

## Effect of PGPB bacteria, compost and digestate on the dry matter yield of cocksfood

Gisela Aguilar-Benítez<sup>1</sup>  
María Myrna Solís-Oba<sup>2</sup>  
Rigoberto Castro-Rivera<sup>2§</sup>  
Valentín López-Gayou<sup>2</sup>  
José Pablo Lara-Ávila<sup>3</sup>  
Marco Antonio Esteves-Luna<sup>4</sup>

<sup>1</sup>Desert Research Institute-Autonomous University of San Luis Potosí. Altair 200, Colonia del Llano, San Luis Potosí. CP. 78377. (gisela.aguilar@uaslp.mx). <sup>2</sup>National Polytechnic Institute-Center for Research in Applied Biotechnology. Ex Hacienda de San Juan Molino, Tecuexcomac-Tepetitla Highway km 1.5, Tlaxcala. CP. 90700 (mirobatlx@hotmail.com; valgayou@gmail.com). <sup>3</sup>Faculty of Agronomy and Veterinary Medicine-Autonomous University of San Luis Potosí, San Luis Potosí-Matehuala highway km 14.5, Ejido Palma de la Cruz, Soledad de Graciano Sánchez, SLP. CP. 78439. (pablo.lara@uaslp.mx). <sup>4</sup>Tlaxcala Altiplano Technological Institute. Federal highway San Martín Texmelucan-Tlaxcala km 7.5, San Diego Xocoyucan, Tlaxcala. CP. 90122. (mael071260@hotmail.com).

§Corresponding author: rcastror@ipn.mx.

### Abstract

The aim of the present study was to determine the effect of compost, digestate and PGPB (plant growth-promoting bacteria) on yield forage accumulation curve, growth rate, forage height and SPAD (soil plant analysis development) units in newly established of cocksfood under greenhouse conditions. The treatments were digestate (60%); compost (10% on dry soil based); bacteria: *Brevibacterium frigoritolerans*, *Bacillus simplex* and *Pseudomonas putida*; positive control (triple 17 fertilization) and negative control (soil without fertilization). A completely random experimental design was used, the experimental unit was a pot with ten tillers, with four replicates by treatment. The highest values ( $p < 0.05$ ) of dry matter yield (6.4 g DM pot), growth rate (0.15 g DM pot d<sup>-1</sup>) and forage height (18.3 cm) were recorded in treatment with compost, with which the final yield was 200% higher than the negative control. The treatment with digestate showed lower values than those obtained with compost but surpassed the rest of the treatments. The best PGPB were *Pseudomonas putida* and *Bacillus simplex*, which outperformed *Brevibacterium frigoritolerans* and negative control by 25 and 37%, respectively. The PGPB bacteria can be a fertilization alternative since the yield was higher than with the negative control and was equal to the yield with chemical fertilization; however, the two organic fertilizers (compost and digestate) favored the higher yield.

**Keywords:** *Bacillus simplex*, *Brevibacterium frigoritolerans*, *Dactylis glomerata*, *Pseudomonas putida*, forage yield and.

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## Introduction

Forage yield is determined by various factors such as temperature, humidity and fertilization (Ahmad *et al.*, 2016). Mineral fertilization of forage grasses is a necessary activity to increase yields and to cope with the low natural or induced fertility of the soils, this practice is expensive and has adverse environmental implications if done improperly. An alternative to the application of mineral fertilizers in forage crops is organic fertilizers, which provide N in organic forms such as proteins and amino acids, more or less stable, which gradually become mineralized in forms assimilable by plants (Ramos and Terry, 2014).

The use of plant growth promoting bacteria (PGPB) promotes better absorption of nutrients in the soil due, among others, to the ability of bacteria to solubilize phosphorous, fix nitrogen and synthesize siderophores, also produce phyto-stimulant substances (auxins, gibberellins, cytokinins) or can act as stress controllers (biological control through antagonistic activity against phytopathogenic microorganisms) in the plant (De-Bashan *et al.*, 2007; Pérez-Montañaño *et al.*, 2014; Menna *et al.*, 2017; Singh *et al.*, 2017).

Lopes *et al.* (2018) evaluated the effect of inoculation of PGPB bacteria on forage yield in *Brachiaria brizanta*, subjected to different radiation intensities, the results were higher yield, chlorophyll content, leaf area index and root weight ( $p < 0.05$ ) with pure and associated bacteria treatments compared to controls.

