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# Spatial and temporal organization of aquatic insect assemblages in two subtropical river drainages

Organización espacial y temporal de ensamblajes de insectos acuáticos en dos cuencas subtropicales

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# ABSTRACT

Background. The spatial and temporal changes of assemblages of aquatic insect can be used to detect the anthropic impacts that influence the biological communities. Goals. We compared the assemblages of aquatic insect in 1997 and 2014 in two subtropical river drainages, the association with water characteristics, and we discuss their implications for ecosystems conservation. Methods. True diversity of the aquatic insect fauna at family level and their community structure for 27 study sites in 1997 and 2014 were assessed. Multivariate analyzes were used to compare aquatic insect assemblages and the abundance of functional feeding groups. Results. There were significant differences in the dissolved oxygen (DO) of the water between 1997 and 2014, decreasing its values. Other variables correlated to DO were also modified, with a decrease in pH and an increase in temperature. We found a correlation between reduction of DO and water pH with a decline in the overall abundance of aquatic insects; also, with shifts in the community structure, from the decrease of groups such as some Ephemeroptera and scrapers, to the increase in opportunistic families such as Chironomidae, Culicidae, and other predator families such as Coenagrionidae, Corixidae and Veliidae, and more abundance of collectors. Families such as Heptageniidae and Caenidae decreased in abundance, as well as other benthic groups. Conclusions. The assemblages of aquatic insect are useful to indicate a generalized degradation of environmental conditions across localities and time in two subtropical river drainages, related to water quality degradation symptoms such as reduction of pH levels and dissolved oxygen, usually associated with anthropogenic stressors.

**Keywords:** environmental degradation, functional feeding groups, macroinvertebrates, true diversity, water quality.

# RESUMEN

Antecedentes. Los cambios espaciales y temporales de los ensamblajes de insectos acuáticos pueden ser utilizados para detectar los impactos antrópicos que influyen en las comunidades biológicas. Objetivos. Comparamos los ensamblajes de insectos acuáticos en 1997 y 2014 en dos cuencas subtropicales, su asociación con las características del agua y discutimos sus implicaciones para la conservación de los ecosistemas. Métodos. Se evaluó la diversidad verdadera a nivel de familia, de la fauna de insectos acuáticos en 27 sitios de estudio en 1997 y 2014. Se utilizaron análisis multivariados para comparar los ensamblajes de insectos acuáticos y la abundancia de los grupos funcionales de alimentación. Resultados. Se obtuvieron diferencias significativas en el oxígeno disuelto (OD) del agua entre 1997 y 2014, disminuyendo sus valores. También observó una disminución de pH y una tendencia a un incremento de la temperatura. Se identificó una relación entre la disminución de oxígeno y valores menores de pH con una reducción general en la abundancia de insectos acuáticos; asimismo, se observa una relación con cambios en los ensamblajes como lo son una disminución en la representación de grupos como Ephemeroptera y raspadores, el incremento de familias como Chironomidae, Culicidae, Coenagrionidae y Veliidae, y una mayor abundancia de colectores. Familias como Heptageniidae y Caenidae disminuyeron en abundancia, así como otros grupos bentónicos. Conclusiones. Los ensamblajes de insectos acuáticos son útiles para indicar una degradación generalizada de las condiciones a través de las localidades y el tiempo en las dos cuencas subtropicales de estudio, con



síntomas de degradación de la calidad del agua como la disminución de los niveles de pH y oxígeno disuelto, generalmente asociados con factores de estrés antropogénicos.

**Palabras clave**: calidad de agua, degradación ambiental, diversidad verdadera, grupos funcionales de alimentación, macroinvertebrados.

#### INTRODUCTION

Among the aquatic ecosystems, rivers benefit human communities by providing a supply of water, nutrient retention, removal of toxins, microclimate stability, opportunities for tourism, and are valued by local cultures (Brismar, 2002; Dudgeon, 2019). The main cause of loss of the ecological integrity and deterioration of these ecosystems are human activities (Carpenter *et al.*, 2011; Dudgeon, 2019), while the major threats for freshwater biodiversity are overexploitation, water pollution and flow modification, the invasion of exotic species, land use change, and climate change (Dudgeon, 2019). At the basin scale, land use changes influences in the stream conditions modifying water characteristics, sediment supply and deposition, affecting bank stability, and consequently the aquatic biota (Strayer *et al.*, 2003; Townsend *et al.*, 2003; Allan, 2004).

In order to design adequate proposals for new management and conservation of fluvial ecosystems, it has been proposed to study selected indicator groups and how the ecological elements and processes in these catchments have changed in the long term to reveal their current ecological condition and future threats (Ramírez & Gutiérrez-Fonseca, 2014a). Historical analyzes of changes in aquatic communities offers information about the current conservation status of aquatic ecosystems, in order to infer factors that have impacted these systems and obtain insight into the changing conditions of the surrounding watershed (Karr, 1981; Fausch *et al.*, 1990). Aquatic macroinvertebrates have a range of preferences for environmental conditions, so shift in the assemblages may reflect changes in the aquatic ecosystem and human impacts over time (Li *et al.*, 2012). However, the long-term perspectives and historical comparisons in aquatic macroinvertebrate communities remains often short (Jackson & Füreder, 2006).

Aquatic macroinvertebrates are especially useful to evidence changes in river ecosystems due to anthropic impacts (Barbour et al., 1999; Bonada et al., 2006; Ligeiro et al., 2013). Among them, aquatic insects are generally the most abundant and diverse, as they are one of the most ecologically important groups (Macadam & Stockan, 2015) especially in tropical and subtropical zones (Dudgeon, 2008). They are the main primary consumers and are responsible for transferring the energy of primary productivity to other trophic levels of food chains, and there are elements within this group that are important predators (Hanson et al., 2010; Macadam & Stockan, 2015). Aquatic insects can have highly specific functions in the ecosystem, such as filterers, gatherers, shredders, predators, piercers and scrapers (Merritt et al., 2008; Hanson et al., 2010; Ramírez & Gutierrez-Fonseca, 2014b). Importantly, because aquatic insects deploy a wide array of generalist and specialist feeding strategies, occupy several microhabitats, and have different responses and sensitivities to habitat degradation, they are considered highly useful biological indicators of stream ecological condition (Karr & Chu, 1999).

On the other hand, we must also bear in mind that macroinvertebrate assemblages can not only be affected by pollution or degradation. It has been seen that these assemblages vary due to the flow regime and sediment deposition (Díaz-Rojas *et al.*, 2020). So that high areas of a basin may have greater diversity because the variations of flows given by the slopes allows greater heterogeneity in the landscape than in the low sections where the slope decreases as well as flow (Mesa, 2010). Likewise, when the flow increases in the rainy season, the communities are modified (Quesada-Alvarado *et al.*, 2020). Therefore, it is important to consider the seasonality of the samples, being also throughout the year that differences have been seen in the macroinvertebrate assemblages as the flows and chemical composition of the rivers is modified, for example, by the leachate of the soil in rainy season (Leal-Bastidas *et al.*, 2021).

