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## Exploring the prospect of marine microalgae *Isochrysis galbana* as sustainable solid biofuel feedstock

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**Abstract:** Among the renewable biomass fuel alternatives, microalgae are the most important future choices owing to its fast growth rate and great capability for CO<sub>2</sub> fixation. There are various species in the world, in which each species has its characteristics. This work presents a prospect of marine microalgae *Isochrysis galbana* for renewable fuel feedstock regarding its biomass abundance, physicochemical properties, and thermal characteristic. The seawater medium in the Erlenmeyer flask was used for the algal culturing. The biomass abundance, in term of specific growth rate and doubling time, was assessed by calculating the culture medium cells number with a hemocytometer and optical microscope. Harvesting was done by precipitating biomass with caustic soda, subsequently filtering, and washing it with distilled water. The biomass sediment had been sun-dried for three days, and then dried biomass was crushed by using the mortar to be a powder. The proximate analysis was arranged by conducting an experiment in according to the test method of ASTM D 3173-11, ASTM D 3175-11, ASTM D 3172-13 and ASTM D 3174-12 for specifying the content of moisture, volatile matter, fixed carbon, and ash of the sample, respectively. The heating value was estimated by using adiabatic bomb calorimeter. The chemical composition of biomass was determined by Energy-dispersive X-ray (EDX) spectrometry. The biomass cellular macromolecular compounds were also evaluated by Fourier transform infrared (FTIR) spectroscopy and compared with its residue. Through eight days observation, it was noticeable that *Isochrysis galbana* has a specific growth rate of 0.18 d<sup>-1</sup> and a doubling time of 3.85 d. The respective moisture, volatile matter, fixed carbon, and ash content were 12.98, 40.10, 7.47, and 39.45 (% air-dried basis). The energy content algal biomass was 16.22 MJ kg<sup>-1</sup>. This current investigation encourages that *Isochrysis galbana* can be viable as one of a future sustainable solid biofuel feedstock.

**Keywords:** Sustainable, fuel feedstock, biomass, marine microalgae, *Isochrysis galbana*

## 1. INTRODUCTION

Limited supplies of petroleum-based fuel and a massive impact on atmospheric CO<sub>2</sub> levels as a result of

its burning processes have encouraged intense interest to a renewable and environmentally-friendly fuel.

Discovering adequate supplies of renewable and clean fuel for the future survival is the most daunting challenges since they significantly determine economic prosperity, global stability, and quality of life. This phenomenon induces appealing debate and questions over new fuel choices, resulted from new raw materials, to partly

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complement or entirely substitute present petroleum-based fuels. Biomass has recently turned out to be a topic of passionate interest.

Yet, determining to which biomass with their yield promises for biofuel candidates appears as most gorgeous challenges. The use of first-generation biofuels, which have focused on higher plants such as sugarcane, corn, oil palm, soybean, algae, and others, raises new issues associated with the global food security and food markets or the loss of ecosystems. Second-generation biofuels resulted from lignocellulosic biomass such as forest and agriculture residues, as well as nonfood vegetable feedstock, can address some of the above problems; nevertheless, there is an alarm over competing land use or required land use changes (Brennan & Owende, 2010). While most bioenergy alternatives miss opportunities as replacement fossil fuels due to various constraints, aquatic microorganism-based biofuels have a good prospect to generate a significant amount of renewable energy without disruptions. Based on current knowledge and technology projections, microalgae are taken into account to be third-generation biofuels which are technically feasible as the alternative energy reserves, and they are without major drawbacks compared to the first and second generation biofuels (Brennan & Owende, 2010).

Microalgae have become one of the most important renewable energy sources owing to their flexible cultivation and neutrality towards the natural environment. Some species of microalgae can be cultured in brackish water on non-arable land (Brennan & Owende, 2010; Schwenk et al., 2013), and consequently, they may not bring in land-use change. Cultivation of microalgae could be conducted under non-controlled conditions, open bioreactors (Da Rosa, Carvalho, Goldbeck, & Costa, 2011), as well as open ponds (Crowe et al., 2012; Sanchez, González, Maceiras, Cancela, & Urrejola, 2011), and microalgae use sunlight as the primary source of energy. Therefore, a non-controlled cultivation is easy to build and operate with low initial investment (Da Rosa et al., 2011). Producing and harvesting microalgae have uses in wastewater treatment and CO<sub>2</sub> fixation (Christenson & Sims, 2011; Wang, Li, Wu, & Lan, 2008), and by naturally fixing atmospheric CO<sub>2</sub> via photosynthesis they are ten times more efficient than terrestrial plants (Cantrell, Ducey, Ro, & Hunt, 2008). Hence, it would reduce environmental impacts. Microalgal biomass is arranged by 50% carbon by dry weight, and its element is originated from carbon dioxide

(Sánchez Mirón et al., 2003). Producing 1 kg of algal biomass requires 1,83 kg of CO<sub>2</sub> (Yusuf, 2007); thereby, microalgae cultivation is critical for global warming mitigation (Sukarni, Sudjito, Hamidi, Yanuhar, & Wardana, 2015). Moreover, algae cultivation does not undermine the production of food, fodder and other products derived from crops (Yusuf, 2007). Microalgae can be a year-round harvest (Phukan, Chutia, Konwar, & Kataki, 2011) providing a reliable and continuous supply of biomass. They have higher annual biomass productivity on an area basis due to their fast growth rate and high biomass yield (Sukarni, Sudjito, Hamidi, Yanuhar, & Wardana, 2014). Thus, they will be prospective as essential future energy reserves to address the fossil fuels problems.

Exploring the feasibility of microalgae for future biomass fuel feedstock obligates to investigate their biomass abundance as well as their physicochemical properties. The algal biomass productivity will ensure their continuous supply of biomass fuel feedstock. The algal physicochemical properties are also critical parameters because they will provide a strong effect on the combustion characteristics. The latest references associated with the microalgal biomass fuel specify that very few studies have been focused on the microalgal biomass characterization. Phukan, Chutia, Konwar, and Kataki (2011) have investigated the potential bio-energy feedstock of *Chlorella sp. MP-1* regarding its physical and chemical characteristics by using bomb calorimeter, thermogravimetric analysis (TGA), CHN analyzer and Fourier transform infrared (FTIR) spectroscopy. Sukarni, et al. (2014) have studied the potential of marine microalgae *Nannochloropsis oculata* concerning its abundance and physicochemical properties. However, the potential of *Isochrysis galbana* as a biomass fuel feedstock has not been explored yet. In connection with the broadening opportunities for seeking alternative energy sources, exploring the potential of *Isochrysis galbana* as solid fuel feedstock is very critical. Hence, for optimizing the usefulness of this biomass associated with the propriety of combustion technology and the potential future applications, it requires further in-depth research.

This work endeavored to fill partially in the gap, by investigating the properties of *Isochrysis galbana* in accordance with its prospect as biomass fuel feedstock. The samples were characterized in relation to their biomass growth rate, heating value, moisture and ash

content, as well as their chemical composition. The cellular macromolecular content of biomass and residue have also been evaluated by using FTIR spectroscopy. The study also analyzed the biomass thermal characteristic under a nitrogen and air atmosphere.

## 2. MATERIALS AND METHODS

### 2.1 SPECIES CULTIVATION AND GROWTH EVALUATION

The marine microalga *Isochrysis galbana* species was obtained from the Center for Development of Brackish Water Aquaculture (Balai Besar Pengembangan Budidaya Air Payau-BBPBAP), Jepara, Indonesia. Eight Erlenmeyer flasks with seawater medium were used to cultivate the species. During the cultivation process, three-time measurement was performed for salinity, pH, temperature, and dissolved oxygen of the medium, i.e. at the beginning, middle and end of the period. The fertilizer, whose composition was shown in Table 1, enriched the cultivation medium. The species' culturing duration was eight days.

