New Isoniazid Complexes, Promising Agents Against Mycobacterium tuberculosis

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Abstract. Tuberculosis (TB) is a public health disease that produces several millions deaths annually worldwide. Due to this critical situation and the appearance of drug-resistant microbial strains, innovation in TB drug discovery is a research priority. In this work, the synthesis and characterization by elemental analysis, thermogravimetry, conductimetric measurements and spectroscopies UV-Vis, IR and EPR of [Cu(INH)(H2O)]SO42H2O (Cu-INH) and [CoCl(INH)2(H2O)]Cl2.5H2O (Co-INH) complexes with isoniazid (INH) are reported. Besides, the lipophilicity, the activity against Mycobacterium tuberculosis (MICCu-INH = 0.78 µg/mL and MICCo-INH = 0.19 µg/mL) and the cytotoxicity (IC50 = 48.8 and 625 µg/mL for the copper and cobalt complexes, respectively) were measured and the selectivity index (62.5 for Cu-INH and 3205 for Co-INH) was calculated. These results indicate that these complexes are good candidates for further studies. Keywords: copper and cobalt complexes, isoniazid, antimycobacteria activity, tuberculosis, spectroscopy.

Introduction

Tuberculosis (TB) is a public health disease that produces several millions deaths annually worldwide [1]. In developing countries is a leading cause of mortality. Nowadays, the number of infections is growing also in developed countries, especially in immunocompromised patients such as those co-infected with human immunodeficiency virus (HIV), and in individuals receiving anti-tumour therapy or diabetes [2-4].

A big associated problem is the alarming increase of drug-resistant microbial strains (MDR-TB) that makes difficult the effective control of the disease. Besides, to further complicate the matter, drug-drug interactions between TB drugs and anti-HIV treatments or other chronic disease medications such as those used in diabetics are observed [2].

Due to this critical situation, innovation in TB drug discovery and evolving strategies to bring new agents with best performance is a current health priority.

One strategy to obtain new antimycobacterial drugs is the development of new molecules by reengineering old drugs with the aim to improve the antimycobacteria activity and to obtain better resistance profiles, bioavailability and tolerability, among others [3]. In particular, the coordination of metal cations with organic drugs is a promising strategy that has been successful in many cases with different pharmacological activities [5-7]. In particular, in the research of new antimycobacteria compounds with best activity and/or lower resistant effects, several metallic complexes were reported in the literature. For example, silver complexes with α-hydroxyxcarboxilic acids have proved to be effective antimycobacterial compounds, potential candidates for antiseptic or disinfectant products [8]. On the other hand, several series of homoleptic and heteroleptic iron, copper and ruthenium compounds were developed being more active than the free ligands against Mycobacterium tuberculosis (M. tuberculosis) [9-14].

Taking into account these antecedents and as a part of our work in the research of new antimicrobial molecule [15-19], in this article the development of new Cu(II) and Co(II) complexes with isoniazid (INH) is reported and the activity against M. tuberculosis is evaluated. Besides, the cytotoxicity in mammalian cells was determined and the selectivity index was calculated.

Isoniazid (Figure 1), the first effective drug discovered for the treatment of tuberculosis [20], is included in the group of the first-line antimycobacterial drugs used in prevention and treatment of tuberculosis. However, shortly after its discovery,
INH resistant *M. tuberculosis* strains were reported [21]. Although this, INH is one of the most used drugs today. INH is able to coordinate with metal cations through different chemical groups: heterocyclic nitrogen from the pyridine ring and/or carbonylic O and N atoms of the hydrazide group. For this versatility it is also an interesting ligand from the chemical point of view.

### Results and discussion

#### Analytical results

The elemental analyses of the complexes were:

- **Cu-INH** (Code Cu-INH), *Anal C* 21.2%, N 11.9%, H 3.4%, calcld for C₆N₃H₁₃O₈SCu, C 21.5%, N 12.0%, H 3.7%; yield: 70%, 135mg.

