade papillary serous	7	5.93
Endometrioid	21	17.80
Clear cell	11	9.32
Mixed histology/other	21	17.80
FIGO clinical stage		
I	25	21.19
II	5	4.24
III	57	48.31
IV	31	26.27
Cytoreduction		
Taken to cytoreduction	113	95.76
Optimal cytoreduction	85	75.22*
Chemotherapy		
Received neoadjuvancy	74	62.71
Received adjuvancy	90	76.27
Recurrence/progression of disease		
Neither	40	33.90
Recurrence or progression	78	66.10
Sensitivity to chemotherapy		
Non-platin sensitive	58	49.15
Platin sensitive/no recurrence	60	50.85
Site of recurrence		
Peritoneal	16	20.51**
Nodal	23	29.48**
Distance	27	34.61**
Undetermined/other	12	15.38**
MAD1L1 genotype		
Wild type	26	22.03
Heterozygous	49	41.53
Homozygous polymorphic	43	36.44

*Including only patients taken to cytoreduction.

**Including only patients with recurrence/progression of disease.

SD: standard deviation; Q1-Q3: first quartile-third quartile; BMI: body mass index; ECOG: Eastern Cooperative Oncology Group; FIGO: International Federation of Gynecology and Obstetrics.

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Table S2. Sociodemographic, clinical, anatomical, and histopathological variables in relation to platin sensitivity of patients with ovarian adenocarcinoma, treated at the National Cancer Institute – Mexico from 2005 to 2018 (n = 118)

Variable	Non-platin sensitive ^a (n = 58)		Platin sensitive ^b (n = 60)		p-value*
	Mean	SD	Mean	SD	
Age, years	51.50	11.90	52.71	11.73	0.577
BMI, kg/m ²	26.94	5.50	27.39	5.08	0.642
Variable	Median	Q1-Q3	Median	Q1-Q3	p-value*
Initial CA125, IU/mL	1506.50	415.80-3823.50	324.50	93.50-2094.00	0.150
Initial HE4, IU/mL	116.15	84.83-172.80	49.05	36.23-73.03	0.285
Tumor size, cm	6.00	4.15-7.00	16.00	12.00-21.50	<0.001
Variable	n	%	n	%	p-value**
Initial ECOG performance status					
0	40	68.97	49	81.67	0.141
1	15	25.86	6	10.00	
2	3	5.17	4	6.67	
3	0	0.00	1	1.67	
Histological type					
High-grade papillary serous	44	75.86	19	31.67	<0.001
Low-grade papillary serous	0	0.00	2	3.33	
Endometrioid	5	8.62	16	26.67	
Clear cell tumor	2	3.45	9	15.00	
Other/mixed histology	7	12.07	14	23.33	
FIGO clinical stage					
I	0	0.00	26	43.33	<0.001
II	1	1.72	3	5.00	
III	35	60.34	22	36.67	
IV	22	37.93	9	15.00	
Cytoreduction					
Yes	53	91.38	60	100.00	0.061
Optimal cytoreduction ^c					
Yes	29	54.72	56	93.33	<0.001
Chemotherapy					
Received neoadjuvancy	53	89.83	21	35.00	<0.001
Received adjuvancy	43	72.88	47	78.33	1.000
Recurrence/progression of disease					
Recurrence or progression	58	100.00	20	33.33	<0.001
Site of recurrence^d					
Peritoneal	5	8.62	4	20.00	0.035
Nodal	11	18.97	5	25.00	
Distant	25	43.10	9	45.00	
Multiple locations/other	17	29.31	2	10.00	
MAD1L1 genotype					
Wild type	7	12.07	19	31.67	0.024
Heterozygous	25	43.10	24	40.00	
Polymorphic	26	44.83	17	28.33	

^ap-value calculated using Student's t-test.^{**}p-value calculated using Chi-squared test.^aDefined as patients who had a recurrence of disease before 1 year after the end of treatment or with progression of disease.^bDefined as patients who did not recur after treatment or who recurred over a year after the end of treatment.^cConsidering only patients who were cytoreduced.^dConsidering only patients with recurrence/progression of disease.

SD: standard deviation; Q1-Q3: first quartile-third quartile; FIGO: International Federation of Gynecology and Obstetrics; MAD1: mitotic arrest deficiency 1. ECOG: Eastern Cooperative Oncology Group; BMI: body mass index.