The daily growth evaluation was conducted during the cultivation period by calculating the number of cells in the cultivation medium using hemocytometer coupled with an optical microscope (CH2 Olympus Optical Co. Ltd, Tokyo, Japan). The sample calculation was done every day at 10:00 AM for three-time measurement and then averaged. The specific growth rate ( $\mu$  in unit  $d^{-1}$ ) of the biomass was specified according to Sukarni, et al. (2014) and calculated as following:

$$\mu = d \ln C_n / dt = (\ln C_t - \ln C_o) / (t - t_o) \quad (1)$$

Where  $C_n$  is the cell number density (cells  $ml^{-1}$ ),  $t$  is time (d),  $C_t$  is cell density at time ( $t$ ), and  $C_o$  is cell density at the start of the exponential phase ( $t_o$ ). The doubling time (d), i.e. the time required to double the cell, is determined as the following:

$$t_d = \ln 2 / \mu \quad (2)$$

### 2.2 HARVESTING, DRYING AND PULVERIZING

The biomass' harvesting process has been carried out by precipitating the biomass suspension with caustic soda

and then filtering and washing it twice with distilled water. The precipitated materials were subsequently sun-dried for three days. The fine particles of biomass were attained by crushing the biomass chunks with a mortar. A vacuum desiccator was employed to store the pulverized samples.

### 2.3 PROXIMATE ANALYSIS AND HIGHER HEATING VALUE (HHV) CALCULATION

The proximate analysis, which determines contents of moisture, volatile matter, fixed carbon and ash in the biomass samples, was performed by an experiment in according to the test method of ASTM D 3173-11, ASTM D 3175-11, ASTM D 3172-13 and ASTM D 3174-12, respectively. The adiabatic bomb calorimeter was employed to determine the heating value of biomass. The sample about 0.5 g was used for 10 min with 99.5% pure oxygen.

### 2.4 CHEMICAL COMPOSITION ANALYSIS

The chemical composition of the algal biomass was determined by an Energy-dispersive X-ray (EDX) spectrometry (FEI, Inspect-S50) coupled with X-ray microanalysis capabilities (AMETEK EDAX TSL). Minimizing the surface electron charging effect of samples was achieved by gold-coating so that the distort images did not take place during the samples' evaluation.

The macromolecular content of biomass, which represented the presence of carbohydrate, protein, and lipid in the algal biomass, was evaluated by using Shimadzu FTIR spectroscopy. Before measuring process, the potassium bromide (KBr) powders were mixed with the samples and then mixed samples were pressed into tablets. The scanning of samples was conducted from 400 to 4000  $cm^{-1}$ . Likewise, investigation of macromolecular content change after the thermal processes of biomass had performed by the similar method.

### 2.5 THERMAL ANALYSIS

A METTLER TOLEDO TGA/DSC1 simultaneous analyzer was employed in the current study to continuously monitor mass changes of *Isochrysis galbana* sample as a result of thermal degradation in the nitrogen and air environment, respectively. The sample followed a

fixed linear heating program of  $10\text{ }^{\circ}\text{C min}^{-1}$ . The small samples (approximately 10 mg) were loaded into an  $\text{Al}_2\text{O}_3$  ceramic crucible and then inserting into the reactor, which subsequently was heated up from ambient temperature to  $1000\text{ }^{\circ}\text{C}$ . During the process of pyrolysis and combustion, the TGA furnace was flowed by nitrogen as an inert and air atmosphere as an oxidizer, where each test was supplied with a constant flow rate of  $100\text{ ml min}^{-1}$ . The continuous on-line records of mass loss as a function of temperature and time were obtained to plot the thermogravimetric analysis (TGA) curve as well as the derivative thermogravimetric (DTG) one.

Table 1. The Walne fertilizer composition.

Substances	Composition (ppm)
$\text{NaNO}_3$	100
$\text{NaH}_2\text{PO}_4$	20
$\text{H}_3\text{BO}_3$	33.6
$\text{NaEDTA}$	45
$\text{FeCl}_3$	1.3
$\text{MnCl}_2$	0.36
$\text{Na}_2\text{SiO}_3$	10-80
<b>Vitamin B12</b>	0.001

### 3. RESULTS AND DISCUSSION

#### 3.1 THE CULTURE CONDITIONS AND BIOMASS PRODUCTIVITY

The appropriateness of microalgae for biomass fuel candidates, in terms of the sustainability, is largely determined by the biomass abundance. Hence, microalgae productivity is one of the most important parameters which must be examined. In this work, the *Isochrysis galbana* culture condition was shown in Figure 1. Its environmental parameters, represented by the salinity, pH, temperature and dissolved oxygen (DO) of the medium, were depicted in Table 2. The microalgae are very sensitive to their growth environment. Hence, these environmental parameters are interesting in relation to both biomass productivity and properties.

The *Isochrysis galbana* biomass abundance during the eight days cultivation period was shown in Figure 2.

It was started at around  $3 \times 10^6$  cells  $\text{ml}^{-1}$  under the initial cultivation, and then the density reached  $1044 \times 10^6$  cells  $\text{ml}^{-1}$  under the end one. In terms of growth kinetic, *Isochrysis galbana* had  $0.18\text{ d}^{-1}$  for the specific growth rate and  $3.85\text{ d}$  for the doubling time. It meant that  $0.26$  cell was double, or division per day and average increases in biomass were  $9.3 \times 10^5$  cells  $\text{d}^{-1}$ . It was clearly that *Isochrysis galbana* had good biomass productivity.

The productivity of *Isochrysis galbana* compared with other species cultured under different conditions and media was presented in Table 3. In the growth kinetics point of view, it could be observed that *Isochrysis galbana* had higher productivity than *Chlorella vulgaris* and of the *Nannochloropsis oculata* did, in which both were grown in the Bold's Basal medium and the Guillard f2 one (Converti, Casazza, Ortiz, Perego, & Del Borghi, 2009), respectively. Additionally, the *Isochrysis galbana* productivity equals *Dunaliella salina* cultured in outdoors-closed tubular photobioreactor (García-González, Moreno, Manzano, Florencio, & Guerrero, 2005) and almost equals *Nannochloropsis gaditana* cultivated in the indoor system (Selvakumar & Umadevi, 2014). Table 3 also indicated that *Isochrysis galbana* productivity is less than *Chlorella protothecoides* (Illman, Scragg, & Shales, 2000), *Nannochloropsis oculata* (Sukarni et al., 2014) and *Scenedesmus obliquus* (De Moraes & Costa, 2007), in which all three species were cultured in a low nitrogen Guillards marine medium, a traditionally natural open pond, and tubular photobioreactor, respectively. Due to the sensitivities of microalgae to the environmental culturing (Alkhamis & Qin, 2013; Kaplan, Cohen, & Abeliovich, 1986), its comparison could not exactly show an algal biomass productivities benchmark, but at least could provide an overview for future algae cultivation. Therefore, *Isochrysis galbana* productivity may still be improved by the environmental conditioning, with regard to the culture medium and nutrients engineering.



Fig. 1. The culture condition of *Isochrysis galbana* species.

Table. 2. The environmental parameters of culture medium. The error represented the standard deviations from eight measurements of eight samples.