- **Co-INH** (Code Co-INH), *Anal C* 31.3%, N17.2%, H4.7%, Cl 15.4%, calcld for C₁₂N₆H₂₁O₅.₅CoCl₂, C 30.8%, N 17.9%, H 4.4%, Cl 15.2%; yield: 76%, 196 mg.

These results agree with the proposed stoichiometries.

#### Thermogravimetric and conductimetric measurements

Thermogravimetric results are shown in Table 1. Experimental mass losses agree with calculated values for the two complexes. The thermal process corresponding to Cu-INH firstly shows the release of two hydration water molecules at 58.5 °C while the release of the coordinated water molecule took place at 112.9 °C, as expected. The decomposition of the organic moiety starts above 140 °C forming the copper oxide at the end of calcination. Similarly, the thermogram of Co-INH indicates that the complex has 2.5 hydration water molecules and one coordinated molecule.

Results of the conductimetric measurements in dimethyl sulfoxide (DMSO) solutions are shown in Table 2. According to previous reports the conductivities obtained for both complexes are in agreement with the assigned formula for a 1:1 electrolyte type in DMSO [22-23]. In addition, the conductivity was measured in the same solutions after 48 hs and no major changes were observed showing the stability of the complexes in DMSO.

#### Spectroscopic measurements

**Infrared spectra**

The IR spectra of the complexes were compared with those of the free ligands and previously reported complexes [24-27]. Table 3 shows the tentative assignments of the main bands of free isoniazid and the metal complexes.

**Analysis of Cu-INH spectrum**

According to bibliographic reports [24, 25], the INH spectrum presents two bands at 3300 and 3209 cm⁻¹ corresponding to asymmetric and symmetric stretching of NH₂ group. Upon copper complexation the ν(NH₂) shifted to lower frequencies while the ν(N-H) didn’t shift in the complex showing that this group does not participate in the coordination. On the other hand, INH spectrum shows one band at 1667 cm⁻¹ assigned as ν(C=O). In the complex

### Table 1. Thermal data for the metal-INH complexes.

<table>
<thead>
<tr>
<th>Complex</th>
<th>Temperature range (°C)</th>
<th>DrTGA minima (°C)</th>
<th>Mass loss (%) Found/Calc.</th>
<th>Removed group</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cu-INH</td>
<td>20-80</td>
<td>58.5</td>
<td>10.6/10.3</td>
<td>2 H₂O</td>
</tr>
<tr>
<td></td>
<td>80-140</td>
<td>112.9</td>
<td>4.6/5.1</td>
<td>H₂O</td>
</tr>
<tr>
<td>Co-INH</td>
<td>20-80</td>
<td>40.4</td>
<td>9.2/9.6</td>
<td>2.5 H₂O</td>
</tr>
<tr>
<td></td>
<td>80-130</td>
<td>100.3</td>
<td>3.9/3.8</td>
<td>H₂O</td>
</tr>
</tbody>
</table>

### Table 2. Conductivity (µS) in DMSO at 25 °C.

<table>
<thead>
<tr>
<th>Complex</th>
<th>Λ DMSO (µS) at 25°C</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cu-INH</td>
<td>35.3</td>
</tr>
<tr>
<td>Co-INH</td>
<td>52.0</td>
</tr>
</tbody>
</table>

### Table 3. Main IR bands and their assignment for Cu-INH and Co-INH.

<table>
<thead>
<tr>
<th>INH/Cu-INH/Co-INH</th>
<th>Assignment</th>
</tr>
</thead>
<tbody>
<tr>
<td>—</td>
<td>ν(O-H)ₐwat</td>
</tr>
<tr>
<td>3300 (m), 3209 (m)</td>
<td>ν(NH₂)</td>
</tr>
<tr>
<td>3114 (s)</td>
<td>ν(N-H)</td>
</tr>
<tr>
<td>1667 (vs)</td>
<td>ν(C=O)</td>
</tr>
<tr>
<td>1635 (vs)</td>
<td>ν(C=N)py</td>
</tr>
<tr>
<td>1492 (m)</td>
<td>δ(CNH)</td>
</tr>
<tr>
<td>1334 (s)</td>
<td>ν(C-N)amide</td>
</tr>
<tr>
<td>1142 (s)</td>
<td>ν(N-N)</td>
</tr>
<tr>
<td>888 (m)</td>
<td>δ(C-N-C)py</td>
</tr>
<tr>
<td>504 (m)</td>
<td>ν(C-O)</td>
</tr>
</tbody>
</table>