In the tropics, the influence of hydrological, physical and chemical alterations upon macroinvertebrate communities remains poorly understood (Md Rawi et al., 2014; Ramírez et al., 2015). Several studies on the ecology of aquatic insects in Latin America have been reported, with emphasis on the relationship with abiotic factors (Ramírez & Gutierrez-Fonseca, 2014a). For example, recently, Kohlmann et al. (2021) include in their study a analyzes of the relationships between functional feeding groups of aquatic macroinvertebrates with physicochemical such as NO<sup>3-</sup> K<sup>+</sup>, biochemical oxigen demand (BDO), oxygen saturation, and pH; Díaz-Rojas et al. (2020) relates depth, flow velocity, channel width and roughness of the substrate with macroinvertebrate assemblages composition and functional traits; Quesada-Alvarado et al. (2020) describe the relationship between the aquatic macroinvertebrate assemblages with physicochemical and habitat variables, such as NO<sup>3-</sup>, substrate and flow; and, Mosquera-Restrepo & Peña-Salamanca (2019) explain the relationships between aquatic macroinvertebrates assemblages with dissolved oxygen, BOD, total dissolved solids, and turbidity.

In Mexico, this approach has been used to assess the biotic integrity of rivers in the Río Chiquito basin in the State of Michoacán (Piñón-Flores *et al.*, 2014), variation of macro-invertebrates in the Laguna de Tecocomulco in the State of Hidalgo for one year period (Rico-Sánchez *et al.*, 2014), and impacts of mining activities in three rivers of the Sierra Gorda Biosphere Reserve (Rico-Sánchez *et al.*, 2022). However, these studies involve brief spatial and temporal scale. For these reasons, the present study has the main goal of compare aquatic insect assemblages data of 1997 with data of 2014. Community structure, diversity, functional feeding groups, and the associations with water characteristics were described in two major subtropical river drainages in east-central Mexico, to interpret the ecological impairment indicated by the patterns founded.

#### MATERIAL AND METHODS

**Study area:** The study area includes the Pánuco and Lerma-Chapala river drainages, located in east-central Mexico (Fig. 1). It has a subtropical area in the northeast, located in the Eastern Sierra Madre and the Neovolcanic Belt. Central Mexico has the most degraded river drainages in the country (Mercado-Silva *et al.*, 2006). The Lerma-Chapala and Pánuco river drainages are two of the most important basins of this region, and have been highly impacted by loss of vegetation cover (>30%), expansion of cultivated pastures for livestock, increased agricultural activities, combined with expanded industrialization and urbanization

(Cuevas et al., 2010). The Lerma-Chapala river drainage shows an evident problem of physical and chemical anthropogenic transformation, and is considered as the most degraded in Mexico (Cotler-Avalos *et al.*, 2004). At the present, the headwaters of both drainages are being considered for special protection status as water reserves in Mexico by the National Commission of Water (Comisión Nacional del Agua, 2011).

A total of 27 sampling sites were selected in permanent rivers and were sampled in 1997 and 2014. We chose some of the main waterways in the following five states (Fig. 1) which included: 1) Aguascalientes: San Pedro River and Calvillo River; 2) Jalisco: Grande River; 3) Guanajuato: Laja River and Apaseo River; 4) Querétaro: Extóraz River, Huimilpan River, Querétaro River, San Juan River, Jalpan River and Santa María River; and, 5) San Luis Potosí: Verde River. The field work was conducted in the dry season (from February to May), when the conditions of habitat and biological community of rivers are more stable (Pérez-Munguía *et al.*, 2007; Lyons *et al.*, 1995) and the effect of the human activities are more evident (Moncayo-Estrada *et al.*, 2015).

Data collection: Water physical and chemical parameters were measured with a multimeter probe (Hach Hydromet Quanta, Loveland,

Colorado, USA), and we included pH, dissolved oxygen (mg/L), and temperature (°C). Aquatic insects were sampled using a D-net (300 mm of diameter and 300 µm of mesh size) in all different types of reachable habitat, with a sample effort of 60 minutes per study site. During 1997, aquatic insects were preserved in alcohol in 125 ml jars and brought back to the laboratory and separated from detritus. During 2014, insects were separated in situ and were deposited into a plastic vial and preserved in 80% alcohol solution for further transport to the laboratory (Biotic Integrity Lab at Universidad Autónoma de Querétaro). The aquatic insects were identified to the taxonomic level of family using specialized keys (e.g., Arce-Pérez & Roughley, 1999; Merritt et al., 2008; Bueno-Soria, 2010; Springer et al., 2010). We used the family taxonomic level because it has proven to be a good indicator of the level of ecological disturbance in fluvial ecosystems (cf. Marshall et al., 2006; Serrano-Balderas et al., 2016; Wright & Ryan, 2016), allows for the categorization of functional traits for the different families in most cases, and highest taxonomic level still providing sufficient resolution regarding biological traits of the organisms, saving time and effort to reach lower taxonomic categories. The functional feeding groups (FFG) were obtained from Ramírez & Gutiérrez-Fonseca (2014b).



Figure 1. Geographic location of study sites. 1 = Fracción Sánchez, 2 = Planta-La Hacienda, 3 = Puente la Plazuela, 4 = Pinihuan, 5 = Canoas, 6 = Quinta Matilde, 7 = El Realito, 8 = Quiotillos, 9 = El Salto, 10 = Presa del Carmen, 11 = Presa de Rayas, 12 = Comonfort, 13 = La Quemada, 14 = Los Galvanes, 15 = El Xote, 16 = El Qasis, 17 = Chuveje, 18 = Carpintero, 19 = Rascón, 20 = Tamasopo, 21 = Jalpan, 22 = Ayutla, 23 = Santa María (before of Adjuntas), 24 = El Carrizal (Santa Maria after of Adjuntas), 25 = Río Grande, 26 = Calvillo, 27 = Sabinolandia (El Salto de los Salados).

#### Statistical analysis

**Water physicochemistry**: Paired T-test were made to assess the difference of the values of each water parameter between two years (1997 and 2014). To analyze and elucidate patterns of all physical and chemical parameters in both years, a principal component analysis (PCA) was conducted, and we normalize all variables using division by their standard deviations because the variables are measured in different units.

Aquatic insect assemblages: We calculated the true diversity as proposed by Jost (2006, 2007), through assessing of effective numbers of elements at family level, that refers to the numbers of taxa equally probable and necessary to obtain a diversity value (Jost, 2007). This approach is considered logical and works intuitively, unlike other indices such as the Shannon entropy, which measures the uncertainty degree of a species (Jost, 2006). True alpha and gamma diversity of first order was obtained to sensitize the index to the abundant species, because in aquatic insect communities are common to find this pattern of very abundant and rare taxa, which means an inequitable distribution of abundances among all taxa. Jackknife estimator was used because it is appropriate for this group of organisms (Basualdo, 2011; Martínez-Sanz et al., 2010). In addition, the true beta diversity was obtained as the effective number of elements in the data set (true gamma diversity) divided by the average number of effective elements of the samples (true alpha diversity); where, one is the minimum number which we can obtained, indicating that all communities are exactly the same, and maximum value are equal to the total of communities (N) (Jost, 2007). We applied paired T test to assess the difference of diversity values between 1997 and 2014, and a similarity percentage analysis (SIMPER) using the Bray-Curtis similarity measure (multiplied with 100), based in abundance per family and FFG was used to identify which taxon discriminates among periods (Clarke, 1993).

**Responses of aquatic insect assemblages to water physicochemistry**: To assess the effect of water physicochemistry on aquatic insect assemblages we made a non-metric multidimensional scaling (NMDS) based on the Bray-Curtis similarity index, which can be used with cero values in data sets (Bray & Curtis, 1957). NMDS was applied using the abundance per family and per FFG. In the NMDS the environmental variables were associated to the axis and represented with vectors in the plot (Hammer *et al.*, 2001). We correlated by the Spearman method the PCA values and NMDS scores to understand the relationships between abiotic variables and aquatic insect assemblages overall, considering the intrinsic relationships, as was used by Escalera-Vázquez & Zambrano (2010) in a study of the effect of variation in abiotic factors on fish assemblages.