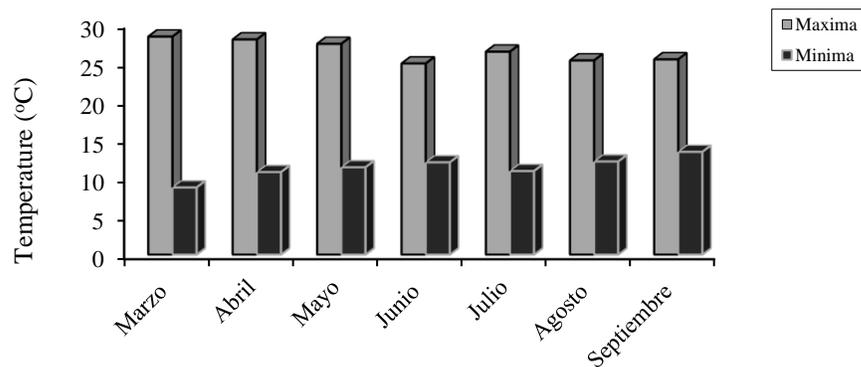
Digestates are the liquid by-product of anaerobic digestion for the production of biogas, from organic solid waste. This by-product is used as a fertilizer since they contain phytohormones (gibberellins and indolacetic acid) dissolved in organic matter, a considerable amount of microorganisms, as well as other bioactive compounds that have the potential to promote plant growth and increase stress tolerance biotic and abiotic (Yu *et al.*, 2010).

The benefits of compost in crops have been reported in different works. Beltrán *et al.* (2017) estimated the forage yield of Triticale fertilized with compost obtained from feces of dairy cattle and observed that the combination of this fertilizer with inorganic fertilizer promotes a higher dry matter yield, forage height and higher stem density, compared to unfertilized plants and fertilized only with inorganic fertilizer. For the foregoing, the aim of the present work was to evaluate the effect of the different types of organic fertilization on the regrowth pattern of cocksfoot, under greenhouse conditions.

## Materials and methods

### Description of the experimental area

The experiment was carried out in a tunnel-type plastic greenhouse with side windows of the Center for Research in Applied Biotechnology of the National Polytechnic Institute, located in Tepetitla de Lardizabal, Tlaxcala (19° 16' 50.3" north latitude, 98° 21' 58.1" west longitude, 2 221 masl). The average outdoor temperature is shown in Figure 1.



**Figure 1. Maximum average temperature and minimum average environmental temperature, in Tepetitla de Lardizabal, Tlaxcala.** <https://www.accuweather.com/es/mx/tepetitla/240244/weather-forecast/240244>.

The cocksfoot seeds (*Dactylis glomerata* L.) were donated by the Forage Laboratory of the Livestock Program of the Postgraduate College. The digestate was obtained from the Experimental Farm of the Department of Zootecnics of the Autonomous University Chapingo (UACH). The compost was donated by the Zacatenco composting unit of the National Polytechnic Institute. The soil used as a substrate was obtained from the experimental plot of the CIBA IPN Unidad Tlaxcala, which was identified as fluvisol with a sandy texture.

### Experiment development

Cocksfoot was sown by placing 15 seeds in plastic pots containing 1.5 kg of soil (experimental unit). Once the seedlings emerged, manual thinning was performed to leave only 10 stems per pot and an establishment period of 45 days was left after planting. Subsequently, a uniform cut was made at a height of five cm, to reduce the effect of the covariate, and the PGPB bacteria were inoculated and organic fertilizers were applied in the early spring of 2018.

### Selection and inoculation of PGPB batteries

Serial dilutions were made of the soil, compost and digestate samples. 1 ml was grown in Petri dishes at three dilutions  $10^{-2}$ ,  $10^{-4}$  and  $10^{-6}$ , with an incubation period of 24 h at 30 °C. Pure cultures were obtained to describe the particular characteristics and with the selected morphotypes, *Paenibacillus*, *Variovorax*, *Lysobacter*, *Azospirillum*, *Streptomyces*, *Streptomyces*, *Pikovskaya*, *Ashby*, *NFb* and *NBRIP* were inoculated into the selective media (Bashan and Holguin, 1997; Noumavo *et al.*, 2013; Beghalem *et al.*, 2017).

Bacteria that grew in these media were re-inoculated into *Pikovskaya*, *Ashby*, *NFb* and *NBRIP* specific media to evaluate their potential as potassium solubilizers, nitrogen fixers and phosphorus solubilizers, respectively. The strains selected for this experiment were identified as *Pseudomonas putida*, *Bacillus simplex* and *Brevibacterium frigoritolerans*, which were previously identified by means of 16S rRNA sequencing.