s = strong, vs = very strong, m = medium, w = weak, sh = shoulder, br = broad.
this band shifted to 1655 cm\(^{-1}\) remarking the copper coordination through this group. Similar shifts were observed in complexes where the metal coordinate through the \(-\text{C}=\text{O}\) group [27, 28]. This behavior permits to propose that INH acts as a bidentate ligand coordinating through the \(\sim\text{NH}_2\) and the \(\sim\text{C}=\text{O}\) groups.

Regarding the \(\nu\)(C-N)\(_{\text{amide}}\) and \(\nu\)(N-N) bands the coordination affects only the first one. A plausible explanation would be that the coordination through the \(\sim\text{C}=\text{O}\) group weakens the \(\text{C}^{-}\text{O}\) bond and consequently reinforces the \(\sim\text{N-H}\) one, shifting the \(\nu\)(C-N)\(_{\text{amide}}\) to a higher frequency.

Upon complex formation, the shifts of pyridine vibrations in the high-frequency region usually are small, whereas those corresponding to bendings are more evident and shifted to higher frequencies [26]. As Table 3 shows, in the high-frequency region the band corresponding to \(\nu\)(C=O) shifts to 1655 cm\(^{-1}\), confirming the coordination through the heterocyclic nitrogen atom. Especially the band at 504 cm\(^{-1}\) assigned as a ring C-C-C bending [29] shifted to 570 cm\(^{-1}\), confirming the coordination through pyridine. Due to the fact that the aromatic pyridine ring is rigid, the coordination through the heterocyclic N atom should be given with another copper ion different than that coordinated with the hydroxamine group, in a dimeric or polymeric structure.

Regarding the metal-ligand bands, only one band was observed at 281 cm\(^{-1}\) and it was assigned as \(\nu\)(Cu-O) taking into account previous work [26, 29].

The inorganic sulfates present only one stretching band usually found in the range 1140-1080 cm\(^{-1}\). Besides, sulfate bonds can bend giving rise to one or two bands in the 680-610 cm\(^{-1}\) range, sharper than the stretching bands [26]. In the case of Cu-INH the bands at 1118 and 610 cm\(^{-1}\) confirm the presence of a sulfate counterion.

Besides, the broad band at 3409 cm\(^{-1}\) confirms the presence of water molecules in the complex.

**Analysis of the Co-INH spectrum**

The Co-INH spectrum shows the shifts of \(\nu\)(NH\(_2\)) and \(\nu\)(NH\(_2\)) to lower frequencies (see Table 3), according to the cobalt coordination through this group [26]. Besides, the \(\nu\)(C=O) shifts to lower frequencies in a similar behavior to that observed in Cu-INH spectrum. The \(\nu\)(N-H) does not change in the complex showing that this group does not participate in the coordination. Taking into account this analysis, we can propose that isoniazid acts as a bidentate ligand.

Unlike the case of the Cu-INH, in the Co-INH spectrum no significant changes in the pyridine vibrations were observed. This would discard the coordination through the heterocyclic nitrogen atom.

On the other hand, only one band was observed at 279 cm\(^{-1}\) and it was assigned as \(\nu\)(Co-O) taking into account previous reports [26, 29].

Besides, the broad band at 3378 cm\(^{-1}\) confirms the presence of water molecules in the complex.

**Electronic spectra**

Copper(II) complexes of lower coordination number can exist in a wide range of stereochemistries, some of them distorted, making it difficult to use electronic spectroscopy for identifying structures [30]. In the case of Cu-INH, the electronic spectrum in a nujol suspension shows one poorly defined broad band at 600 nm almost overlapped by ligand bands. This result agrees with the report of similar complexes with a CuN\(_2\)O\(_2\) chromophore [30].