The paired T-test and Spearman rank order correlation analysis (Zar, 2014) were made using the statistical software SPSS version 20 (IBM corp., 2011). The true diversity values (Jost, 2006, 2007) were estimated with SPADE software (Chao & Shen, 2010). The multivariate analysis PCA, NMDS and SIMPER (Quinn & Keough, 2002) were obtained using PAST version 3.07 (Hammer *et al.*, 2001).

#### RESULTS

Water physicochemistry: The paired T-test shows a significantly decrease (p<0.001) of dissolved oxygen between 1997 and 2014 considering all study sites (i.e., both basins), from 8.2 ± 3.3 mg/L to 3.6 ± 2.2 mg/L. There are no significant differences between temperature (20.3 ±

4.31 to 20.5  $\pm$  4.8 °C) and pH (8.01  $\pm$  0.43 to 7.8  $\pm$  0.35) of both years (p>0.05). Nevertheless, the PCA analysis showed a subtle tendency gradient of segregation of data between 1997 and 2014. Three of the main components (PC) had moderately related variables (0.75-0.50). Of these PC1 (eigenvalue=1.76) explained 58.73% of the variance, PC2 (eigenvalue=0.75) explained 25% and PC3 (eigenvalue=0.48) 16.26%. The first component (PC1) showed moderate positively association with the three variables with correlation coefficients of 0.63(pH), 0.58 for dissolved oxygen (DO) and 0.51 for temperature. While the second was strongly positively associated (>0.75) with temperature (0.81) and moderate negatively associated with DO (-0.55) and not so with the pH (-0.15). Whereas the PC3 showed a negative association with pH (-0.76) and positive with DO (0.59) and temperature (0.25). The study sites ordination resulted located diagonally from upper left corner to the lower right corner, following a decrease of dissolved oxygen and pH values, and from the lower left corner to the upper right corner following an increase in temperature. The sites located in the upper left corner zone, comprises mainly the sites sampled in 2014 (Fig. 2).

**Aquatic insect assemblages:** A total of 71 aquatic insect families were obtained including both drainages. We collected 47 families during 1997 and 61 families during 2014 (Table 1). We found more representativeness of taxa during 2014 and more gamma diversity for both river drainages (Lerma-Chapala,  ${}^{1}D_{\alpha} = 8.2$  and Pánuco  ${}^{1}D_{\alpha} = 7.82$  during 1997; and Lerma-Chapala,  ${}^{1}D_{\alpha} = 9.17$  and Pánuco  ${}^{1}D_{\alpha} = 12.13$  during 2014). The beta diversity was higher in 2014 ( ${}^{1}D_{\beta} = 2.8$ ) than in 1997 ( ${}^{1}D_{\beta} = 1.4$ ) on Lerma-Chapala River drainage, and lower in 2014 ( ${}^{1}D_{\beta} = 1.4$ ) than 1997 ( ${}^{1}D_{\beta} = 1.61$ ) on Pánuco River drainage. Global gamma and beta diversity was also higher in 2014 ( ${}^{1}D_{\gamma} = 13.34$ ;  ${}^{1}D_{\beta} = 2.08$ ) than 1997 ( ${}^{1}D_{\gamma} = 9.30$ ;  ${}^{1}D_{\beta} = 1.81$ ). However, the Lerma-Chapala river drainage showed higher alpha and beta diversity of families in the 70% of study sites during 1997 (Table 2). These results are consistent with the results of paired T-test that showed not significantly difference of alpha diversity of all sites between years (p=0.181).

The SIMPER analysis showed that the main families with contribution for abundance dissimilarity between 1997 and 2014 were Chironomidae (24.8%), Baetidae (16.5%), Coenagrionidae (8.5%), Veliidae (4.9%), Corixidae (4.2%), Culicidae (3.9%), Caenidae (3.7%) and Heptageniidae (1.3%). The abundance of Chironomidae was  $61 \pm 185$  in 1997 and 116 ± 257.7 in 2014; of Baetidae was  $60.7 \pm 104.9$  in 1997 and 38.9 ± 57.2 in 2014; Coenagrionidae showed 2.44 ± 6 in 1997 and 46 ± 129.3 in 2014; Veliidae 2.26 ± 5.1 in 1997 and 22.2 ± 55 in 2014; Corixidae showed 6.48 ± 30.7 of mean abundance in 1997 and 20 ± 51 during 2014; Culicidae 0.5 ± 1.5 in 1997 and 40.7 ± 195.2 in 2014; Caenidae 25.3 ± 99.3 in 1997 and 2.29 ± 13.5 during 2014; and Heptageniidae 3.22 ± 10.15 in 1997 and 0.07 ± 0.38 in 2014.

We found the six FFG: gatherers, filterers, predators, shredders, piercers, and scrapers. The most abundant FFG in both years was the gatherers, follow by predators, and piercers were the rarest (Table 3). SIMPER analysis based on abundance per FFG showed that gatherers contributed with 55.6% to the dissimilitude, with change from 79% of gatherers in 1997 to 44% in 2014; predators contributed with 32% and the quantity of individuals changed from  $23 \pm 23$  in 1997 to  $120 \pm 188$  in 2014; filterers contributed with 8.9% and changed from  $8 \pm 33$  to  $48 \pm 194$  individuals between 1997 and 2014; scrapers contributed with 2% and changed from  $3.5 \pm 10.1$  in 1997 to  $0.8 \pm 2.4$  in 2014; shredders contributed with 0.7% and piercers with 0.5%.



Figure 2. Principal Components Analysis based on physicochemical parameters in rivers of two subtropical river drainages in east-central Mexico (Lerma-Chapala and Pánuco). Data from 1997 (triangles), data from 2014 (circles). Ayutla = Ayu; Calvillo = Cal; Canoas = Can; Carpintero = Car; Chuveje = Chu; Comonfort = Com; El Carrizal (Santa María after of Adjuntas) = SMD; El Oasis = EO; El Realito = ER; El Salto = ES; El Xote = EX; Fracción Sánchez = FS; Jalpan = Jal; La Hacienda = LH; La Quemada = LQ; Los Galvanes = LG; Pinihuan = Pin; Presa de Rayas = PR; Presa del Carmen = PC; Puente la Plazuela = PP; Quinta Matilde = QM; Quiotillos = Qui; Rascón = Ras; Río Grande = RG; Sabinolandia (El Salto de los Salados) = Sab; Santa María (before of Adjuntas) SM; Tamasopo = Tam.

Relationships between aquatic insect assemblages and water characteristics: The NMDS based on number of individuals per site (Fig. 3), showed a pattern where taxa were ordinated in a gradient of dissolved oxygen and pH decrease from the upper left corner to the lower right corner. Additionally, the ordination analysis showed a gradient of decrease in abundance per site in the same direction (from the upper left to the lower right corner). We found some sites with contrasting differences between years, where Comonfort (Com) had 1008 individuals in 1997 and 21 individuals in 2014; Sabinolandia (Sab) changed from 298 individuals in 1997 to 36 in 2014; Fracción Sanchez (FS) increased the number of individuals from 85 to 2491 in 1997 and 2014, respectively; however, ~40% (1017 individuals) belong to Culicidae family and the insect diversity decrease in time (Table 2). The NMDS analysis shows a relationship between low concentrations of dissolved oxygen and lower pH values with fewer number of individuals per site; however, it shows no tendency of grouping by sampling years.