Cultivation periods (d)	Salinity (‰)	pH	Temperature (°C)	DO (mg l <sup>-1</sup> )
1 <sup>st</sup>	25.25 ± 1.28	8.56 ± 0.50	24.13 ± 0.48	7.24 ± 0.26
4 <sup>th</sup>	38.75 ± 1.98	8.69 ± 0.46	23.88 ± 0.42	7.33 ± 0.15
8 <sup>th</sup>	40.38 ± 1.60	9.00 ± 0.00	24.31 ± 0.27	7.53 ± 0.11

Table. 3. The kinetic growth rate comparison of various microalgae species.

Species	Cultivation	$\mu$ (d <sup>-1</sup> )	$t_d$ (d)	Ref
<i>Dunaliella salina</i>	Outdoors -closed tubular photobioreactor	0.18	3.85	(1)
<i>Scenedesmus obliquus</i>	Tubular photobioreactor	0.22	3.15	(2)
<i>Chlorella vulgaris</i>	Bold's Basal medium	0.14	4.95	(3)
<i>Nannochloropsis oculata</i>	The Guillard f2 medium	0.07	9.90	(3)
<i>Nannochloropsis gaditana</i>	Indoor Culture	0.199	3.48	(4)
<i>Chlorella protothecoides</i>	Low nitrogen Guillards Marine medium	0.27	2.6	(5)
<i>Nannochloropsis oculata</i>	Open pond	0.27	2.6	(6)
<i>Isochrysis galbana</i>	Erlenmeyer flask Indoor Culture	0.18	3.85	This study

(1) García-González et al. (2005); (2) De Morais & Costa (2007); (3) Converti et al. (2009); (4) Selvakumar & Umadevi (2014); (5) Illman et al. (2000); (6) Sukarni et al. (2014)

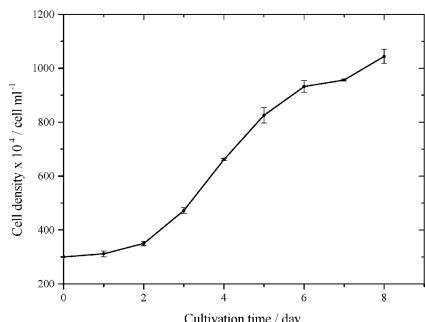


Fig. 2. The biomass abundance of *Isochrysis galbana* during eight days cultivation. The error denoted the relative standard error of mean from three replications of each sample.

### 3.2 THE PHYSICOCHEMICAL PROPERTIES

The main physical properties of solid fuels were the moisture content, volatile matter, fixed carbon, and ash, where overall parameters are ordinary known as proximate values. Moreover, the most critical parameter in relation with the biomass fuel was the heating value. The *Isochrysis galbana* proximate results, as well as the heating value and their comparison with another biomass, were summarized in Table 4.

Table 4 indicated that *Isochrysis galbana* had nearly 12.98% of moisture content. This moisture would be released in the first stage of the thermal conversion process. As shown in Table 4, the moisture content of biomass varied widely, depending largely on the type of biomass. Moreover, the biomass moisture content was also strongly influenced by the drying process and storage mode. Comparing sun-dried *Isochrysis galbana* to another biomass, it clearly that it had a relatively higher moisture content. This result pointed out that *Isochrysis galbana* which sun-dried for three days appears less dry as the fuel feedstock. Therefore, the longer sun-dried process or other drying methods may be needed for reducing this moisture content.

The moisture content was the critical parameter of solid fuel owing to its impact on the overall performance of combustion processes. Because releasing moisture at the initial step of combustion required an amount of energy, biomass with higher moisture content would take more energy for drying out, and subsequently for heating their vapor. It would reduce the maximum potential energy resulted during the biomass combustion processes and their overall efficiency, as well. As the moisture release needs a quantity of energy in the initial step of combustion, it also determines the evaporation time, so that higher moisture content implies more residence time in the chamber. The increasing residence time of biomass in the chamber before the occurrence of gasification and combustion, this leads to required larger equipment during combustion processes and consequently affected the size of the entire combustion system. Moreover, moisture in biomass also affected both burning reaction rate and reaction zone thickness which directly determined the forming of gas emission and influenced the devolatilization rate, respectively.

After releasing moisture during the thermal conversion process, as a result of the increasing temperature, a most organic element of the biomass underwent softening, swelling, and subsequently thermal cracking. It led to the vaporized material known as volatile matter. This volatile matter would determine both initial ignition and burnout time in the first step of combustion processes. Finding out volatile matter amount was needed to determine the quantity accuracy of supplied air to the combustion zone where volatiles was released. The sufficiency of air supplied to this zone affected the combustion efficiency. As shown in Table 4, the volatile

content of *Isochrysis galbana* was 46.08% on the dry weight basis. This volatile content was lower than that of both the terrestrial plant and the other aquatic biomass. This lower volatile content implied on its combustion reactivity. Owing to volatile matter was the most flammable components of biomass, the lower volatile meant harder biomass to be burned or less in reactivity. Conversely, the higher volatile in the fuel, the easier the ignition would be. Therefore, *Isochrysis galbana* with less volatile matter content was more difficult to be burned than other tabulated biomass.

Non-combined carbon, generally known as fixed carbon, is the portion left over once the volatile matter is entirely released, apart from ash and moisture. That carbon was burned to form char. The portion of volatile matter and fixed carbon in the biomass, which expressed as VM/FC ratio, is the critical parameter determining the reactivity of biomass. The higher this ratio was, the easier the biomass was ignited. Conversely, lower ratio made the biomass ignition more difficult. Table 4 showed that the fixed carbon content of *Isochrysis galbana* was 8.58% on dry weight basis (7.47 %, adb). Its content was higher than that of bagasse and *Chlorella vulgaris* (Agrawal & Chakraborty, 2013), and comparable to *Enteromorpha clathrata* and *Nannochloropsis oculata* biomass. It meant that *Isochrysis galbana* was less in reactivity and had higher residence stage until the combustion was completed compared to both of bagasse and *Chlorella vulgaris*. Nevertheless, its fixed carbon content significantly determined the heating value, and it impacted on the heat generation during the combustion processes.

The remaining mass left after the complete combustion of char was ash. It contained inorganic materials, which were incombustible parts of the fuel. These materials were derived from a mineral fraction of the biomass origin. The ash content of biomass fuel was critically associated with the appropriateness of combustion and gas-cleaning technology choices. Furthermore, the residual handling mode and overall operating costs of the combustion system were greatly determined by the ash content of the fuel. Hence, biomass fuels with low ash content were desirable. Table 4 indicated that *Isochrysis galbana*, whose ash content was 45.33% on dry weight basis (39.45%, adb), was higher ash than other biomass feedstocks except for the *Chlorella vulgaris* (Agrawal & Chakraborty, 2013). Because the inorganic element of algal biomass composed ash content,

which was mostly influenced by their environmental culturing, the same algae species might vary in their ash content. This phenomenon could be observed in the case of *Chlorella vulgaris* as shown in Table 4. Therefore, the ash content of *Isochrysis galbana* may achieve lower levels by the appropriate culture condition. A more detailed insight into ash content of *Isochrysis galbana* in relation to its culturing condition is necessary to be observed. This phenomenon will be revealed in future studies.

The higher heating value of *Isochrysis galbana*, which denoted the maximum quantity of energy possibly recoverable from a given its biomass, was comparable to both *Cotton residue* and *Nannochloropsis oculata* (Table 4). Moreover, this value was higher than bagasse. This value provided the strengthening of the *Isochrysis galbana* to be prospective feedstock of solid fuel.