On the other hand, the Co-INH is a pale pink powder, typically of octahedral complexes [31]. It is known that high spin six coordinate cobalt(II) complexes generally show, in a visible region, a multiple structured band assigned to \(^{4}\text{T}_{1g}(\text{F}) \rightarrow ^{4}\text{T}_{1g}(\text{P})\) [30]. In the reflectance spectrum of Co-INH three bands at 606, 695 and 725 nm, and a shoulder at 668 nm are observed.

**EPR spectra**

The EPR spectrum of a polycrystalline sample of Cu-INH at X-band is shown in Figure 2.

This spectrum shows two main resonances centered around 290 and 325 mT and that are attributed to transitions allowed by the interaction between the magnetic moment of the unpaired electron and the spectrometer magnetic field (Zeeman interaction). Those two resonances are characteristic of an axially symmetric paramagnetic center with principal components of the g-tensor given by \(g_\perp = 2.023\) and \(g_\parallel = 2.236\) as calculated directly from the resonance positions (in units of magnetic field) and from the frequency of microwave radiation used in the experiment. The relation \(g_\parallel > g_\perp\) indicates that the unpaired electron is in a \(d_{2}x^2-y^2\) ground state orbital. Copper ions have nuclear spin of 3/2 that would give rise to extra resonance lines in their spectrum (hyperfine interaction). However, no such lines are observed in Figure 2, which is a situation usually found in the cases where copper - copper interactions are observed [32-36].

![Fig. 2. X-band EPR spectrum of a polycrystalline sample of Cu-INH.](image-url)
It is also worth noting the absence of extra resonances for both low (below 275 mT) and high (above 375 mT) fields and also of the so-called half-field transition, which is typical of copper pairs [37]. Together these features suggest that the copper ions in Cu-INH are arranged as a polymeric structure in the solid state, as shown in Figure 3.

As a result of the experimental analytical data and the spectroscopic studies (IR, UV-Vis and EPR) the proposed structures for the complexes are shown in Figure 3.

**Lipophilicity and solubility**

Before the study of lipophilicity, the stability of complexes in physiological solution was determined. A thin-layer chromatography using the experimental conditions presented in 3.4 was performed. The solutions analyzed were: saturated solution of Cu-INH in physiological serum, saturated solution of Cu-INH in water, saturated solution of Co-INH in physiological serum and saturated solution of Co-INH in water. After the experiment, only one spot was observed in each Co-INH chromatogram and the \( R_f \) were 0.73 in both solutions, showing that the presence of Cl\(^-\) does not affect the species in the physiological solution. The same behavior was observed in the case of Cu-INH but the \( R_f \) was almost 0 according to the low solubility of the complex in methanol.

With the aim to determine the lipophilic character of the complexes that can be correlated with biological properties like the passage through lipophilic barriers, distribution coefficients (D) between n-octanol and physiological solutions were determined. Besides, the solubility in water and DMSO was estimated. These results are presented in Table 4.

The D values obtained for the complexes are higher than that of the free ligand. According to the results shown in Table 4, the compound most soluble in water (isoniazid) was the less lipophilic while both complexes presented lower solubility and were more lipophilic than isoniazid, as expected.

The solubilities in DMSO are higher than the values obtained in aqueous solution. For this reason this solvent was used in the microbiological assays.

**Anti-M. tuberculosis activity and cytotoxicity on mammalian cells**

The minimum inhibitory concentration (MIC) values against *M. tuberculosis*, the IC\(_{50}\) and the selectivity index of the metal-INH complexes and the free ligand are presented in Table 5. None of the metallic salt dilutions (CuSO\(_4\) and CoCl\(_2\)), used as control, inhibited the growth of mycobacterium. In the case of Cu-INH, the molecular weight of the monomeric complex was used to calculate the MIC, due to the uncertainty in knowing the molecular weight in solution. If polymeric structures exist in solution the calculated MIC values expressed in μM will be lower.