The inoculation of the bacteria was carried out directly in the soil, in the rhizosphere of the tillers, at the beginning of the season, with a sterile 1 mL syringe of nutrient broth at a concentration of  $1 \times 10^8$  CFU mL<sup>-1</sup> per experimental unit.

### Treatment and experimental design

A completely randomized design and four repetitions per treatment were used, which were: concentration of digestate (60%), compost (10% in dry soil), bacteria: *Brevibacterium frigoritolerans*, *Bacillus simplex*, *Pseudomonas putida*, positive control (fertilization with triple 17) and the negative control (soil without fertilization).

### Variables evaluated

The dry matter yield was obtained from cuts at weeks one, two, three, four, five, six and seven. The harvest height was five centimeters, placing all the cut forage in previously labeled paper bags. The harvested plant material was washed and weighed fresh, and then dried in a forced air stove at 70 °C, for 48 h to a constant weight, and the proportion of dry matter was determined.

Before each cut, the height of the forage to the recently exposed top leaf was recorded in randomly chosen plants, with a graduated ruler of 30 cm and an accuracy of 0.1 cm (Castillo *et al.*, 2009).

The growth rate (TC) was calculated with the dry matter yield data per cut using the following formula.

$$TC = \frac{FC}{t}$$

Where: FC= forage harvested (g MV pot) and t= days elapsed between one cut and the next. The chlorophyll content (SPAD units) were recorded before each cut, taking 3 samples per experimental unit, placing the Apogee instruments MC-100 instrument sensor on the exposed upper leaf with the ligule well differentiated.

### Statistical analysis

The values grouped by week of growth were plotted using the SigmaPlot V.10 statistical software and analyzed using the GLM and PROC MIXED procedures of the Statistical Software SAS<sup>®</sup> version 9.0 for Windows<sup>®</sup>. The treatment means comparison was made using Tukey at a significance level of 5%.

## Results and discussion

The highest average height of the cocksfoot was obtained with the compost treatment (10.5 cm); however, statistically there was no difference ( $p > 0.05$ ) with the treatment with digestate and *Pseudomonas putida*, in addition these last two treatments did not present differences ( $p > 0.05$ )

with the rest of the treatments. The average height was higher ( $p > 0.05$ ) in the plants harvested at seven weeks and this variable showed no difference ( $p > 0.05$ ) in the cuts at five and six weeks, despite the fact that at six weeks the higher forage yield (Table 1).

**Table 1. Height of cocksfoot plant (*Dactylis glomerata* L.) at different weeks of growth after a homogenization cut.**

Treatments	Growth weeks							Pom.	SEM	Sig.
	1	2	3	4	5	6	7			
Compost	6.13Ad	6.9Ad	8.6Ac	9.6Ac	11.3Ab	12.6Ab	18.3Aa	10.5A	0.42	**
Digestate	5.1Bd	5.6Bd	6.9BCd	9.1Ac	9.6Bbc	11.4Ab	16.2Aa	9.13AB	0.55	**
<i>P. putida</i>	5.2Be	5.3Be	7.7ABd	8.7Acd	10.4ABcb	12.4Ab	15ABa	9.25AB	0.59	**
<i>B. simplex</i>	5.2Bc	5.6Bc	7.1BCbc	7.1Bbc	7.8Cb	8.3Bb	10.3Ca	7.36B	0.54	**
<i>B. frigiditolerans</i>	5.3Bc	5.4Bc	5.3Dc	5.9Bbc	6.3Cbc	7.7Bb	13.8ABa	7.12B	0.56	**
Chemical	5.1Bd	5.2Bd	5.5Dd	6.3Bcd	7.6Cbc	9.3Bb	12.2BCa	7.31B	0.56	**
Soil	5.3Bd	5.9ABcd	6.3CDcd	6.4Bcd	7.3Cbc	8.6Bab	9.5Da	7.06B	0.42	**
Average	5.35e	5.73e	6.76ed	7.58cd	8.64bc	10.3b	13.63a			**
SEM	0.15	0.32	0.35	0.39	0.43	0.61	0.86			
Sig.	**	**	**	**	**	**	**	**		

Different lowercase literals in a row are statistically different Tukey ( $p < 0.05$ ). Different capital letters in columns are statistically different Tukey ( $p < 0.05$ ). Average (Prom.); significance (Sig.); \* = 0.05; \*\* = 0.01.