The elemental composition of *Isochrysis galbana*, compared with *Nannochloropsis oculata* and *Nannochloropsis sp.*, as well as with both *Cystoseira tamariscifolia* and *Bifurcaria bifurcata*, was shown in Figure 3. The carbon and oxygen content of *Isochrysis galbana*, which was expressed in (wt.%) were  $(14.43 \pm 3.25)$  and  $(45.68 \pm 1.11)$ , respectively. The error indicated the relative standard error of mean from three replication measurements of the sample. These compositions would strongly influence the biomass combustion behavior and the residual characteristics. As a result of the thermal attack on the organically bounds of substance in the biomass, most of the oxygen would be released and then supplied partially oxygen which was required for the combustion processes. In the completed combustion mode, carbon would be burned and accompanied by releasing the amount of heat to form  $\text{CO}_2$  subsequently.

The inorganic elements presented in *Isochrysis galbana* comprised: (1) major ( $>1\%$ ); (2) minor ( $1-0.1\%$ ); and (3) trace ( $<0.1\%$ ) elements; in relation to the elemental concentrations. Its major elements and their approximate composition in (wt.%) were Mg ( $27.98 \pm 1.57$ ), Ca ( $5.87 \pm 3.97$ ), Si ( $1.67 \pm 0.40$ ), Cl ( $1.66 \pm 0.52$ ) and Na ( $1.10 \pm 0.26$ ). The minor elements were P ( $0.88 \pm 0.45$ ) and S ( $0.65 \pm 0.35$ ) while the only one trace element was found, namely Al ( $0.07 \pm 0.07$ ). It was already mentioned that the error indicated the same measurement condition of each sample with both carbon and oxygen.

The inorganic elements composition of algal biomass depended on many factors including the type of species, growing processes and conditions, the age of biomass

culturing, fertilizer, harvesting time, collection technique, transport and storage mode, pollution, and processing. These characteristics are more changeable and significantly different in comparison among algal biomass depending on aforementioned factors. Figure 3 shows the differences of inorganic elements composition of *Isochrysis galbana* (eight-day cultivation in Erlenmeyer flask), *Nannochloropsis oculata* (seventh-day cultivation in the traditionally open pond) and *Nannochloropsis sp.* (unknown for its culturing condition). Two different species of brown macroalgae, *Cystoseira tamariscifolia* and *Bifurcaria bifurcata* that both originated from the same growing environment, season and harvesting technique (Ainane, Abourriche, & Kabbaj, 2015), also had a different composition. Hence, Figure 3 was asserted that the elemental variations among the three species of microalgae, as well as the difference between both species of macroalgae, were depending on various factors, included the culture conditions and type of species. Thereby identify the details of the elemental composition of each raw material, which originated from different species and culture medium, was very critical because it would highly impact to its characteristics in the furnace.

During the combustion processes, inorganic elements could significantly influence the ash melting and deposit formation, aerosol and fly ash emissions as well as the material corrosion and the byproduct utilization of ashes. The respective highest inorganic elements contents of *Isochrysis galbana* biomass, i.e., alkaline earth metal (Mg and Ca), would frequently increase the ash melting point. The presence of Al in the form of its oxide might also potentially increase the ash melting point, as well. Conversely, the attendance of chlorides and the low melting alkali such as sodium might considerably decrease the ash melting point. Besides chlorides and sodium, the existence of phosphorus could also greatly reduce the melting point of ash (Du, Yang, Qian, Wang, & Chen, 2014; Obernberger, Brunner, & Barnthaler, 2006).

A lot of alkali metals and chlorides was vaporized during combustion processes to form gasses such as  $\text{HCl}$  (g),  $\text{Na}_2\text{SO}_4$ (g) and  $\text{NaCl}$  (g). Since the heat exchange section of boilers cooled down, they condensed and formed a large amount of fine ash portion. The integration of Cl in the ash was influenced primarily by the concentration of alkali and earth-alkali metals as well as Si in the fuel, which they could react with Cl. The main effect of Cl in the biomass was the corrosive effect of chloride salts.

The sulfur (S) presented in the biomass forms mostly gaseous  $\text{SO}_2$ , partly also  $\text{SO}_3$ , and both alkali and earth-alkali sulfates. Owing to the succeeding cooling of flue gas in the boiler section of combustion plant,  $\text{SOx}$  could form sulfates and condensed on the heat exchanger surfaces, or formed fine fly ash particles, or reacted directly with fly ash particles deposited on heat exchanger surfaces by sulphation reaction. The remaining S rests in the flue gas as aerosols and in gaseous formed as  $\text{SO}_2$  (in insignificant amounts as  $\text{SO}_3$ ) (Obernberger et al., 2006). Overall, in-depth knowledge of the inorganic elements composition of algal biomass is critical regarding the technological countermeasures which include proper temperature control on the grate and in the furnace as well as in the boiler section.

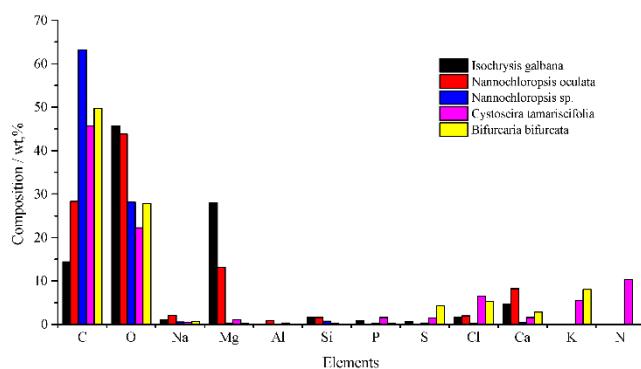


Fig. 3. The EDX elemental comparison of microalgae (*Isochrysis galbana*, *Nannochloropsis oculata* (Sukarni et al., 2014), and *Nannochloropsis* sp. (Patil et al., 2011) and brown macroalgae (*Cystoseira tamariscifolia* and *Bifurcaria bifurcata* (Ainane et al., 2015)

### 3.3 THERMAL CHARACTERISTIC

Figure 4 shows the mass loss and the mass loss rate curve obtained during the thermal processes of *Isochrysis galbana* sample at a heating rate of  $10\text{ }^{\circ}\text{C min}^{-1}$  under nitrogen and air atmosphere, respectively. The both graph has the similar trend, indicated the thermal degradation of the sample was less affected by the kind of the flowing gas. Nevertheless, the presence of  $\text{N}_2$  has caused the increasing in material degradation at high temperature as compared to that of  $\text{O}_2$ . Figure 4 also signified that onset volatile degradation temperature remains same, but the burnout temperature has been shifted toward higher temperature because of the presence of  $\text{O}_2$ .

Figure 4 revealed that three prominent peaks could be encountered during the thermal processes of the samples. The first stage, which taken place from ambient temperature to nearby  $180\text{ }^{\circ}\text{C}$ , was the mass loss relates to the release of physically adsorbed water and low molecular weight hydrocarbons. The second stage which characterized by a strong peak started at around  $330\text{ }^{\circ}\text{C}$  and finished at about  $430\text{ }^{\circ}\text{C}$  attributed to the main pyrolysis and combustion stage. The third stage was the zone where the solid residue of highly non-volatile carbon compounds resulted from stage two gasified and combusted at high temperature.

### 3.4 FTIR ANALYSIS OF ISOCHRYYSIS GALBANA BIOMASS

The FTIR spectroscopy provides information on the chemical structure of *Isochrysis galbana* biomass by identifying the peaks of its spectra, which are produced by the vibrations of functional groups. The FTIR spectra of *Isochrysis galbana* biomass and its residue  $1000\text{ }^{\circ}\text{C}$  were depicted in Figure 5. The band spectra in Figure 5 indicated that *Isochrysis galbana* biomass contains a number of atomic groups and structures. Band intensities reveal the chemical bonds in *Isochrysis galbana* biomass which is comprised into three groups: (1) The “carbohydrate band spectra” (2) The “protein band spectra” and (3) the “lipid band spectra”; in relation to the distinct transmittance in different frequency regions.