The NMDS based on relative abundance of insects per FFG (Fig. 4) showed that study sites were ordinated on axis one (left to right) in a gradient from low to high number of predators. The second axis (bottom to up) show a gradient of a greater number of filterers, a smaller number of scrapers and gatherers, and lower values of pH and dissolved oxygen. We found important changes in functional feeding groups at some study sites such as Comonfort (Com) where the abundance of gatherers decreased from 82.9% in 1997 to 9.5% in 2014 and the abundance of filterers (from 0.1% to 52.4%) and predators (from 0.2% to 38.1%) increased drastically between 1997 and 2014. At the Ayutla

(Ayu) location, the abundance of gatherer decreased from 84.9% to 62.7%, the abundance of filterers and predator increased from 1.9% to 11.6% and from 3.8% to 22.8%, respectively. Fracción Sánchez (FS) showed considerable increase of filterers from 10.6% to 40.8% from 1997 to 2014. This analysis shows a slightly relationship between lower levels of dissolved oxygen and pH with high abundance of filterers and lower abundance of scrapers.

Most of the correlations between the values of PCA with the values from Axis of the NMDS were not significant. The only significant correlation was based on abundance per FFG, using the axis 2 ( $r_{xy} = -0.28$ , p = 0.04). The correlation shows that the increase in water temperature and the decrease in dissolved oxygen is related with more abundance of filterers and less abundance of scrapers; however, in this case the correlation coefficient is very low showing that this pattern is not consistent.

#### DISCUSSION

The rivers in the Lerma-Chapala and Pánuco river drainages showed symptoms of biological and environmental degradation based on differences in aquatic insect diversity and taxa abundance, FFG and water quality parameters such as dissolved oxygen and pH. The aquatic insect structure and the relationship with water physiochemical variables through the space and time were difficult to interpret at the basin scale. However, our analyzes provided general patterns such as the condition of the aquatic insect fauna and water characteristics in two major subtropical river drainages in east-central Mexico.

Table 1. Number of individuals and functional feeding groups per site in rivers in two sub-tropical river drainages in east-central Mexico (Lerma-Chapala River and Pánuco River). Values show the number of individuals. Sampling sites are in parentheses. FFG = Functional feeding group, 1 = El Salto, 2 = Presa del Carmen, 3 = Presa de Rayas, 4 = Comonfort, 5 = La Quemada, 6 = Los Galvanes, 7 = El Xote, 8 = Río Grande, 9 = Calvillo, 10 = Sabinolandia (El Salto de los Salados), 11 = Fracción Sánchez, 12 = La Planta-La Hacienda, 13 = Puente la Plazuela, 14 = Pinihuan, 15 = Canoas, 16 = Quinta Matilde, 17 = El Realito. 18 = Quiotillos, 19 = El Oasis, 20 = Chuvejé, 21 = Carpintero, 22 = Rascón, 23 = Tamasopo, 24 = Jalpan, 25 = Ayutla, 26 = Santa María (above of Adjuntas), 27 = El Carrizal (Sta. Ma. below of Adjuntas).

Family	FFG	1997	2014			
Ephemeroptera						
Baetidae	Gatherer	25(1), 26(2), 1(3), 294(4), 135(5), 93(6), 35(9), 35(10), 44(11), 4(12), 5(13), 45(14), 14(17), 182(18), 466(19), 21(20), 19(22), 13(23), 100(24), 18(25), 11(26), 52(27)	7(2), 52(3), 185(5), 29(6), 21(8), 185(9), 1(11), 3(12), 109(13), 127(15), 12(16), 3(17), 99(18), 5(19), 47(20), 6(22), 13(24), 112(25), 12(26), 21(27)			
Ephemerellidae	Gatherer	10(11), 66(18)	0			
Polymitarcyidae	Gatherer	1(2), 1(3), 1(13), 12(18)	0			
Caenidae	Gatherer	519(4), 2(7), 24(10), 52(18), 16(19), 29(20), 6(21), 6(22), 8(23), 13(24), 6(26), 1(27)	70(5), 10(18)			
Leptophlebiidae	Gatherer	9(5), 1(6), 16(10), 5(18), 12(19), 63(20), 5(21), 15(26)	9(14), 1(15), 4(16), 15(22), 1(23), 9(24), 5(25), 6(26)			
Leptohyphidae	Gatherer	5(26)	1(2), 4(5), 134(13), 6(15), 7(16), 9(18), 10(19), 3(20), 7(22), 1(23), 5(24), 2(27)			
Heptageniidae	Scraper	9(4), 1(9), 2(10), 1(19), 1(20), 4(21), 1(22), 2(23), 1(24), 1(25), 12(26), 52(27)	2(20)			
Ephemeridae	Gatherer	22(24)	0			
Odonata						
Gomphidae	Predator	7(7), 1(10), 1(17), 3(20)	2(6), 16(13), 10(14), 1(15), 1(16), 9(19), 3(21), 6(22), 12(24), 46(25), 5(26), 15(27)			
Coenagrionidae	Predator	2(1), 23(2), 1(3), 14(5), 1(7), 18(9), 2(12), 2(14), 1(21), 1(24), 1(26)	20(2), 178(3), 1(4), 88(5), 67(6), 1(7), 1(8), 5(9), 70(12), 20(13), 4(14), 10(15), 11(16), 6(17), 663(18), 21(19), 9(20), 3(22), 2(23), 43(24), 6(25), 5(26), 8(27)			
Lestidae	Predator	0	3(3), 3(14), 50(15), 107(16), 20(20)			
Platystictidae	Predator	0	2(26)			
Macromiidae	Predator	0	1(27)			
Libellulidae	Predator	1(10), 4(17), 1(18), 1(19), 4(20), 40(24)	5(2), 2(3), 7(4), 8(7), 3(9), 54(13), 1(14), 6(15), 6(16), 3(18), 3(19), 1(20), 8(24), 9(25), 2(27)			
Aeshnidae	Predator	2(1), 5(2), 8(5), 2(6), 14(10), 1(11), 3(12), 17(20), 2(26)	1(1), 4(2), 13(3), 1(16), 14(18), 1(22)			
Calopterygidae	Predator	2(1), 1(6), 1(10), 5(18), 3(24)	4(12), 3(14), 1(15), 3(19), 2(20), 3(25), 4(26), 1(27)			
Protoneuridae	Predator	3(5), 1(14)	1(9), 3(21)			
Plecoptera						
Perlidae	Predator	1(13), 1(14), 2(18)	5(14)			
Hemiptera						
Corixidae	Predator	160(4), 1(8), 10(18), 4(25)	1(1), 143(3), 167(5), 71(6), 159(9)			
Hebridae	Predator	0	1(5), 1(13), 1(26), 4(27)			
Veliidae	Predator	2(1), 1(4), 1(5), 13(10), 5(11), 2(12), 1(13), 5(14), 2(15), 2(16), 24(18), 1(20), 1(21), 1(22)	$\begin{array}{l} 6(2),\ 10(5),\ 7(8),\ 2(11),\ 2(12),\ 212(13),\ 3(14),\ 202(15),\\ 3(16),\ 4(17),\ 47(18),\ 42(20),\ 1(21),\ 1(23),\ 43(25),\ 15(26) \end{array}$			