As Table 5 shows, both complexes were active against *M. tuberculosis*. The MIC values are lower than that of the ethambutol (MIC = 5.62 μg/mL, 27.5 μM) and p-aminosalicylic acid (MIC = 1.25 μg/mL, 8.16 μM), antimycobacterial agents in clinical use.

Besides, Cu-INH complex (MIC = 2.2 μM) is less active than isoniazid (MIC = 0.063μg/mL, 0.46 μM) while Co-INH (MIC = 0.41 μM) has similar activity but both complexes presented MIC values well below 25 μg/mL, considered as a limit to continue the anti-mycobacterial studies, according to pipeline by Pavan *et al.*[38].

One thing to note is that Co-INH was more active than Cu-INH and more lipophilic. However, isoniazid was less lipophilic than the complexes but it was the most active one. INH is a prodrug which is active after transformation, by a peroxidase, to isonicotinic acid that inhibits the synthesis of the mycolic acid required for the biosynthesis of mycobacterial cell wall [9]. INH enters into the cell by passive transportation [39] [40]. In spite of its polarity this molecule is a very powerful drug. In the case of our complexes, two mechanisms would be possible:

![Fig. 3. Proposed structures for Cu-INH (A) and Co(INH) (B).](image)

**Table 4.** Distribution coefficients (D) between n-octanol and physiological solutions, and estimated solubility (S) of the complexes.

<table>
<thead>
<tr>
<th>Complex</th>
<th>D/ logD*</th>
<th>S (mg/100 mL)</th>
<th>S (mg/100 mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>INH</td>
<td>0.07/−1.2</td>
<td>1667</td>
<td>815</td>
</tr>
<tr>
<td>Cu-INH</td>
<td>0.16/−0.79**</td>
<td>3.0</td>
<td>5.8</td>
</tr>
<tr>
<td>Co-INH</td>
<td>0.33/−0.48**</td>
<td>75</td>
<td>150</td>
</tr>
</tbody>
</table>

* strength force = 0.15 M, 25 °C.
** pH = 4.98 (for Cu-INH) and 4.38 (for Co-INH).

**Table 5.** MIC (mg/mL), IC\(_{50}\) and Selectivity Index for Cu-INH, Co-INH.

<table>
<thead>
<tr>
<th>Compound</th>
<th>MIC* (μg/mL)</th>
<th>MIC* (μM)</th>
<th>IC(_{50}) (μg/mL)</th>
<th>IC(_{50}) (μM)</th>
<th>SI**</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>VERO</td>
<td>J774A.1</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cu-INH</td>
<td>0.78</td>
<td>2.2</td>
<td>62.5/178</td>
<td>58.6/167</td>
<td>80.1</td>
</tr>
<tr>
<td>Co-INH</td>
<td>0.19</td>
<td>0.41</td>
<td>&gt;500/1070</td>
<td>&gt;500/1070</td>
<td>2631.6</td>
</tr>
</tbody>
</table>

* The obtained MIC values were the same in the triplicate assays
** The selectivity index (SI) was calculated by the ratio between VERO/MIC.
the complexes would pass through the cell wall by a different mechanism than isoniazid or they would act through a different mechanism of action. Due to the fact that the mode of action of isoniazid requires the biotransformation to isonicotinic acid, the stable coordination to a metal center would block the whole process leading to an inactivation of isoniazid. In our case, both new metal complexes are active allowing to conclude that the mode of action of the complexes could be completely different to that of free isoniazid. If this hypothesis could be verified through further studies these compounds would be good drug candidates.

Another interesting aspect of these complexes is the cytotoxicity shown on two different mammalian cells lineages. The models chosen were the VERO (ATCC CCL81) and J774.A1 (ATCC TIB-67) cells. VERO cells are considered normal cells and do not represent any disease. These cells are largely used as phenotypic screening in drug discovery in response of the cytotoxicity of the compounds. J774A.1 are macrophage cells, that represent the first ones of the immunological response. Also, mycobacteria has the ability to survive inside them. All the complexes showed moderate cytotoxicity on these cell lines and good selectivity indexes (Table 5).