Peaks in the region where they are supposed to be carbohydrate belonging are referred to the “carbohydrate band spectra.” These spectra were distinguished by the weak band at  $856\text{ cm}^{-1}$  which indicated the  $\alpha$ -D glucans and C–H aromatic deformation of polysaccharides of the cell wall. Likewise, this weak band pointed out the Si–OH bending of silanol (Corradi da Silva et al., 2008; Huang, Jin, Zhang, Cheung, & Kennedy, 2007; Lecellier et al., 2014; Marshall, Javaux, Knoll, & Walter, 2005; Parida, Dash, Patel, & Mishra, 2006; Peng, Zhang, Zeng, & Kennedy, 2005; Peng, Zhang, Zeng, & Xu, 2003; Tianqi, Hanxiang, Manyi, & Tianwei, 2007). The prominent feature at  $1010\text{ cm}^{-1}$  contributing to C–O and C–O–C stretching of polysaccharides, Si–O stretching of siloxanes silicate frustules, as well as its features indicated the Si–OH bond stretching of silanes (Banyay, Sarkar, & Gräslund, 2003; Benning, Phoenix, Yee, & Konhauser,

2004; Benning, Phoenix, Yee, & Tobin, 2004; Dean, Nicholson, & Sigee, 2008; Dean, Sigee, Estrada, & Pittman, 2010; El-Toni et al., 2012; Goo et al., 2013; Guo et al., 2013; Jiang, Yoshida, & Quigg, 2012; Mayers, Flynn, & Shields, 2013; Meng, Yao, Xue, & Yang, 2014; Parida et al., 2006; Stehfest, Toepel, & Wilhelm, 2005).

The “protein band spectra” was shown by the medium peak at  $1255\text{ cm}^{-1}$  associated with asymmetric stretching of nucleic acids phosphodiester (P=O) (Banyay et al., 2003; Benning, Phoenix, Yee, & Konhauser, 2004; Benning, Phoenix, Yee, & Tobin, 2004; Dean et al., 2008, 2010; Duygu et al., 2012; Jangir, Charak, Mehrotra, & Kundu, 2011; Mayers et al., 2013; Mecozzi, Pietroletti, & Di Mento, 2007; Murdock & Wetzel, 2009; Sigee, Dean, Levado, & Tobin, 2002). The most prominent spectrum at  $1454\text{ cm}^{-1}$  originated from the  $\text{CH}_2$  and  $\text{CH}_3$  bending of methyl and this feature also specified the C–O stretching of the carboxylic group (Banyay et al., 2003; Benning, Phoenix, Yee, & Konhauser, 2004; Benning, Phoenix, Yee, & Tobin, 2004; Dean et al., 2008; Jiang et al., 2012; Murdock & Wetzel, 2009; Sigee et al., 2002; Stehfest et al., 2005). The peak in the spectrum originating from C=O stretching of esters was the medium band at  $1651\text{ cm}^{-1}$ , which was associated with the unique protein amide I characteristic (Benning, Phoenix, Yee, & Konhauser, 2004; Benning, Phoenix, Yee, & Tobin, 2004; Dean et al., 2008, 2010; Gao, Yang, & Wang, 2013; Jiang et al., 2012; Kamnev, Ristić, Antonyuka, Chernyshev, & Ignatov, 1997; Mayers et al., 2013; Murdock & Wetzel, 2009; Phukan et al., 2011; Ragusa et al., 2002; Sigee et al., 2002; Stehfest et al., 2005; Wysokowski et al., 2013). The small feature at  $1786\text{ cm}^{-1}$  as a result of C=O stretching of esters (Gao et al., 2013; Jiang et al., 2012; Mayers et al., 2013; Mecozzi et al., 2007; Murdock & Wetzel, 2009; Phukan et al., 2011; Stehfest et al., 2005; Tan, Balasubramanian, Das, Obbard, & Chew, 2013), and less pronounced band at  $2520\text{ cm}^{-1}$  owing to superimposed O–H and amine salts  $-\text{NH}_3^+$  stretching (Silverstein, Webster, & Kiemle, 2005) were also invented to be protein band spectra. The broad spectrum at around  $3296\text{ cm}^{-1}$  resulted from N–H stretching of amide A has clearly represented the protein of *Isochrysis galbana* biomass.

The presence of lipid constituent was confirmed by the appearance of two weak peaks at  $2856\text{ cm}^{-1}$  and  $2927\text{ cm}^{-1}$  related to both symmetric and asymmetric  $\text{CH}_2$  stretching of methylene (Jiang et al., 2012; Marshall et al., 2005; Mayers et al., 2013; Phukan et al., 2011; Sigee et al.,

2002), respectively. The weak peak at  $1786\text{ cm}^{-1}$  originated from C=O stretching of esters was also supposed to be lipid structure belonging.

These overall FTIR bands spectra have justified the existence of three primary constituents in the *Isochrysis galbana* biomass, i.e. carbohydrate, protein, and lipid. These constituents were the primary substances, which composed several microalgae cell structures. The algal cell wall was composed of a polymer of 1,4 linked  $\beta - D -$  glucose and an amorphous mucilaginous material constituted of polysaccharides, lipids, and proteins. The plasma membrane was composed of phospholipid bilayer and protein membrane. The chloroplast consisted of several membranes composed of lipid and protein, and it contains small spherical lipid droplets between the thylakoids. The mitochondrion's inner membrane contained of proteins greater than 70%. All cell structure constituents could be gradually cracked in the course of the thermal processes when the temperature extended to its decomposition temperature, and the fraction of material would be released to be volatile matter, while the residual constituent formed char as a solid material. Both volatile and char would be combusted during the combustion processes. The latest leftover material after the final combustion processes was ash as the last remaining material. This ash material was mainly arranged from the inorganic ingredient of biomass. The overall thermal decomposition phenomenon have been shown by progressive declining intensity in the spectra of biomass residue  $1000\text{ }^{\circ}\text{C}$ , mainly at around  $3590 - 1630\text{ cm}^{-1}$ .

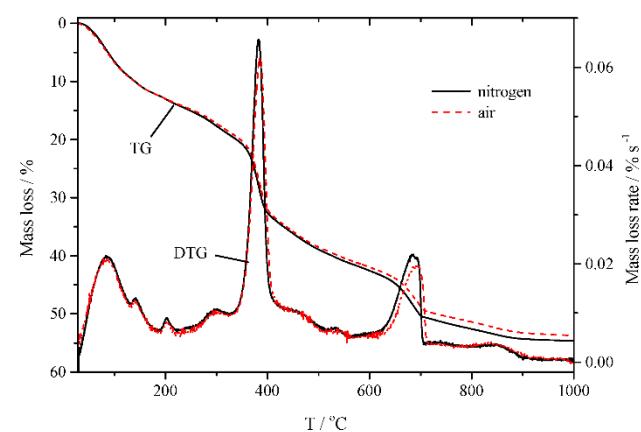


Fig. 4. TG-DTG profile of *Isochrysis galbana* under nitrogen and air atmosphere at  $10\text{ }^{\circ}\text{C min}^{-1}$ .

Table. 4. The physical properties of some biomass.