Family	FFG	1997	2014				
Mesoveliidae	Predator	0	2(12)				
Gerridae	Predator	3(7), 32(10), 10(12), 6(13), 17(15), 7(16), 23(17), 1(18), 10(20), 7(23)	1(3), 4(5), 9(13), 2(15), 7(16), 8(18), 2(21), 5(27)				
Belostomatidae	Predator	2(1), 6(3), 1(4), 2(7), 1(16), 2(18), 2(19)	5(2), 5(5), 4(7), 1(8), 17(12), 6(13), 1(15), 3(16), 3(17), 1(20), 9(21)				
Naucoridae	Predator	1(6), 4(10), 4(10), 5(11), 17(18), 3(19), 1(22), 5(24)	40(13), 2(21), 3(24), 2(25)				
Notonectidae	Predator	4(16), 5(18)	3(5), 1(6), 1(8), 23(9), 2(11), 1(12), 3(15), 21(16), 43(18), 14(20), 21(27)				
Saldidae	Predator	1(17)	0				
Pleidae	Predator	10(5), 72(6), 3(19)	96(18)				
Macroveliidae	Predator	0	47(18)				
Nepidae	Predator	0	1(2), 1(5), 1(6), 1(11), 4(12), 11(13), 2(16), 2(18)				
Megaloptera							
Corydalidae	Predator	1(10), 1(18), 5(21)	3(14), 3(19), 2(20), 1(23), 1(25), 5(26), 1(27)				
Trichoptera							
Hydroptilidae	Piercer	1(7), 1(9), 2(24)	9(5), 2(15), 1(20), 1(24), 3(25), 12(26)				
Polycentropodi- dae	Filterer	0	12(15), 1(16), 2(18), 8(20), 1(21)				
Philopotamidae	Filterer	12(10), 1(21)	1(16), 3(24), 25(26), 4(27)				
Odontoceridae	Shredder	0	1(20)				
Hydrobiosidae	Predator	0	1(14), 2(25), 1(27)				
Limnephilidae	Shredder	2(5), 1(10)	0				
Calamocerati- dae	Shredder	0	3(20), 2(22)				
Lepidostomati- dae	Shredder	0	3(20)				
Leptoceridae	Gatherer	0	1(15), 1(20), 1(24)				
Hydropsychidae	Filterer	25(10), 11(24)	1(12), 1(14), 1(19), 2(24), 11(26), 3(27)				
Coleoptera							
Gyrinidae	Predator	1(12), 18(14), 7(16), 4(18)	1(5), 1(6), 2(8), 29(16), 1(18), 1(20), 7(24), 3(25)				
Dytiscidae	Predator	1(1), 2(2), 1(5), 1(18), 3(20)	4(1), 12(3), 5(5), 13(9), 94(11), 12(13), 31(16), 4(18), 2(19), 5(20), 1(22), 4(25)				
Hydrophilidae	Predator	2(6), 1(7), 1(12), 1(17), 3(18)	2(2), 6(3), 16(5), 3(7), 2(8), 82(9), 1(10), 26(11), 1(12), 11(13), 1(15), 6(16), 3(17), 4(18), 2(19), 1(20)				
Helophoridae	Gatherer	0	1(11)				
Staphylinidae	Gatherer	0	1(5), 1(9), 1(15)				
Psephenidae	Scraper	1(18), 5(19), 1(26)	0				
Scirtidae	Scraper	1(9)	3(5), 11(12), 2(25)				
Dryopidae	Shredder	1(5), 1(22)	1(26)				
Elmidae	Gatherer	1(1), 2(4), 2(5), 1(6), 1(9), 19(10), 1(13), 4(18), 9(20), 16(22), 3(23), 154(24), 16(26)	4(13), 3(15), 5(19), 11(20), 1(22), 3(23), 19(24), 9(25), 7(26), 5(27)				
Limnichidae	Gatherer	1(20), 3(24)	0				

Family	FFG	1997	2014
Lutrochidae	Shredder	0	9(25), 5(26)
Ptiliidae	Scraper	0	1(8), 4(20)
Haliplidae	Shredder	1(5), 6(9)	1(7),1(13), 1(15), 1(20)
Diptera			
Tipulidae	Shredder	0	1(5), 1(27)
Ceratopogoni- dae	Predator	1(1), 1(2), 11(5), 1(8), 2(13), 3(17), 5(24), 2(25)	1(3), 5(5), 1(6), 1(19), 8(25), 3(26), 3(27)
Chironomidae	Gatherer	21(1), 63(2), 5(3), 19(4), 72(5), 7(6), 4(7), 2(8), 16(9), 92(10), 11(11), 25(12), 33(13), 1(14), 5(15), 5(16), 4(17), 39(18), 40(19), 67(20), 8(21), 11(22), 11(23), 980(24), 27(25), 80(26)	$\begin{array}{llllllllllllllllllllllllllllllllllll$
Simuliidae	Filterer	1(2), 1(4), 3(5), 1(10), 1(11), 2(20), 160(24), 1(25)	11(2), 7(5), 7(8), 1(14), 1(16), 1(19), 2(22), 9(24), 65(25), 15(26), 8(27)
Syrphidae	Gatherer	0	1(1), 6(10), 1(11)
Dixidae	Gatherer	0	4(5), 8(18), 3(24)
Culicidae	Filterer	2(2), 8(11), 1(12), 1(15), 2(17), 1(18), 1(20)	12(1), 11(4), 5(5), 29(10), 1017(11), 10(13), 15(18), 1(22)
Thaumaleidae	Scraper	1(18)	0
Tabanidae	Predator	1(1), 1(2), 1(10), 1(13), 1(19), 1(24)	2(14), 2(17)
Stratiomyidae	Gatherer	0	1(11), 1(13), 3(25), 1(27)
Muscidae	Predator	0	1(22)
Ephydridae	Gatherer	6(9)	4(9), 12(11)
Psychodidae	Gatherer	2(4), 6(24)	0
Chaoboridae	Predator	2(5), 1(13), 1(20), 5(24)	0
Athericidae	Predator	0	2(20)
Empididae	Predator	1(25)	0
Lepidoptera			
Crambidae	Shredder	3(10), 1(13), 1(15), 1 (18),	1(18), 2(25), 1(26)

It has been shown that the loss of vegetation cover in watershed, mainly riparian vegetation, is a contributing factor in the increase of temperature in freshwater bodies (Allan, 2004; Quinn *et al.*, 1997). The processes of urbanization are related with degradation symptoms such as the reduction of dissolved oxygen in the water (De Jesús-Crespo & Ramírez, 2011; Ometo *et al.*, 2000). These patterns are consistent with the decrease of both dissolved oxygen concentrations and pH values, and an increase in water temperature finding in our study and whit the fact that Cuevas *et al.* (2010) estimates that the Lerma-Chapala and Pánuco River drainages have lost about 30% and 50% of the vegetation cover respectively, due to the expansion of cultivated pastures, increased agriculture, and urbanization.

Biodiversity measurement has been considered has good indicator of ecosystem stability (Maclaurin & Sterelny, 2008). However, some authors argue that alpha diversity often do not present systematic patterns among habitats, which does not always make them as good indicators of the severity of human impacts (cf. Magurran, 2016; Pandolfi & Lovelock, 2014). Another constraint is the fact that all diversity metrics are limited by the ability of researchers to measure them in field, i.e., the community is rarely perfectly measured varying across taxonomic groups, environments, and traits (Jarzyna & Jetz, 2016). The absence of significant differences in the diversity of aquatic insects and the inconsistent patterns in this biological measure in our study, comparing our data from 1997 with data of 2014, are similar to other research, where different gradients of urbanization or river ecosystems degradation were analyzed at the basin scale with no clear responses and patterns in the richness and evenness of aquatic invertebrates (Bonada *et al.*, 2006; Quinn *et al.*, 1997).