SCREENING TOOL FOR RESTRICTIVE AND OBSTRUCTIVE VENTILATORY ABNORMALITIES IN A POPULATION-BASED SURVEY

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Table S1: Prediction of pre-bronchodilator ventilatory spirometric abnormalities from pre-bronchodilator 6 s pocket spirometry

Spirometry parameters	AUC ^a	Sensitivity (95% CI)	Specificity (95% CI)	PPV ^b (95% CI)	NPV ^c (95% CI)	Best cutoff point ^d
FEV₁ %P						
Restrictive	91.8 (88.4-95.3)	87.5 (77.6-94.1)	84.3 (81.2-87.0)	38.7 (31.1-46.6)	98.4 (96.9-99.2)	87
Obstructive	89.0 (82.3-95.7)	81.3 (63.6-92.8)	84.6 (81.6-87.3)	21.0 (14.2-29.2)	98.9 (97.6-99.6)	87
Any ventilatory abnormality	91.0 (87.7-94.2)	84.8 (76.4-91.0)	84.3 (81.2-87.0)	47.1 (39.8-54.5)	97.1 (95.3-98.3)	87
FEV₆ %P						
Restrictive	87.1 (82.1-92.2)	88.9 (79.3-95.1)	75.8 (72.3-79.1)	29.4 (23.4-35.9)	98.4 (96.8-99.3)	85
Obstructive	73.3 (64.3-82.4)	62.5 (43.7-78.9)	77.4 (73.9-80.6)	11.7 (7.2-17.6)	97.2 (95.4-98.5)	84
Any ventilatory abnormality	82.9 (78.2-87.6)	80.0 (71.1-87.2)	75.8 (72.3-79.1)	35.3 (29.2-41.7)	95.8 (93.7-96.4)	85
FEV₁/FEV₆ %P						
Restrictive	61.2 (54.8-67.5)	62.5 (50.3-73.6)	60.1 (56.2-64.0)	15.1 (11.2-19.6)	93.4 (90.6-95.6)	105
Obstructive	83.0 (74.4-91.7)	66.7 (48.2-82.0)	89.0 (86.3-91.3)	23.9 (15.6-33.9)	98.1 (96.6-99.0)	97
Any ventilatory abnormality	67.9 (62.3-73.5)	69.5 (59.8-78.1)	60.1 (56.2-64.0)	22.3 (17.9-27.2)	92.3 (89.3-94.7)	105
Crude FEV₁/FEV₆						
Restrictive	60.2 (53.7-66.7)	73.6 (61.9-83.3)	46.0 (42.1-50.0)	13.4 (10.2-17.1)	93.9 (90.7-96.3)	88
Obstructive	80.7 (71.8-89.7)	63.6 (45.1-79.6)	88.5 (85.8-90.9)	22.3 (14.4-32.1)	97.9 (96.4-98.9)	79
Any ventilatory abnormality	66.6 (61.0-72.3)	67.6 (57.8-76.4)	60.0 (56.0-63.8)	21.8 (17.4-26.7)	91.8 (88.8-94.3)	86

^aArea under the curve; ^bpositive predictive value; ^cnegative predictive value; ^dby the highest Youden index, affording the same value to sensitivity and specificity; FEV₁: forced expiratory volume in 1 s; %P: percentage of predicted.

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Table S2. Prediction of post-bronchodilator ventilatory abnormalities from pre-bronchodilator 6 s pocket spirometer