	Biomass	Proximate Analysis					Ref
		Moisture	VM	FC	Ash	HHV	
		(wt%)	(wt%)	(wt%)	(wt%)	(MJ kg <sup>-1</sup> )	
<b>Terrestrial plant</b>	<i>Bagasse</i>	-	70.90*	7.00*	22.10*	14.258	(1)
	<i>Sugar cane straw</i>	-	76.20*	14.60*	9.20*	17.19	(2)
	<i>Cotton residue</i>		72.80*	20.59*	6.61*	16.90	(2)
<b>Aquatic biomass</b>	<i>Chlorella sp. MP-1</i>	6.8	72.19	15.08	5.93	18.59	(3)
	<i>Chlorella vulgaris</i>	-	55.37*	34.35*	10.28*	21.88**	(4)
	<i>Chlorella vulgaris</i>	-	-	-	-	18	(5)
	<i>Chlorella minutissima</i>	-	-	-	-	21	(5)
	<i>Chlorella vulgaris</i>	9.1	37.3	5.0	48.6	-	(6)
<i>Enteromorpha clathrata</i>		13.30	41.82	7.79	37.09	7.89**	(7)
	<i>Sargassum natans</i>	10.46	48.85	11.60	29.09	8.68**	(7)
	<i>Nannochloropsis oculata</i>	3.99	64.76	7.76	23.50	-	(8)
<i>Isochrysis galbana</i>		-	67.45*	8.08*	24.47*	16.80	(8)
	<i>Isochrysis galbana</i>	12.98	40.10	7.47	39.45		This study
	<i>Isochrysis galbana</i>		46.08*	8.58*	45.33*	16.22	This study

\* indicated VM, FC and Ash in *dry basis* (db), other demonstrated on an *air-dried basis* (adb)

\*\* indicated lower heating value (LHV) in unit of MJ kg<sup>-1</sup>

(1) Parikh, Channiwala, & Ghosal (2005); (2) Yin (2011); (3) Phukan et al. (2011); (4) Chen, Ma, & Liu (2011); (5) Illman et al. (2000); (6) Agrawal & Chakraborty (2013); (7) Wang, Jiang, Han, & Liu (2009); (8) Sukarni et al. (2014)

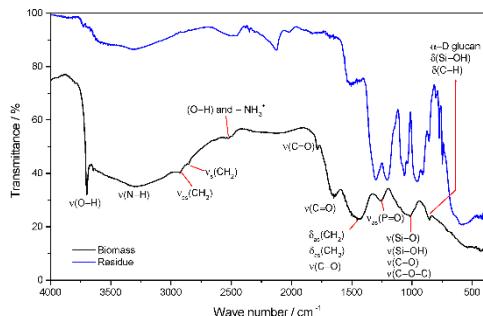


Fig. 5. The FTIR spectra of *Isochrysis galbana* biomass and its residue.

#### 4. CONCLUSIONS

The several significant characteristics of microalgae *Isochrysis galbana* for future solid biofuel have been investigated. In the growth kinetics point of view, with simple and inexpensive nutrient culturing, this alga has comparable growth rate to other algal species. The *Isochrysis galbana* physical properties indicate its visibility as compared to both aquatic biomass and the terrestrial plant. The elemental analysis of *Isochrysis galbana* points out that this biomass has a proportionality value against other algal biomass. The particular attention must be given to the high Mg content of this biomass since it could affect the residual behavior in the combustion system. The macromolecular contents of this alga has also indicated the presence of three primary constituents inside its biomass in terms of carbohydrate, protein, and lipid, which they thermally degraded and then converted into energy during the combustion process. The overall biomass characteristics as already mentioned above provides the essential information regarding the prospect of this alga as a sustainable solid biofuel feedstock and offers *Isochrysis galbana* as a possible candidature for future bioenergy source.

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#### REFERENCES

Agrawal, A., & Chakraborty, S. (2013). A kinetic study of pyrolysis and combustion of microalgae *Chlorella vulgaris* using thermo-gravimetric analysis. *Bioresource Technology*, 128, 72–80. <http://doi.org/10.1016/j.biortech.2012.10.043>

Ainane, T., Abourriche, A., & Kabbaj, M. (2015). Physico-chemical analysis by SEM-EDX and FTIR two brown algae *Cystoseira tamariscifolia* and *Bifurcaria bifurcata*. *BioTechnology An Indian Journal*, 11(5), 185–188.

Alkhamis, Y., & Qin, J. G. (2013). Cultivation of *Isochrysis galbana* in Phototrophic, Heterotrophic, and Mixotrophic Conditions. *BioMed Research International*, 2013.

Banyay, M., Sarkar, M., Gräslund, A. (2003). A library of IR bands of nucleic acids in solution, *Biophysical Chemistry*, 104 (2), 477-488.

Benning, L. G., Phoenix, V. R., Yee, N., & Konhauser, K. O. (2004). The dynamics of cyanobacterial silicification: An infrared micro-spectroscopic investigation. *Geochimica et Cosmochimica Acta*, 68(4), 743–757. [http://doi.org/10.1016/S0016-7037\(03\)00488-5](http://doi.org/10.1016/S0016-7037(03)00488-5)

Benning, L. G., Phoenix, V. R., Yee, N., & Tobin, M. J. (2004). Molecular characterization of cyanobacterial silicification using synchrotron infrared micro-spectroscopy. *Geochimica et Cosmochimica Acta*, 68(4), 729–741. [http://doi.org/10.1016/S0016-7037\(03\)00489-7](http://doi.org/10.1016/S0016-7037(03)00489-7)

Brennan, L., & Owende, P. (2010). Biofuels from microalgae—a review of technologies for production, processing, and extractions of biofuels and co-products. *Renewable and sustainable energy reviews*, 14(2), 557-577. <http://doi.org/10.1016/j.rser.2009.10.009>

Cantrell, K. B., Ducey, T., Ro, K. S., & Hunt, P. G. (2008). Livestock waste-to-bioenergy generation opportunities. *Bioresource Technology*, 99(17), 7941–7953. <http://doi.org/10.1016/j.biortech.2008.02.061>

Chen, C., Ma, X., & Liu, K. (2011). Thermogravimetric analysis of microalgae combustion under different oxygen supply concentrations. *Applied Energy*, 88(9), 3189–3196. <http://doi.org/10.1016/j.apenergy.2011.03.003>

Christenson, L., & Sims, R. (2011). Production and harvesting of microalgae for wastewater treatment, biofuels, and bioproducts. *Biotechnology Advances*, 29(6), 686–702. <http://doi.org/10.1016/j.biotechadv.2011.05.015>

Converti, A., Casazza, A. a., Ortiz, E. Y., Perego, P., & Del Borghi, M. (2009). Effect of temperature and nitrogen concentration on the growth and lipid content of *Nannochloropsis oculata* and *Chlorella vulgaris* for biodiesel production. *Chemical Engineering and Processing: Process Intensification*, 48(6), 1146–1151. <http://doi.org/10.1016/j.cep.2009.03.006>

Crowe, B., Attalah, S., Agrawal, S., Waller, P., Ryan, R., Van Wageningen, J., ... Huesemann, M. (2012). A comparison of *Nannochloropsis salina* growth performance in two outdoor pond designs: Conventional raceways versus the arid pond with superior temperature management. *International Journal of Chemical Engineering*, 1–9. <http://doi.org/10.1155/2012/920608>

Da Rosa, A. P. C., Carvalho, L. F., Goldbeck, L., & Costa, J. A. V. (2011). Carbon dioxide fixation by microalgae cultivated in open bioreactors. *Energy Conversion and Management*, 52(8–9), 3071–3073. <http://doi.org/10.1016/j.enconman.2011.01.008>