The increase in abundance in Chironomidae, Coenagrionidae, Veliidae, Corixidae and Culicidae families, can be related with the environmental degradation. A positive relationship has been reported between the increase of Chironomidae density with land use changes, such as induced grassland and urban sprawl (Jones & Clark, 1987; Quinn et al., 1997). These land use changes are generally associated with an increase in water temperature and sedimentation, and low dissolved oxygen concentrations (Miserendino et al., 2011; Walsh et al., 2005). Chironomids are found in a range of conditions more extensive than any other aquatic insect family; it can exploit an almost complete range of gradient in temperature, pH and oxygen (Ferrington et al., 2008). For this reason, is not surprising that this diverse and opportunistic family showed greater relative abundance in the 2014 when compared to 1997, which was correlated with a decrease of dissolved oxygen and lower values of pH. The larvae of some Odonata are also tolerant and often survive relatively low values of dissolved oxygen and subsist better than many other invertebrates in acidic waters (Suhling et al., 2015), which could explain the increment in individuals of the Coenagrionidae family. Some aquatic Heteroptera, especially Gerromorpha, are good indicators of human disturbance having a high tolerance to eutrophication and acidic waters. Corixidae present a great variation among nutrient and pH tolerance (Lytle, 2015). Accordingly, the increment in these two families of hemipterans, especially Corixidae could be a response of lower pH values. The Culicidae increase in 2014 also can be related with anthropogenic stressors. In this sense, Ribeiro et al. (2012) reported that environmental change, such as the increase in agricultural areas, irrigation ponds, and the reduction in vegetation cover, tends to increase the abundance of opportunistic species of Culicidae, mainly those species that are considered vectors of human diseases (Juliano & Lounibos, 2005).

Although Baetidae is an Ephemeroptera family very common and dominant in tropical and subtropical rivers, in this study showed an abundance decrease (together with Heptageniidae and Caenidae families) from the sampling performed in 1997 compared with the year 2014. Baetidae and Heptageniidae families are reported to be sensitive to land use changes, such as urban and cropland increase (Jones & Clark, 1987; Li *et al.*, 2012; Quinn *et al.*, 1997) because many live attached to boulders and feed on the periphyton (Flowers & De la Rosa, 2010). In general, land use changes can result in an increase in fine sediment deposition, reducing available habitat for benthic organisms (Wood & Armitage, 1997) and resulting in a decrease in periphyton (Yamada & Nakamura, 2002) affecting the establishment and development of families such as Heptageniidae.

In terms of the functional feeding groups (FFG), the increase in gatherers, filterers and predators, and the decrease of scrapers are similar to other studies where a reduction in the of river ecosystem integrity is related with agriculture activities and urbanization processes. In this way, Quinn *et al.* (1997) and Friberg *et al.* (2009) registered an increment of filterers densities and Md Rawi *et al.* (2014) document an increase of predators, filterers, and gatherers in association with environmental degradation. This pattern of increase in collectors (filterers and gatherers) can be an indicator of environmental degradation, because filterers have more availability of suspended particles, and gatherers too with the increase in sediment deposition, which implies, in many cases, more fine particulate organic matter as available feeding resources for these groups. The land use change at basin scale, reduction of riparian vegetation cover, wastewater and pollutants discharges, Table 2. True diversity (number of effective elements) of aquatic insects, alpha (Jack1) beta and gamma (Jack1) of rivers in two sub-tropical river drainages in east-central Mexico (Lerma-Chapala River and Pánuco River) including two years (1997 and 2014). Ayutla = Ayu; Calvillo = Cal; Canoas = Can; Carpintero = Car; Chuveje = Chu; Comonfort = Com; El Carrizal (Sta. Ma. after of adjuntas) = SMD; El Oasis = EO; El Realito = ER; El Salto = ES; El Xote = EX; Fracción Sánchez = FS; Jalpan = Jal; La Hacienda = LH; La Quemada = LQ; Los Galvanes = LG; Pinihuan = Pin; Presa de Rayas = PR; Presa del Carmen = PC; Puente la Plazuela = PP; Quinta Matilde = QM; Quiotillos = Qui; Rascón = Ras; Río Grande = RG; Sabinolandia (El Salto de los Salados) = Sab; Santa María (before of adjuntas) = SM; Tamasopo = Tam. No significant difference of alpha diversity between years were obtained (p = 0.133).

Lerma-Chapala			Pánuco River			
Study site	1997	2014	Study site	1997	2014	
Cal	5.27	1.64	Ayu	3.4	7.4	
Com	3.19	3.24	Can	3.2	5.06	
ES	5.07	1.63	Car	7.53	6.45	
EX	9.24	2.2	Chu	7.57	12.48	
LG	2.95	4.44	EO	1.98	9.5	
LQ	5.15	6.56	ER	5.6	3.21	
PC	4.27	3.59	FS	4.81	2.52	
PR	5.8	4.88	Jal	3.66	10.12	
RG	4.74	2.34	LH	5.27	3.38	
Sab	10.2	1.9	Pin	3.2	11.83	
Gamma	8.2	9.17	PP	4.83	7.93	
Beta	1.49	2.8	QM	5.8	9.13	
			Qui	7.79	4.97	
			Ras	5.31	10.41	
			SM	4.94	14.2	
			SMD	2.12	12	
			Tam	5.44	9.54	
			Gamma	7.82	12.13	
			Beta	1.61	1.4	
		1997	2014			
Global <i>(Both river</i>	Gamma	9.3	13.34			
drainages)	Beta	1.81	2.08			

can cause cumulative and additive effects, which impinges on the river community, changing the habitat, water quality and nutrient amount (Allan, 2004). These disturbances provide favorable conditions to some opportunistic groups such as filterers and gatherers. Sedimentation, for example, restricts the suitability for periphyton and biofilm production (Wood & Armitage, 1997; Yamada & Nakamura, 2002) limiting the success of scrapers that feed on it.

The pattern of increase in temperature shown in the PCA (Fig. 2), were not accurately related with the aquatic insect assemblages. It coincides with Friberg *et al.* (2009) and Buendia *et al.* (2014) who found no correlation or strong effect among water temperature and aquatic macroinvertebrate diversity. On the other hand, Jacobsen *et al.* (1997), report a positive relationship between water temperature increase and aquatic invertebrate richness; however, their study focuses on Ecuado-

Table 3. Relative abundance (%) and mean of number of individuals ( $\mu$ ) per functional feeding groups of aquatic insects during 1997 and 2014 in rivers of two Sub-tropical river drainages of east-central Mexico. BRD = both river drainages; SE = standard deviation G = gatherer; Ft = filterers; Pr = predator; Sh = shredders; Pc = piercers; Sc = scrapers. Superscripts *a*, *b* refers to among-year differences (CG, *p* = 0.012; Ft, *p* = 0.005; Sc, *p* = 0.026).