As a conclusion, the synthesis and characterization of two new isoniazid complexes are reported. According to the spectroscopic analysis the copper complex presents a polymeric structure where the isoniazid acts as a bridge between two different copper ions, coordinating through the heterocyclic nitrogen, the amine and carbonyl groups.

The cobalt complex presents an octahedral geometry where the isoniazid is a bidentate ligand. These two examples show the versatility of isoniazid as ligand.

Both complexes were active against M. tuberculosis showed MIC values well below 25 μg/mL considered as a limit to continue the antimycobacterium studies. Besides, they showed low cytotoxicity on different mammalian cell lines and consequently they presented high selectivity indexes.

Due to these results, Cu-INH and Co-INH are interesting candidates for further studies.

**Experimental**

All starting materials were commercially available research-grade chemicals and were used without further purification.

**Synthesis**

The copper and cobalt complexes were synthesized mixing aqueous solutions of CuSO₄·5H₂O (0.55 mmol, 137 mg) or CoCl₂·6H₂O (0.55 mmol, 130 mg) and isoniazid (1.10 mmol, 150 mg) with continuous stirring for 30 min. The resulting green or pink precipitates corresponding to copper or cobalt complexes respectively were filtered, washed with small portions of water and dried at room temperature.

The elemental analysis was performed with a Carlo Erba EA1108 elemental analyser. Sulphate was identified by precipitation with BaCl₂ and chloride was quantified through a potentiometric method using AgNO₃.

**Thermogravimetric and conductimetric measurements**

Thermogravimetric measurements were performed with a Shimadzu TGA 50 thermobalance, with a platinum cell, working under flowing air (50 mL min⁻¹) and at a heating rate of 0.5 °C min⁻¹ until 80 °C and 1 °C min⁻¹ above 80 °C.

Conductimetric measurements using a Conductivity Meter 4310 Jenway were performed at 25 °C in a DMSO solution (10⁻⁴ M), solvent used in the microbiological assays. The stability of the complexes in this solvent was followed by measuring conductivity of these solutions for 48 h of the dissolution and compared with the initial conductivity.

**Spectroscopic measurements**

IR spectra, in the range between 4000 and 200 cm⁻¹, were recorded on a Bomem M 102 FTIR spectrophotometer using the KBr pellet technique.

Electronic spectra of complexes in a Nujol suspension were registered on a Shimadzu UV-1603 spectrophotometer.

Room temperature EPR measurements of the copper complex were carried out on polycrystalline samples using a JEOL JES-FA200 spectrometer. The experimental parameters were: microwave frequency, 9.4 GHz; microwave power, 10 mW; field modulation,

EPR measurements of the copper complex were carried out on polycrystalline samples using an X-band Varian E109 spectrometer. The experimental parameters were microwave frequency: 9.4 GHz, microwave power: 10 mW, field modulation: 100 kHz, modulation amplitude: 0.2 mT, time constant: 0.03 s, scan time: 2 min. The parameters were optimized to avoid saturation and/or distortions.

**Thin-layer chromatography**

With the aim of studying the stability of complexes in physiological solution and in water, a thin-layer chromatography was performed with a saturated solution of complexes in both solvents. A silica gel G, 250 μm plate and a CH₃OH: NH₃ (100:1.5) solution as a mobile phase were used. 10 μL of each solution (saturated solution of Cu-INH and Co-INH in physiological solution, and saturated solutions of Cu-INH and Co-INH in water) were seeded in the plate. After the chromatography, the spots were detected with UV lamp [41].

**Lipophilicity test and solubility**

Lipophilicity tests at 25 °C were performed determining the partition coefficient (P) between n-octanol and physiological solution [42]. The concentration of the compounds in the physiological solution before and after the contact with n-octanol was determined by UV spectroscopy at 260 nm. This isoniazid band does not shift with complexation.
The solubility at 37 °C in water and DMSO was also estimated.