Corradi da Silva, M. d. L., Fukuda, E. K., Vasconcelos, A. F. D., Dekker, R. F. H., Matias, A. C., Monteiro, N. K., ... Carbonero, E. R. (2008). Structural characterization of the cell wall D-glucans isolated from the mycelium of *Botryosphaeria rhodina* MAMB-05. *Carbohydrate Research*, 343(4), 793–8. <http://doi.org/10.1016/j.carres.2007.12.021>

De Moraes, M. G., & Costa, J. A. V. (2007). Biofixation of carbon dioxide by *Spirulina* sp. and *Scenedesmus obliquus* cultivated in a three-stage serial tubular photobioreactor. *Journal of Biotechnology*, 129(3), 439–45. <http://doi.org/10.1016/j.jbiotec.2007.01.009>

Dean, A. P., Nicholson, J. M., & Sigee, D. C. (2008). Impact of phosphorus quota and growth phase on carbon allocation in *Chlamydomonas reinhardtii*: an FTIR microspectroscopy study. *European Journal of Phycology*, 43(4), 345–354. <http://doi.org/10.1080/09670260801979287>

Dean, A. P., Sigee, D. C., Estrada, B., & Pittman, J. K. (2010). Using FTIR spectroscopy for rapid determination of lipid accumulation in response to nitrogen limitation in freshwater microalgae. *Bioresource Technology*, 101(12), 4499–507. <http://doi.org/10.1016/j.biortech.2010.01.065>

Du, S., Yang, H., Qian, K., Wang, X., & Chen, H. (2014). Fusion and transformation properties of the inorganic components in biomass ash. *Fuel*, 117, 1281–1287. <http://doi.org/10.1016/j.fuel.2013.07.085>

Duygu, D. Y., Udoh, A. U., Ozer, T. B., Akbulut, A., Acikgoz, I., Yildiz, K., & Guler, D. (2012). Fourier transform infrared (FTIR) spectroscopy for identification of *Chlorella vulgaris* Beijerinck 1890 and *Scenedesmus obliquus* (Turpin) Kützing 1833. *African Journal of Biotechnology*, 11(16), 3817–3824.

El-Toni, A. M., Khan, A., Ibrahim, M. A., Labis, J. P., Badr, G., Al-Hoshan, M., ... Sato, T. (2012). Synthesis of double mesoporous core-shell silica spheres with tunable core porosity and their drug release and cancer cell apoptosis properties. *Journal of Colloid and Interface Science*, 378(1), 83–92. <http://doi.org/10.1016/j.jcis.2012.04.006>

Gao, Y., Yang, M., & Wang, C. (2013). Nutrient deprivation enhances lipid content in marine microalgae. *Bioresource Technology*, 147, 484–491. <http://doi.org/10.1016/j.biortech.2013.08.066>

García-González, M., Moreno, J., Manzano, J. C., Florencio, F. J., & Guerrero, M. G. (2005). Production of *Dunaliella salina* biomass rich in 9-cis-beta-carotene and lutein in a closed tubular photobioreactor. *Journal of Biotechnology*, 115(1), 81–90. <http://doi.org/10.1016/j.jbiotec.2004.07.010>

Goo, B. G., Baek, G., Choi, D. J., Park, Y. Il, Synytsya, A., Bleha, R., ... Park, J. K. (2013). Characterization of a renewable extracellular polysaccharide from defatted microalgae *Dunaliella tertiolecta*. *Bioresource Technology*, 129, 343–350. <http://doi.org/10.1016/j.biortech.2012.11.077>

Guo, H., Daroch, M., Liu, L., Qiu, G., Geng, S., & Wang, G. (2013). Biochemical features and bioethanol production of microalgae from coastal waters of Pearl River Delta. *Bioresource Technology*, 127, 422–428. <http://doi.org/10.1016/j.biortech.2012.10.006>

Huang, Q., Jin, Y., Zhang, L., Cheung, P. C. K., & Kennedy, J. F. (2007). Structure, molecular size and antitumor activities of polysaccharides from *Poria cocos* mycelia produced in fermenter. *Carbohydrate Polymers*, 70(3), 324–333. <http://doi.org/10.1016/j.carbpol.2007.04.015>

Illman, A., Scragg, A., & Shales, S. (2000). Increase in *Chlorella* strains calorific values when grown in low nitrogen medium. *Enzyme and Microbial Technology*, 27(8), 631–635. [http://doi.org/10.1016/S0141-0229\(00\)00266-0](http://doi.org/10.1016/S0141-0229(00)00266-0)

Jangir, D. K., Charak, S., Mehrotra, R., & Kundu, S. (2011). FTIR and circular dichroism spectroscopic study of interaction of 5-fluorouracil with DNA. *Journal of Photochemistry and Photobiology. B, Biology*, 105(2), 143–8. <http://doi.org/10.1016/j.jphotobiol.2011.08.003>

Jiang, Y., Yoshida, T., & Quigg, A. (2012). Photosynthetic performance, lipid production and biomass composition in response to nitrogen limitation in marine microalgae. *Plant Physiology and Biochemistry: PPB / Société Française de Physiologie Végétale*, 54, 70–77. <http://doi.org/10.1016/j.plaphy.2012.02.012>

Kamnev, A. A., Ristić, M., Antonyuk, L. P., Chernyshev, A. V., & Ignatov, V. V. (1997). Fourier transform infrared spectroscopic study of intact cells of the nitrogen-fixing bacterium *Azospirillum brasiliense*. *Journal of Molecular Structure*, 409, 201–205.

Kaplan, D., Cohen, Z., & Abeliovich, A. (1986). Optimal growth conditions for *Isochrysis galbana*. *Biomass*, 9(1), 37-48.

Lecellier, A., Mounier, J., Gaydou, V., Castrec, L., Barbier, G., Ablain, W., ... Sockalingum, G. D. (2014). Differentiation and identification of filamentous fungi by high-throughput FTIR spectroscopic analysis of mycelia. *International Journal of Food Microbiology*, 168-169, 32-41. <http://doi.org/10.1016/j.ijfoodmicro.2013.10.011>

Marshall, C., Javaux, E., Knoll, a., & Walter, M. (2005). Combined micro-Fourier transform infrared (FTIR) spectroscopy and micro-Raman spectroscopy of Proterozoic acritarchs: A new approach to Palaeobiology. *Precambrian Research*, 138(3-4), 208-224. <http://doi.org/10.1016/j.precamres.2005.05.006>

Mayers, J. J., Flynn, K. J., & Shields, R. J. (2013). Rapid determination of bulk microalgal biochemical composition by Fourier-Transform Infrared spectroscopy. *Bioresource Technology*, 148, 215-220. <http://doi.org/10.1016/j.biortech.2013.08.133>

Mecozzi, M., Pietroletti, M., & Di Mento, R. (2007). Application of FTIR spectroscopy in ecotoxicological studies supported by multivariate analysis and 2D correlation spectroscopy. *Vibrational Spectroscopy*, 44(2), 228-235. <http://doi.org/10.1016/j.vibspec.2006.11.006>

Meng, Y., Yao, C., Xue, S., & Yang, H. (2014). Application of Fourier transform infrared (FT-IR) spectroscopy in determination of microalgal compositions. *Bioresource Technology*, 151, 347-354. <http://doi.org/10.1016/j.biortech.2013.10.064>

Murdock, J. N., & Wetzel, D. L. (2009). FT-IR Microspectroscopy Enhances Biological and Ecological Analysis of Algae. *Applied Spectroscopy Reviews*, 44(4), 335-361. <http://doi.org/10.1080/05704920902907440>