Spirometry parameters	AUC ^a	Sensitivity (95% CI)	Specificity (95% CI)	PPV ^b (95% CI)	NPV ^c (95% CI)	Best cutoff point ^d
FEV₁ %P						
Restrictive	91.7 (88.1-95.2)	88.7 (78.1-95.3)	82.1 (78.7-85.1)	34 (26.9-41.0)	98.6 (97.1-99.4)	87
Obstructive	89.0 (77.6-1.00)	81.3 (54.4-96.0)	93.9 (91.7-95.7)	26.5 (14.9-41.1)	99.5 (98.4-99.9)	82
Any ventilatory abnormality	91.2 (87.5-94.9)	88.5 (79.2-94.6)	82.1 (78.7-85.1)	39.4 (32.1-47.1)	98.2 (96.6-99.2)	87
FEV₆ %P						
Restrictive	90.4 (86.8-94.0)	91.9 (82.2-97.3)	75.3 (71.6-78.7)	28.1 (22.0-34.8)	98.9 (97.4-99.6)	85
Obstructive	73.3 (57.8-87.3)	50.0 (24.7-75.3)	90.9 (88.2-93.1)	12.9 (5.7-23.9)	98.5 (97.1-99.4)	77
Any ventilatory abnormality	86.8 (82.3-91.2)	85.9 (76.2-92.7)	75.3 (71.6-78.7)	31.5 (25.3-38.2)	97.6 (95.7-98.8)	84
FEV₁/FEV₆ %P						
Restrictive	54.0 (46.7-61.3)	58.1 (44.8-70.5)	52.6 (48.5-56.7)	11.4 (8.1-15.4)	92.3 (88.9-94.9)	106
Obstructive	83.1 (71.1-95.0)	68.8 (41.3-89.0)	88.8 (86.0-91.3)	14.3 (7.4-24.1)	99.1 (97.8-99.7)	97
Any ventilatory abnormality	60.0 (53.1-66.9)	61.5 (49.8-72.3)	54.1 (50.0-58.2)	15.0 (11.3-19.5)	91.4 (88.0-94.1)	106
CRUDE FEV₁/FEV₆						
Restrictive	52.6 (45.1-60.2)	80.6 (68.6-89.6)	26.1 (22.6-29.8)	10.3 (7.7-13.3)	92.8 (87.7-96.2)	90
Obstructive	82.0 (70.7-93.3)	64.7 (38.3-85.8)	92.2 (89.8-94.2)	19.3 (10.0-31.9)	98.9 (97.6-99.6)	76
Any ventilatory abnormality	59.0 (52.0-65.9)	55.7 (44.1-66.9)	59.6 (55.5-63.5)	15.5 (11.5-20.3)	91.0 (87.6-93.6)	86

^aArea under the curve; ^bpositive predictive value; ^cnegative predictive value; ^dby the highest Youden index, affording same value to sensitivity and specificity; FEV₁: forced expiratory volume in 1 s; %P: percentage of predicted.



ECHINOCOCCUS GRANULOSUS HAPLOTYPES IN THE SCIENTIFIC LITERATURE

Supplementary Table 1. *Echinococcus granulosus* haplotypes informed in the literature