Likewise, the compost treatment promoted the highest heights ( $p < 0.05$ ) in all the cutting weeks, while *P. putida* and the digestate showed the highest heights from week four, but their values were lower than those obtained with compost, although superior to the rest of the treatments.

The SPAD units in the compost and digestate treatments were higher ( $p < 0.05$ ), during all the weeks of growth evaluated, whereas with *B. simplex*, *B. frigiditolerans* and in the positive and negative controls the values were not different ( $p < 0.05$ ). With the exception of the treatments with inorganic fertilizer and *B. frigiditolerans*, in the rest of the treatment's differences were observed ( $p < 0.05$ ) in the SPAD values registered in the different weeks of growth (Table 2). With *S. putida* in week one and two there were no differences with respect to the rest of the treatments, but from the third week the SPAD units increased, equaling the values obtained with compost and digestate.

**Table 2. Chlorophyll content (SPAD units  $\text{cm}^2$ ) of cocksfoot (*Dactylis glomerata* L.) in the different weeks of growth after a uniform cut.**

Treatments	Growth weeks							Prom.	SEM	Sig.
	1	2	3	4	5	6	7			
Compost	3.2A	3.1	3.2A	3.2A	3.3A	3.5A	3.4A	3.28	0.15	**
Digestate	3.1A	2.9	3.2A	3AB	3.1AB	3.5A	3.2A	3.16	0.32	**
<i>P. putida</i>	3.3A	3	2.1B	2.9AB	2.2C	2.6B	3.1	2.75	0.42	**

Treatments	Growth weeks							Prom.	SEM	Sig.
	1	2	3	4	5	6	7			
<i>B. simplex</i>	2.1Bb	2.1b	2.2Bb	2.2ABb	2.4BCab	2.3Bb	2.9ABa	2.3C	0.17	**
<i>B. frigoritolerans</i>	2.2B	2.3	2.1B	2.3AB	2.1C	2.4B	2.1C	2.24	0.15	**
Chemical	2.1B	2.4	2.2B	2.2AB	2.2C	2.2B	2.3BC	2.25	0.19	**
Soil	1.2Cc	2.1b	1.9Bb	2.1Bb	2.2Cb	2.3Bb	2.9ABa	2.08	0.14	**
Prom.	2.46	2.57	2.42	2.56	2.49	2.69	2.87			ns
SEM	0.11	0.37	0.14	0.28	0.24	0.23	0.17			
Sig.	**	**	**	**	**	**	**	**	**	

Different lowercase literals in a row are statistically different Tukey ( $p < 0.05$ ). Different capital letters in columns are statistically different Tukey ( $p < 0.05$ ). Average (Prom.); Significance (Sig.); \* = 0.05; \*\* = 0.01.

Regarding the growth rate, the highest values were obtained in the compost and digestate treatments. For this variable, *P. putida* outperformed ( $p < 0.05$ ) the other PGPB bacteria and both controls, and it is observed that the treatment with compost, during the entire evaluation period, was the one with the highest growth rate, otherwise, with digestate this variable only increased in week six (Table 3).

**Table 3. Growth rate (g DM pot d<sup>-1</sup>) of cocksfoot (*Dactylis glomerata* L.) in the different weeks of growth after a uniform cut.**

Treatments	Growth weeks							Prom.	SEM	Sig.
	1	2	3	4	5	6	7			
Compost	0.13Ab	0.1Ac	0.11Ac	0.11Ac	0.11Ac	0.15Aa	0.11Ab	0.11A	0.003	**
Digestate	0.09Ba	0.06BCb	0.058Bb	0.049BC	0.055BCb	0.099Ba	0.06BCb	0.06C	0.006	**
<i>P. putida</i>	0.08BCa	0.07Bab	0.059Bb	0.051Bc	0.059Bc	0.073Ca	0.05BCd	0.07B	0.003	**
<i>B. simplex</i>	0.08Ca	0.06BCb	0.053BC	0.048CD	0.056BCDb	0.085B	0.057Bb	0.05D	0.004	**
<i>B. frigoritolerans</i>	0.08BCa	0.064BC	0.047CD	0.041Ec	0.04Ec	0.066C	0.031Cc	0.054D	0.005	**
Chemical	0.077BC	0.051CD	0.046CD	0.043ED	0.043EDb	0.075Ca	0.047BC	0.054D	0.005	**
Soil	0.064Ca	0.046Da	0.037Db	0.036Fb	0.046CDEb	0.05Dab	0.042BC	0.051E	0.005	**
Prom.	0.085	0.065b	0.058c	0.054	0.058c	0.083a	0.064		0.0011	**
SEM	0.006	0.004	0.003	0.001	0.003	0.006	0.006			
Sig.	**	**	**	**	**	**	**	**	**	