		Year									
		1997					2014				
	Lerma-Chapala			Pánuco	BRD %	Lerma-Chapala		Pánuco		BRD %	
	%	Μ	%	Μ		%	М	%	μ SD		
CG	74.8	155.2± 250.4	82.2	$170.9 \pm 318.5$	79.4ª	56.9	169.5± 180.2	36.2	93.3± 98.9	44.6 <sup>b</sup>	
Ft	2.2	4.5± 11.8	5.4	11.2 ± 41.2	4.2ª	2.8	8.2± 9.1	4.8	12.3± 18.2	4.0 <sup>b</sup>	
Pr	13.8	28.7± 27.6	9.6	19.9 ± 19.7	11.1	21.6	64.2± 73.5Ω	57	147± 228.6	42.7	
Sh	1.1	2.2± 4.3	0.1	$0.1 \pm 0.5$	0.4	0.2	0.5±1.1	1.3	$3.3\pm5.8$	0.8	
Pc	0.1	$0.2 \pm 0.4$	0.1	$0.1 \pm 0.5$	0.1	0.3	0.9± 2.8	0.5	1.2± 2.9	0.4	
Sc	0.6	1.3± 2.8	2.3	4.9± 12.6	1.7ª	0.1	0.4± 1.0	0.6	$1.5\pm3.2$	0.4 <sup>b</sup>	



Figure 3. Non metric multidimensional scaling (NMDS) based on the number of individual of aquatic insects per family, in rivers in the Lerma-Chapala and Pánuco River drainages, with data of 1997 (triangles) and 2014 (circles). Stress: 0.19. Temp = temperature, D0 = dissolved oxygen. Ayutla = Ayu; Calvillo = Cal; Canoas = Can; Carpintero = Car; Chuveje = Chu; Comonfort = Com; El Carrizal (Santa María after of Adjuntas) = SMD; El Oasis = EO; El Realito = ER; El Salto = ES; El Xote = EX; Fracción Sánchez = FS; Jalpan = Jal; La Hacienda = LH; La Quemada = LQ; Los Galvanes = LG; Pinihuan = Pin; Presa de Rayas = PR; Presa del Carmen = PC; Puente la Plazuela = PP; Quinta Matilde = QM; Quiotillos = Qui; Rascón = Ras; Río Grande = RG; Sabinolandia (El Salto de los Salados) = Sab; Santa María (before of Adjuntas) SM; Tamasopo = Tam.

rian mountain streams and temperate lowland streams, and refers to a lower range of temperatures, contrasting with those of this study. Li *et al.* (2012) report that an increase in water temperature was correlated with the scarcity of Ephemeroptera, Plecoptera and Trichoptera. We also found a correlation among lower pH levels and lower dissolved oxygen concentrations with decrease in aquatic insect abundance, more pre-

sence of filterers and a decrement on scrapers. These results appear to differ with Townsend *et al.* (1983) who found more abundance of filter feeders in sites with higher levels of pH in temperate streams that are commonly acid, and they attribute this pattern to the greater range of resources in fewer acid streams.



Figure 4. Non metric multidimensional scaling (NMDS) based on relative abundance of aquatic insects per functional feeding group in rivers of two Sub-tropical river drainages including data from 1997 (triangles) and 2014 (circles) Stress: 0.09. Temp = temperature, D0 = dissolved oxygen. Ayutla = Ayu; Calvillo = Cal; Canoas = Can; Carpintero = Car; Chuveje = Chu; Comonfort = Com; El Carrizal (Santa María after of Adjuntas) = SMD; El Oasis = EO; El Realito = ER; El Salto = ES; El Xote = EX; Fracción Sánchez = FS; Jalpan = Jal; La Hacienda = LH; La Quemada = LQ; Los Galvanes = LG; Pinihuan = Pin; Presa de Rayas = PR; Presa del Carmen = PC; Puente la Plazuela = PP; Quinta Matilde = QM; Quiotillos = Qui; Rascón = Ras; Río Grande = RG; Sabinolandia (El Salto de los Salados) = Sab; Santa María (before of Adjuntas) SM; Tamasopo = Tam.

The negative relationship among dissolved oxygen and aquatic insect abundance found in this study, coincides with other studies where it is suggested that the availability of dissolved oxygen restricts macroinvertebrate diversity (Jacobsen et al., 1997; García-Alzate et al., 2010; Md Rawi et al., 2014). Moreover, it has reported that with a decrease in dissolved oxygen there is an increase in insect predators (Md Rawi et al., 2014), which is in accordance with our data increment of the mean abundance of Coenagrionidae, Veliidae and Corixidae, a predator's groups. Only one location (Fracción Sánchez) showed a substantial increase in the number of individuals, explained by the addition of 1017 individuals (>40% of abundance) of the family Culicidae, but a decrease in insect diversity from 1997 to 2014. The positive relationship between reduced water dissolved oxygen and an increase in the filterers, is mainly explained by the great abundance of individuals of Culicidae (categorized as filterers) in 2014, whose members are independent of water dissolved oxygen as they can obtain this resource directly from the atmosphere (Clements, 1992; Wallace & Walker, 2008).

Across the ecosystems of the world, freshwaters are the most endangered (Nel *et al.*, 2009), and subtropical streams and rivers are especially threatened because are greatly diverse ecosystems usually more than temperate waters (Dudgeon, 2008). Additionally, some pressures are increasing in developing countries (Strayer & Dudgeon, 2010), most located within tropical and subtropical zones (cf. Sachs, 2001). The rivers of the Lerma-Chapala and Pánuco River drainages have been affected by the combined and cumulative negative effects of human activities (Cuevas *et al.*, 2010).

The results of this study show symptoms of both chemical and biological degradation of these subtropical rivers. The evidence is supported by an increase in water temperature, and a decrease in dissolved oxygen concentrations and lower pH water levels along the space and time. For the aquatic insects, there was an increase in opportunistic and tolerant taxa with a corresponding decrease in sensitive groups. Also, the patterns in the FFG included an increase in the collectors (filterers and gatherers) and a decrease in scrapers. These symptoms reflected loss of river functional processes including energy transformation, nutrient turnover, storage and processing of organic matter, retention and cycling of nutrients, and transportation and deposition of sediments in both river drainages, with the most severe changes occurring in the Lerma-Chapala. In this basin the degradation condition was evident, because the anthropic impacts, including loss of habitat, contamination of waters, increase in sediment deposition and loss of riparian vegetation cover caused by human population growth and agricultural and livestock activities has been noticeably greater than in the Pánuco River basin (Cotler-Avalos et al., 2004; Cuevas et al., 2010).

Our data reflected the generalized degradation of rivers of two subtropical river drainages in east-central Mexico, which continues unabated and it is evidence of a deficiency in ecosystems conservation strategies in the country. This degradation it's a risk for the support ecological systems and the public health, because generates the conditions for the proliferation of mosquitoes, capable of transmitting viral infectious diseases such as Dengue (Secretaría de Salud, 2001; Instituto de Diagnóstico y Referencia Epidemiológicos, 2016), Chikungunya (Staples & Fischer, 2014) and Zika (Secretaría de Salud, 2016), which have occurred in Mexico and other tropical and subtropical zones.

This is the first analysis in Mexico that explores the relationship between aquatic insect assemblages and water quality variables, using this information to indicate degradation levels in two major river drainages through a long-term time scale comparison. This research contributes to the understanding of the trends in the responses of the aquatic biota related to water parameters, providing a framework for the application of historical comparison studies for evaluating the ecological conditions in rivers and to interpret the surrounding landscape impairment in other similar subtropical zones.