Reference	Haplotypes informed in literature*	Gene	Country
Alvarez-Rojas (2017)	EG01 ([Q250806]; EGp1 (AB522646); EgAus03 (KT968704); EgAus02 (KT968703); EgRUS7 (AB777904); EgCL1-EgCI21 (KX227116-KX227136))	cox1	Chile
Andresiuk (2013)	C1-C7; N1-N3 (KC579441-KC579451)	cox1 and nad1	Argentina
Barazesh (2019)	H1 (MF54127); H2 (MG672258); H3 (MH542404); H4 and H5 (MH542399, MH542406, and MH542395)	cox1 and nad1	Iran
Bold (2019)	H1 (EcMGL2); H2 (No registered); H3 (EcMGL15); H4 (No registered)	cox1	Mongolia
Boufana (2015)	EGUK01-EGUK03 (AF346403; AB786665; AF346403)	cox1	UK
Boufana (2014)	EgTu01-EgTu39 (KM014606-KM014644)	cox1	Tunisia
Corrêa (2018)	H1-H11 (MF421702-MF421712)	cox1	Chile
De la Rue (2011)	H1 (M84661); H2 (AF458872); H3 (EF367289); H4 (No registered)	cox1	Brazil
Farhadi (2015)	EGH1-EGH13 (KP859559-KP859571)	cox1	Iran
Gorgani-Firouzjae (2018)	H1 ([EgS2] MF346705; [EgB2] MF625021); H2 ([EgS1] MF625022); H3 ([EgS12] MF625020); H4 ([EgB6] MF449137); H5 ([EgB1] MF346706); H6 ([EgHum5] MG099696); H7 ([EgHum4] MG099695); H8 ([EgS11] MF625018); H9 ([EgS6] MF449131); H10 ([EgB3] MF625019)	cox1	Iran
Guo (2018)	H1-H15 (MH211191-MH211205)	cox1	China
Hajiallo (2012)	H1 ([IRCO1] HM563009-11; [IRND1] HM563023-26); H2 ([IRCO1] HM563011, [IRND2] HM563027); H3 ([IRCO1] HM563011, [IRND3] HM563028); H4 ([IRCO1] HM563010, [IRND4] HM563029); H5 ([IRCO1] HM563010, [IRND6] HM563031); H6 ([IRCO2] HM563012, HM563013, [IRND1] HM563023, HM563026); H7 ([IRCO2] HM563012, [IRND3] HM563028); H8 ([IRCO3] HM563014, [IRND5] HM563030); H9 ([IRCO4] HM563016, [IRND7] HM563032); H10 ([IRCO4] HM563017, [IRND8] HM563033); H11 ([IRCO4] HM563015, [IRND9] HM563034); H12 ([IRCO5] HM563018, HM563019, [IRND10] HM563035, HM563036); H13 ([IRCO6] HM563020, [IRND11] HM563037)	cox1 and nad1	Iran
Hammad (2018)	Hap_5; Hap_6; Hap_7; Hap_8; Hap_9 (No registered)	cox1	Iraq
Han (2019)	EgQH1 (MG674403); EgQH2 (MG674404); EgQH3 (MG674405); EgQH4 (MG674406); EgQH5 (MG674407); EgQH6 (MG674408); EgQH7 (MG674409); EgQH8 (MG674410); EgQH9 (MG674411); EgQH10 (MG674412); EgQH11 (MG674413); EgQH12 (MG674414); EgQH13 (MG674415); EgQH14 (MG674416); EgQH15 (MG674417); EgQH16 (MG674418); EmQH1 (MG674419); EsQH1 (MG674420);	cox1	Tibet (China)
Jafari (2018)	IfnN1 (KX186691.1); IfnC1 (KU360325.1); IfnN2 (KU925412.1); IfnC2 (KT254124.1); IfnN3 (KY495928.1); IfnC3 (EF367259.1); IfnN4 (KX853647.1); IfnC4 (KU360316.1); IfnN5 (KX298249.1); IfnC5 (KX29858.1); IfnN6 (KF437795.1); IfnC6 (KU360299.1); IfnN7 (CQ168810.1); IfnC7 (DQ269944.1); IfnN8 (KX010894.1); IfnC8 (KX010872.1); IfnN9 (KP751440.1)	cox1 and nad1	Iran
Kinkar (2017)	SPAS5 (KY766886); SPA1 (KY766900); SPA2 (KY766896); SPA4 (KY766897); SPA3 (KY766903)	Whole mitochondrial genome	Spain
Kinkar (2017)	FIN1- (KY766884)	Whole mitochondrial genome	Algeria (patient detected in Finland)
Kinkar (2017)	TUN1 (KY766885)	whole mitochondrial genome	Túnez
Kinkar (2017)	ARG1 (KY766882)	Whole mitochondrial genome	Argentina
Kinkar (2017)	IND2 (KY766891); IND1 (KY766902)	Whole mitochondrial genome	India
Kinkar (2017)	CHI1 (KY766890)	Whole mitochondrial genome	Chile
Kinkar (2017)	TUR4 (KY766888); TUR1 (KY766901); TUR2 (KY766904); TUR3 (KY766898)	Whole mitochondrial genome	Turkey
Kinkar (2017)	FRA3 (KY766889); FRA1 (KY766893); FRA2 (KY766892)	Whole mitochondrial genome	France