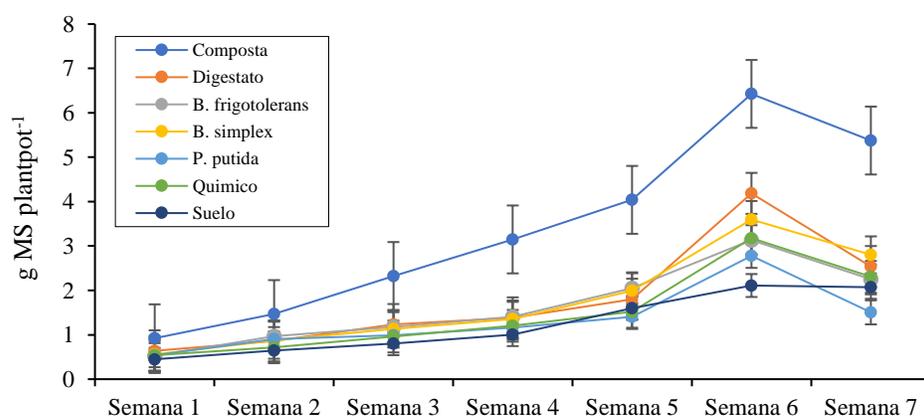
Different lowercase literals in a row are statistically different Tukey ( $p < 0.05$ ). Different capital letters in columns are statistically different Tukey ( $p < 0.05$ ). Average (Prom.); Significance (Sig.); \* = 0.05; \*\* = 0.01.

The highest dry matter yield and the highest forage accumulation ( $p < 0.05$ ) were achieved in week six, regardless of the treatment evaluated. However, the highest average value ( $p < 0.05$ ) per treatment was obtained with compost (3.4 g DM pot), followed by digestate (1.9 g DM pot). The lowest values ( $p < 0.05$ ) of dry matter were obtained with the negative control and *Brevibacterium frigoritolerans* and the treatments of *Pseudomonas putida*, *Bacillus simplex* and the chemical fertilizer showed no differences (Table 4). The highest yield from week one to seven was achieved with the compost, with the digestate no differences were distinguished from week one to five and it was until week six that it was different ( $p < 0.05$ ) from the rest of the treatments (Figure 2).

**Table 4. Forage yield (g DM pot), of cocksfood (*Dactylis glomerata* L.) in the different weeks of growth after a uniform cut.**

Treatments	Growth weeks							Prom.	SEM	Sig.
	1	2	3	4	5	6	7			
Compost	0.92Ag	1.46Af	2.3Ae	3.14Ad	4.03Ac	6.4Aa	5.37Ab	3.4A	0.13	**
Digestate	0.67Bd	0.86BCDd	1.23BCcd	1.39BCcd	1.95BCc	4.18Ba	2.96BCb	1.9B	0.23	**
<i>P. putida</i>	0.53BCe	0.99Bd	1.25Bcd	1.45Bc	2.1Bb	3.1Ca	2.3BCb	1.7C	0.091	**
<i>B. simplex</i>	0.55BCg	0.87BCde	1.3BCde	1.35BCcd	1.99Bc	3.6BCa	2.8Bb	1.6C	0.22	**
<i>B. frigoritolerans</i>	0.54BCc	0.9BCbc	0.98CDbc	1.15CDbc	1.45Cb	2.8CDa	1.5Cb	1.3D	0.21	
Chemical	0.54BCd	0.71CDd	0.96BCDcd	1.2BCDcd	1.5BCc	3.2Ca	2.3BCb	1.6C	0.20	**
Soil	0.44Ce	0.65Dde	0.8Dcd	1Dc	1.6BCb	2.1Da	2.06BCa	1.2D	0.09	**
Average	0.59g	0.91f	1.22e	1.52d	2.05c	3.68a	2.69 b		0.044	**
SEM	0.053	0.067	0.088	0.064	0.16	0.26	0.30	0.06		
Sig.	**	**	**	**	**	**	**	**		**

Different lowercase literals in a row are statistically different Tukey ( $p < 0.05$ ). Different capital letters in columns are statistically different Tukey ( $p < 0.05$ ). Average (Prom.); Significance (Sig.); \* = 0.05; \*\* = 0.01.