Obernberger, I., Brunner, T., & Barnthaler, G. (2006). Chemical properties of solid biofuels—significance and impact. *Biomass and Bioenergy*, 30(11), 973-982. <http://doi.org/10.1016/j.biombioe.2006.06.011>

Parida, S. K., Dash, S., Patel, S., & Mishra, B. K. (2006). Adsorption of organic molecules on silica surface. *Advances in Colloid and Interface Science*, 121(1-3), 77-110. <http://doi.org/10.1016/j.cis.2006.05.028>

Parikh, J., Channiwala, S. a., & Ghosal, G. K. (2005). A correlation for calculating HHV from proximate analysis of solid fuels. *Elsevier Ltd.*, 84(5), 487-494. <http://doi.org/10.1016/j.fuel.2004.10.010>

Patil, P. D., Gude, V. G., Mannarswamy, A., Cooke, P., Munson-McGee, S., Nirmalakhandan, N., ... Deng, S. (2011). Optimization of microwave-assisted transesterification of dry algal biomass using response surface methodology. *Bioresource Technology*, 102(2), 1399-405. <http://doi.org/10.1016/j.biortech.2010.09.046>

Peng, Y., Zhang, L., Zeng, F., & Kennedy, J. F. (2005). Structure and antitumor activities of the water-soluble polysaccharides from *Ganoderma tsugae* mycelium. *Carbohydrate Polymers*, 59(3), 385-392. <http://doi.org/10.1016/j.carbpol.2004.10.009>

Peng, Y., Zhang, L., Zeng, F., & Xu, Y. (2003). Structure and antitumor activity of extracellular polysaccharides from mycelium. *Carbohydrate Polymers*, 54(3), 297-303. [http://doi.org/10.1016/S0144-8617\(03\)00190-5](http://doi.org/10.1016/S0144-8617(03)00190-5)

Phukan, M. M., Chutia, R. S., Konwar, B. K., & Kataki, R. (2011). Microalgae Chlorella as a potential bio-energy feedstock. *Applied Energy*, 88(10), 3307-3312. <http://doi.org/10.1016/j.apenergy.2010.11.026>

Ragusa, S., Cambria, M. T., Pierfederici, F., Scirè, A., Bertoli, E., Tanfani, F., & Cambria, A. (2002). Structure-activity relationship on fungal laccase from *Rigidoporus lignosus*: a Fourier-transform infrared spectroscopic study. *Biochimica et Biophysica Acta*, 1601(2), 155-162.

Sanchez, A., González, A., Maceiras, R., Cancela, Á., & Urrejola, S. (2011). Raceway pond design for microalgae culture for biodiesel. *Chemical Engineering Transactions*, 25, 845-850. <http://doi.org/10.3303/CET1125141>

Sánchez Mirón, A., Cerón García, M. C., Contreras Gómez, A., García Camacho, F., Molina Grima, E., & Chisti, Y. (2003). Shear stress tolerance and biochemical characterization of *Phaeodactylum tricornutum* in quasi steady-state continuous culture in outdoor photobioreactors. *Biochemical Engineering Journal*, 16(3), 287-297. [http://doi.org/10.1016/S1369-703X\(03\)00072-X](http://doi.org/10.1016/S1369-703X(03)00072-X)

Schwenk, D., Seppälä, J., Spilling, K., Virkki, A., Tamminen, T., Oksman-Caldentey, K. M., & Rischer, H. (2013). Lipid content in 19 brackish and marine microalgae: Influence of growth phase, salinity and temperature. *Aquatic Ecology*, 47(4), 415-424. <http://doi.org/10.1007/s10452-013-9454-z>

Selvakumar, P., & Umadevi, K. (2014). Mass Cultivation of Marine Micro alga *Nannochloropsis gaditana* KF410818 Isolated from Visakhapatnam offshore and Fatty Acid Profile Analysis for Biodiesel Production. *Journal of Algal Biomass Utilization*, 5(1), 28-37.

Sigee, D. C. D., Dean, A., Levado, E., & Tobin, M. J. (2002). Fourier-transform infrared spectroscopy of *Pediastrum duplex*: characterization of a micro-population isolated from a eutrophic lake. *European Journal of Phycology*, 37(1), 19-26. <http://doi.org/10.1017/S0967026201003444>

Silverstein, R. M., Webster, F. X., & Kiemle, D. (2005). *Spectrometric Identification of Organic Compounds* (7th Editio). John Wiley & Sons.

Stehfest, K., Toepel, J., & Wilhelm, C. (2005). The application of micro-FTIR spectroscopy to analyze nutrient stress-related changes in biomass composition of phytoplankton algae. *Plant Physiology and Biochemistry: PPB / Société Française de Physiologie Végétale*, 43(7), 717-726. <http://doi.org/10.1016/j.plaphy.2005.07.001>

Sukarni, Sudjito, Hamidi, N., Yanuhar, U., & Wardana, I. N. G. (2014). Potential and properties of marine microalgae *Nannochloropsis oculata* as biomass fuel feedstock. *International Journal of Energy and Environmental Engineering*, 5(4), 279–290. <http://doi.org/10.1007/s40095-014-0138-9>

Sukarni, Sudjito, Hamidi, N., Yanuhar, U., & Wardana, I. N. G. (2015). Thermogravimetric kinetic analysis of *Nannochloropsis oculata* combustion in air atmosphere. *Frontiers in Energy*, 9(2), 125–133. <http://doi.org/10.1007/s11708-015-0346-x>

Tan, S.-T., Balasubramanian, R. K., Das, P., Obbard, J. P., & Chew, W. (2013). Application of mid-infrared chemical imaging and multivariate chemometrics analyses to characterise a population of microalgae cells. *Bioresource Technology*, 134, 316–23. <http://doi.org/10.1016/j.biortech.2013.01.060>

Tianqi, W., Hanxiang, L. I., Manyi, W., & Tianwei, T. A. N. (2007). Integrative extraction of ergosterol, (1→ 3)- $\alpha$ -D-glucan and chitosan from *Penicillium chrysogenum* mycelia. *Chinese Journal of Chemical Engineering*, 15(5), 725–729.

Wang, B., Li, Y., Wu, N., & Lan, C. Q. (2008). CO<sub>2</sub> bio-mitigation using microalgae. *Applied Microbiology and Biotechnology*, 79(5), 707–18. <http://doi.org/10.1007/s00253-008-1518-y>

Wang, S., Jiang, X. M., Han, X. X., & Liu, J. G. (2009). Combustion Characteristics of Seaweed Biomass. 1. Combustion Characteristics of *Enteromorpha clathrata* and *Sargassum natans*. *Energy & Fuels*, 23(10), 5173–5178. <http://doi.org/10.1021/ef900414x>

Wysokowski, M., Behm, T., Born, R., Bazhenov, V. V., Meissner, H., Richter, G., ... Ehrlich, H. (2013). Preparation of chitin-silica composites by in vitro silicification of two-dimensional *Ianthella basta* demosponge chitinous scaffolds under modified Stöber conditions. *Materials Science & Engineering. C, Materials for Biological Applications*, 33(7), 3935–3941. <http://doi.org/10.1016/j.msec.2013.05.030>

Yin, C. (2011). Prediction of higher heating values of biomass from proximate and ultimate analyses. *Fuel*, 90(3), 1128–1132. <http://doi.org/10.1016/j.fuel.2010.11.031>

Yusuf, C. (2007). Biodiesel from microalgae. *Biotechnology Advances*, 25(3), 294–306. <http://doi.org/10.1016/j.biotechadv.2007.02.001>