Kinkar (2017)	ALB1 (KY766883)	Whole mitochondrial genome	Albania
Kinkar (2017)	IRA4 (KY766887); IRA3 (KY766899); IRA1 (KY766894); IRA2 (KY766895);	Whole mitochondrial genome	Iran
Karamian (2017)	Cox1-Erlan-1 (KP751430); Cox1-Erlan-2 (KP751426)	cox1	Iran
Konyaev (2013)	EgRUS7 (AB777904); EgRUS10 (AB777907); EgRUS11 (AB777908); EgRUS8 (AB777905); EgRUS9 (AB777906); EcRUS7 (AB777914)	cox1	Russia
Laatamna (2019)	EG01 ([Q250806]; EG40 (AB88617); EgA01 (KX020323); EgA16 (KX020337); EgAus03 (KT968704); EgAlg01 (MG808282); EgAlg02 (MG808283); EgAlg03 (MG808284); EgAlg04 (MG808285); EgAlg05 (MG808286); EgAlg06 (MG808287); EgAlg07 (MG808288); EgAlg08 (MG808289); EgAlg09 (MG808290); EgAlg10 (MG808291); EgAlg11 (MG808292); EgAlg12 (MG808293); EgAlg13 (MG808294); EgAlg14 (MG808295); EgAlg15 (MG808296); EgAlg16 (MG808297); EgAlg17 (MG808298); EgAlg18 (MG808299); EgAlg19 (MG808300); EgAlg20 (MG808301); EgAlg21 (MG808302); EgAlg22 (MG808303); EgAlg23 (MG808304); EgAlg24 (MG808305); EgAlg25 (MG808306); EgAlg26 (MG808307); EgAlg27 (MG808308); EgAlg28 (MG808309); EgAlg29 (MG808310); EgAlg30 (MG808311); EgAlg31 (MG808312); EgAlg32 (MG808313); EgAlg33 (MG808314); EgAlg34 (MG808315); EgAlg35 (MG808316); EgAlg36 (MG808317); EgAlg37 (MG808318); EgAlg38 (MG808319); EgAlg39 (MG808320); EgAlg40 (MG808321); EgAlg41 (MG808322); EgAlg42 (MG808323); EgAlg43 (MG808324); EgAlg44 (MG808325); EgAlg45 (MG808326); EgAlg46 (MG808327); EgAlg47 (MG808328); EgAlg48 (MG808329); EgAlg49 (MG808330); EgAlg50 (MG808331); EgAlg51 (MG808332); EgAlg52 (MG808333); EgAlg53 (MG808334); EgAlg54 (MG808335); EgAlg55 (MG808336); EgAlg56 (MG808337); EgAlg57 (MG808338); EgAlg58 (MG808339); EgAlg59 (MG808340); EgAlg60 (MG808341); EgAlg61 (MG808342); EgAlg62 (MG808343); EgAlg63 (MG808344); EgAlg64 (MG808345); EgAlg65 (MG808346); EgAlg66 (MG808347); EgAlg67 (MG808348); EgAlg68 (MG808349)	cox1	Algeria
Laurimäe (2018)	IRA3 (MH300931); IRA4 (MH300932)	Whole mitochondrial genome	Iran
Laurimäe (2018)	ARG4 (MH300955); ARG5 (MH300956); ARG5 (MH300957); ARG5 (MH300958); ARG5 (MH300960); ARG7 (MH300962); ARG7 (MH300963); ARG8 (MH300964); ARG9 (MH300965); ARG10 (MH300966); ARG11 (MH300967); ARG12 (MH300968); ARG12 (MH300970)	Whole mitochondrial genome	Argentina
Laurimäe (2018)	KEN2 (MH300937); KEN3 (MH300938)	Whole mitochondrial genome	Kenya
Laurimäe (2018)	SUD1 (MH300939); SUD1 (MH300942); SUD1 (MH300943); SUD1 (MH300944); SUD1 (MH300945); SUD1 (MH300946); SUD1 (MH300947); SUD1 (MH300948); SUD1 (MH300949); SUD2 (MH300950); SUD2 (MH300951); SUD3 (MH300952);	Whole mitochondrial genome	Sudan
Laurimäe (2018)	MAU1 (MH300953); MAU2 (MH300954)	Whole mitochondrial genome	Mauritania
Laurimäe (2018)	Gmon (MH300971)	Whole mitochondrial genome	Mongolia
Laurimäe (2018)	MEX1 (MH300972); MEX2 (MH300973); MEX3 (MH300974); MEX4 (MH300975); MEX5 (MH300976); MEX5 (MH300977); MEX6 (MH300978); MEX7 (MH300979); MEX8 (MH300980); MEX9 (MH300981)	Whole mitochondrial genome	Mexico
Laurimäe (2018)	FRA2 (MH300987); FRA3 (MH300988); FRA4 (MH300989); FRA4 (MH300990); FRA4 (MH300991); FRA5 (MH300992); FRA5 (MH300993); FRA5 (MH300994); FRA6 (MH300995); FRA6 (MH300997); FRA6 (MH301000); FRA7 (MH301001); FRA8 (MH301002); FRA10 (MH301010); FRA10 (MH301011); FRA10 (MH301012); FRA12 (MH301015); FRA13 (MH301016); FRA14 (MH301017)	Whole mitochondrial genome	France