**Figure 2. Cocksfood dry matter yield under different fertilization schemes, during the spring time of 2018.**

In the literature it has been reported that the factors that determine the speed and magnitude of growth of a forage plant are: climate (radiation, photoperiod, temperature, humidity and precipitation) (McKenzie *et al.*, 1999), soil (physical, chemical characteristics, topographic and fertilization); species (photosynthetic route, genetic potential of different species and varieties); and grassland management (intensity and frequency of defoliation) (Mendoza-Pedroza *et al.*, 2018). All the factors do not act separately and the growth of the plant responds to an interaction between them (Jiménez and Martínez, 1984; Hernández-Garay *et al.*, 1997; Moliterno, 2002; Ganderats *et al.*, 2003). However, in this experiment the forage species evaluated was subjected to the same conditions of temperature, humidity and radiation, modifying the source of fertilization; therefore, it is evident that the fertilization source affects the yield of the species.

It has been reported that the addition of compost modifies the physical, chemical and microbiological properties of the soil, generating more favorable conditions for the plant, which is reflected in a higher yield (Ramos and Terry 2014; Liu *et al.*, 2016). Furthermore, the benefits of compost have been evidenced in various studies with vegetables and there are few studies published in temperate pastures, so the results presented in this document provide a first of study in forage grasses with temperate climates.

Regarding the results obtained with the digestate, these coincide with what was stated by Rancane *et al.* (2015); Walsh *et al.* (2012), who observed an increase in forage yield with the addition of digestate. For their part, Tilvikiene *et al.* (2018) evaluated the biomass yield in cocksfoot fertilized with different concentrations of digestates for five consecutive years and noted that fertilization with this fertilizer causes high concentrations of structural components (cellulose and hemicellulose) in the biomass, which has a greater impact forage yield.

Montalvo-Aguilar *et al.* (2018), warned that irrigation with digestate (60%) after each cut favors the increase in forage yield of *Lolium perenne*, obtaining more than 150% of yield compared to the control (soil alone). Likewise, with the highest concentrations of digestate, the highest protein levels were observed (28% at the four-week cutoff frequency), and this variable is correlated with the values obtained in this experiment with the SPAD units (González-Torres *et al.*, 2009).

The results obtained with the different PGPB bacteria coincide with that reported by Lopes *et al.* (2018), since they showed higher yields compared to the control that was unfertilized and inoculated soil. However, in the results obtained in this work only with *Pseudomonas putida*, high values were obtained in the evaluated variables. Rangel *et al.* (2014) report that when evaluating PGPB bacteria in the yield of corn, wheat and sorghum, only *Azospirillum brasilense* registered beneficial effects, exceeding 55 and 49% in soil alone and inorganic fertilizer, respectively.

Criollo *et al.* (2012) evaluated the effect of PGPB bacteria associated with *Penissetum clandestinum*, at different regrowth times (70, 100 and 130 days after sowing), reporting that bacteria of the *Pseudomonas* genus showed an effect on yield up to 130 days after regrowth, surpassing 150% the root weight with respect to the control. In C4 grasses, it is mentioned that for the *Brachiaria* genus, inoculating with PGPB bacteria is equivalent to a dose of 40 kg of nitrogen per hectare (Hungria *et al.*, 2016).

The bacteria evaluated in this experiment have been reported as PGPB bacteria, by different authors, and their benefits as phosphate solubilizers, nitrogen fixers, and siderophore synthesizers have been mainly evaluated in culture media and *in vitro* conditions (Rashid *et al.* (2012). Other authors have evaluated them in the field such as Terry *et al.* (2005) who reported the beneficial effects of the genera *Pseudomonas* and *Bacillus* on tomato (*Lycopersicon esculentum* L.). However, the species evaluated in this work have not been evaluated in grasses of zootechnical interest and have not been reported in scientific articles.

## Conclusions

PGPR bacteria can be a fertilization alternative since the evaluated variables increased their values with respect to the soil without fertilization and were equal to chemical fertilization; however, the two organic fertilizers (compost) and the digestate were superior to all the treatments evaluated. From an economic point of view, bacteria may be the best fertilization option, since these were only inoculated at the beginning of the experiment, while digestates were applied at each uniform cut and the percentage of compost in the soil is equivalent to one approximate dose of 60 t ha<sup>-1</sup>.

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