Anti-M. tuberculosis activity

The anti-MTB activity of the compounds was determined by the REMA (Resazurin Microtiter Assay) method according to Palomino et al., 2002 [43]. Stock solutions of the tested compounds were prepared in dimethyl sulfoxide (DMSO) and diluted in Middlebrook 7H9 broth (Difco) supplemented with oleic acid, albumin, dextrose and catalase (OADC enrichment - BBL/Becton-Dickinson), to obtain final drug concentration ranges of 0.09-25 µg/mL. The isoniazid was dissolved in distilled water, and used as standard drug. A suspension of the MTB H37Rv ATCC 27294 was cultured in Middlebrook 7H9 broth supplemented with OADC and 0.05% Tween 80. The culture was frozen at -80 °C in aliquots. After two days was carried out the CFU/mL of a aliquot. The concentration was adjusted by 5x10^6 CFU/mL and 100 µL of the inoculum was added to each well of a 96-well microplate together with 100 µL of the compounds. Samples were set up in triplicate. The plate was incubated at same conditions for 24 h, to allow cell adhesion. The compounds were dissolved in DMSO (5%) and subjected to two-fold serial dilution from 1250 to 3.9 µg/mL. Cells were exposed to the compounds at concentrations of 0.09-25 µg/mL, at 37 °C, in a humidified 5% CO₂ atmosphere. After reaching confluence, the cells were detached and counted. For the cytotoxicity assay, 1 x 10^5 cells/mL were seeded in 200 µL of complete medium in 96-well plates (NUNC™). The plates were incubated at same conditions for 24 h, to allow cell adhesion prior to drug testing. The compounds were dissolved in DMSO (5%) and subjected to two-fold serial dilution from 1250 to 3.9 µg/mL. Cells were exposed to the compounds at various concentrations for 24 h. Resazurin solution was then added to the cell cultures and incubated for 6 h. Cell respiration, as an indicator of cell viability was detected by reduction of resazurin to resorufin, whose pink colour and fluorescence indicates cell viability. A persistent blue colour of resazurin is a sign of cell death. The fluorescence measurements (530 nm excitation filter and 590 nm emission filter) were performed in a Tecan Spectrafluor Plus microfluorimeter. The IC₅₀ value was defined as the highest drug concentration at which 50% of the cells are viable relative to the control. Each test was set up in triplicate.

Cytotoxicity on mammalian cells

In vitro cytotoxicity assays (IC₅₀) were performed first on the VERO epithelial cells (ATCC CCL81). Compounds with low cytotoxicity were investigated on the J774A.1 (ATCC TIB-67) mouse cell line. Both studies as recommended by Pavan et al., 2010 [44]. The cells were routinely maintained in complete medium (DMEM for VERO and RPMI-1640 (VitroCell®) for J774A.1) supplemented with 10% heat-inactivated fetal bovine serum (FBS) plus gentamicin (50 mg/L) and anfotericin B (2 µg/mL), at 37 °C, in a humidified 5% CO₂ atmosphere. After reaching confluence, the cells were detached and counted. For the cytotoxicity assay, 1 x 10⁵ cells/mL were seeded in 200 µL of complete medium in 96-well plates (NUNC™). The plates were incubated at same conditions for 24 h, to allow cell adhesion prior to drug testing. The compounds were dissolved in DMSO (5%) and subjected to two-fold serial dilution from 1250 to 3.9 µg/mL. Cells were exposed to the compounds at various concentrations for 24 h. Resazurin solution was then added to the cell cultures and incubated for 24 h. The fluorescence measurements (530 nm excitation filter and 590 nm emission filter) were performed in a Tecan Spectrafluor Plus microfluorimeter. The IC₅₀ value was defined as the highest drug concentration at which 50% of the cells are viable relative to the control. Each test was set up in triplicate.

Selectivity Index

The selectivity index (SI) was calculated by dividing IC₅₀ for the VERO cells by the MIC for the pathogen; if the SI is ≥10, the compound is then investigated further.

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