Laurimäe (2018)	POL1 (MH301003); POL2 (MH301004); POL3 (MH301005); POL4 (MH301006); POL5 (MH301007)	Whole mitochondrial genome	Poland
Laurimäe (2018)	LIT1 (MH301020)	Whole mitochondrial genome	Lithuania
Laurimäe (2018)	UKR1 (MH301021); UKR1 (MH301022)	Whole mitochondrial genome	Ukraine
Laurimäe (2018)	SER1 (MH300984)	Whole mitochondrial genome	Serbia
Laurimäe (2018)	ROM2 (MH300983)	Whole mitochondrial genome	Romania
Oskouei (2016)	S1 (No registered); S2 (No registered); S5 (No registered); S7 (No registered); S8 (No registered); S9 (KJ540227); S10 (No registered); S11 (No registered); S13-S18 (No registered); S19 (KJ540229); S20 (KJ540231); S21-S23 (No registered)	cox1	Iran
Marinova (2017)	H1 ([C1] KR070984, [S1] KR070969, [L1] KR070973, [ATP1] KR070978); H2 ([C1] KR070964, [N3] KR070986, [S1] KR070969, [L1] KR070973, [ATP1] KR070978); H3 ([C1] KR070964, [N5] KR070984, [L2] KR070974, [ATP1] KR070978); H4 ([C1] KR070964, [N2] KR070985, [S1] KR070969, [L1] KR070973, [ATP1] KR070978); H5 ([C1] KR070964, [N6] KR070989, [S1] KR070969, [L1] KR070973, [ATP1] KR070978); H6 ([C1] KR070964, [N4] KR070987, [S1] KR070969, [L1] KR070973, [ATP2] KR070979); H7 ([C2] KR070965, [N1] KR070984, [S1] KR070969, [L1] KR070973, [ATP1] KR070978); H8 ([C1] KR070964, [N7] KR070991, [S1] KR070969, [L1] KR070973, [ATP1] KR070978); H9 ([C3] KR070966, [N7] KR070990, [S2] KR070970, [L4] KR070976, [ATP4] KR070981); H10 ([C3] KR070966, [N7] KR070990, [S2] KR070970, [L3] KR070975, [ATP3] KR070980); H11 ([C3] KR070966, [N7] KR070990, [S3] KR070971, [L3] KR070975, [ATP3] KR070980); H12 ([C3] KR070966, [N7] KR070990, [S2] KR070970, [L3] KR070975, [ATPS1] KR070982); H13 ([C4] KR070967, [N7] KR070990, [S2] KR070970, [L3] KR070975, [ATP3] KR070980); H14 ([C5] KR070968, [N9] KR070992, [S4] KR070972, [L5] KR070977, [ATP6] KR070983)	cox1, nad1, rrnS, rRNA, atp6	Bulgaria
Matini (2019)	Hamc1 (MG792551); Hamc2 (MG792552); Hamc3 (MG792553); Hamc4 (MG792554); Hamc5 (MG792555); Hamc6 (MG792556); Hamc7 (MG792557); Hamc8 (MG792558); Hamc9 (MG792559); Hamc10 (MG792560); Hamc11 (MG792561); Hamc12 (MG792562); Hamc13 (MG792563)	cox1	Iran
Metwally (2018)	Echs1-Echs12 (No registered); Echs1 (No registered); Echs2 (No registered)	cox1	Saudi Arabia
Moro (2009)	Egra-A (AB458672); Egra-B (AB458673); Egra-C (AB458674); Egra-D (AB458675); Egra-E (AB470527); Ecan-A (AB458676); Ecan-B (AB458677); Ecan-C (AB458678)	cox1	Peru
Nikmanesh (2014)	H1 (KF612381, KF612357); H2 (KF612356); H3 (KF612390, KF612360); H4 (KF612380, KF612358); H5 (KF612376, KF612349); H6 (KF612395, KF612350); H7 (KF612396, KF612351); H8 (KF612394, KF612343); H9 (KF612386, KF612351); H10 (KF612397, KF612351); H11 (KF612400, KF612372)	cox1 and nad1	Iran
Orsten (2018)	TUK01-TUK28 (no registered)	cox1	Turkey
Parra (2012)	H1 ([Lor1] JN604097, [Lor2] JN604107); H2 ([Lor1] JN604097), [Lor3] JN604108); H3 ([Lor1] JN604097, [Lor4] JN604109); H4 ([Lor1] JN604097, [Lor5] JN604110); H5 ([Lor1] JN604097, [Lor6] JN604111); H6 ([Lor2] JN604098), [Lor7] JN604109); H7 ([Lor2] JN604107); H7 ([Lor3] JN604107); H7 ([Lor4] JN604107); H7 ([Lor5] JN604107); H8 ([Lor4] JN604106); H8 ([Lor5] JN604110); H9 ([Lor5] JN604101); H10 ([Lor6] JN604101); H10 ([Lor7] JN604102); H10 ([Lor8] JN60410		