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### REFERENCES

- ALLAN, J. D. 2004. Landscapes and riverscapes: The influence of land use on stream ecosystems. *Annual Review of Ecology, Evolution,* and Systematics 35(1): 257-284.
- ARCE-PÉREZ, R. & R. E. ROUGHLEY. 1999. Lista anotada y claves para los Hydradephaga (Coleoptera: Adephaga: Dytiscidae, Noteridae, Haliplidae, Gyrinidae) de México. *Dugesiana* 6(2): 69-104.
- BARBOUR, M. T., J. GERRITSEN, B. D. SNYDER & J. B. STRIBLING. 1999. Rapid bioassessment protocols for use in streams and wadeable rivers: Periphyton, benthic macroinvertebrates and fish. 2nd ed. U.S. Environmental Protection Agency; Office of Water. Washington, D.C. EPA 841-B-99-002.
- BASUALDO, C. V. 2011. Choosing the best non-parametric richness estimator for benthic macroinvertebrates databases. *Revista de La Sociedad Entomológica Argentina* 70(1-2): 27-38.
- BONADA, N., N. PRAT, V. H. RESH & B. STATZNER. 2006. Developments in aquatic insect biomonitoring: A comparative analysis of recent approaches. *Annual Review of Entomology* 51(1): 495-523.
- BRAY, J. R. & J. T. CURTIS. 1957. An ordination of the upland forest communities of Southern Wisconsin. *Ecological Monographs* 27(4): 325-349.
- BRISMAR, A. 2002. River Systems as Providers of Goods and Services: A Basis for Comparing Desired and Undesired Effects of Large Dam Projects. *Environmental Management* 29(5): 598-609.
- BUENDIA, C., C. N. GIBBINS, D. VERICAT & R. J. BATALLA. 2014. Effects of flow and fine sediment dynamics on the turnover of stream invertebrate assemblages. *Ecohydrology* 7(4): 1105-1123.
- BUENO-SORIA, J. 2010. Guía ilustrada para la identificación de géneros de larvas de insectos del Orden Trichoptera de México. Universidad Nacional Autónoma de México. México D.F. 228 p.
- CARPENTER, S. R., E. H. STANLEY & M. J. VANDER-ZANDEN 2011. State of the world's freshwater ecosystems: physical, chemical, and biological changes. *Annual review of Environment and Resources* 36: 75-99.

- CHAO, A. & T. J. SHEN. 2010. Program SPADE (Species Prediction And Diversity Estimation). Available online at: http://chao.stat.nthu.edu.tw (downloaded May 25, 2018).
- CLARKE, K. R. 1993. Non-parametric multivariate analyses of changes in community structure. *Australian Journal of Ecology* 18(1): 117-143.
- CLEMENTS, A. N. 1992. *Biology of Mosquitoes: Development, Nutrition and Reproduction.* Springer, Netherland. 540 p.
- Comisión Nacional del Agua. 2011. Identificación de reservas potenciales de agua para el medio ambiente en México. Secretaria de Medio Ambiente y Recursos Naturales. México D.F. 85 p.
- COTLER-AVALOS, H., A. PRIEGO-SANTANDER, C. RODRÍGUEZ, C. ENRÍQUEZ-GUADARRAMA & J. C. FERNÁNDEZ. 2004. Determinación de zonas prioritarias para la eco-rehabilitación de la cuenca Lerma-Chapala. *Gaceta Ecológica* 71: 79-92.
- CUEVAS, M. L., A. GARRIDO, J. L. PÉREZ DAMIÁN & D. LURA-GONZÁLEZ. 2010. Procesos de cambio de uso de suelo y degradación de la vegetación natural. *In*: Cotler-Ávalos, H. (ed.). *Las cuencas hidrográficas de México: Diagnóstico y priorización*. Instituto Nacional de Ecología/ Fundación Gonzalo Río Arronte I.A.P. Mexico, pp. 96-103.
- DE JESÚS-CRESPO, R. & A. RAMÍREZ. 2011. Effects of urbanization on stream physicochemistry and macroinvertebrate assemblages in a tropical urban watershed in Puerto Rico. *Journal of the North American Benthological Society* 30: 739-750.
- DIAZ-ROJAS, C. A., Á. J. MOTTA-DIAZ & N. ARANGUREN-RIAÑO. 2020. Estudio de la diversidad taxonómica y funcional de los macroinvertebrados en un río de montaña Andino. *Revista de Biología Tropical* 68: 132-149.
- DUDGEON, D. 2008. Tropical stream ecology. Elsevier UK. 316 p.
- DUDGEON, D. 2019. Multiple threats imperil freshwater biodiversity in the Anthropocene. *Current Biology 29*(19): 960-967.
- ESCALERA-VÁZQUEZ, L. H. & L. ZAMBRANO. 2010. The effect of seasonal variation in abiotic factors on fish community structure in temporary and permanent pools in a tropical wetland. *Freshwater Biology* 55(12): 2557-2569.
- FAUSCH, K. D., J. LYONS, J. R. KARR & P. L. ANGERMEIER. 1990. Fish Communities as Indicators of Environmental Degradation. *American Fisheries Society Symposium* 8(1): 123-144.
- FERRINGTON, L. C., M. B. BERG & W. P. COFFMAN. 2008. Chironomidae. In: Merritt, R. W., K. W. Cummins & M. B. Berg (eds.). An introduction to the aquatic insects of North America. Kendall/Hunt Publishing Company, Dubque, Iowa, pp. 847-989.
- FLOWERS, R. W. & C. DE LA ROSA. 2010. Capítulo 4: Ephemeroptera. *Revista de Biología Tropical* 58(4): 63-93.
- FRIBERG, N., J. B. DYBKJÆR, J. S. OLAFSSON, G. M. GISLASON, S. E. LARSEN & T. L. LAURIDSEN. 2009. Relationships between structure and function in streams contrasting in temperature. *Freshwater Biology* 54(10): 2051-2068.
- GARCÍA-ALZATE, C. A., C. ROMÁN-VALENCIA, M. I. GONZALES & A. M. BARRERO. 2010. Composition and temporal variation of aquatic insect community (Insecta) in Sardineros Creek, Verde River drainage, upper Cauca, Colombia. *Revista de Investigaciones de la Universidad del Quindío* 21: 21- 28.

- tistics Software Package for Education and Data Analysis. *Paleontologia Electronica* 4(1): 9.
- HANSON, P., M. SPRINGER & A. RAMIREZ. 2010. Capítulo 1: Introducción a los grupos de macroinvertebrados acuáticos. *Revista de Biología Tropical* 58: 3-37.
- IBM CORP. 2011. IBM SPSS Statistics for Windows, Version 20.0. Armonk, N.Y.
- INSTITUTO DE DIAGNÓSTICO Y REFERENCIA EPIDEMIOLÓGICOS. 2016. Laboratorio de Arbovirus y Virus Hemorrágicos. Disponible en línea en: http:// www.indre.salud.gob.mx/interior/lab\_arbovirus\_1.html (consultado el 25 Mayo 2018).
- JACKSON, J. K. & L. FÜREDER. 2006. Long-term studies of freshwater macroinvertebrates: A review of the frequency, duration and ecological significance. *Freshwater Biology* 51(3): 591-603.
- JACOBSEN, D., R. SCHULTZ & A. ENCALADA. 1997. Structure and diversity of stream invertebrate assemblages: The influence of temperature with altitude and latitude. *Freshwater Biology* 38(2): 247-261.
- JARZYNA, M. A. & W. JETZ. 2016. Detecting the Multiple Facets of Biodiversity. Trends in Ecology & Evolution 31(7): 527-538.
- JONES, R. C. & C. C. CLARK. 1987. Impact of Watershed Urbanization on Stream Insect Communities1. *Journal of the American Water Resources Association* 23(6): 1047-1055.
- Jost, L. 2006. Entropy and diversity. Oikos 113(2): 363-375.
- JOST, L. 2007. Partitioning diversity into independent alpha and beta components. *Ecology* 88(10): 2427-2439.
- JULIANO, S. A. & L. P. LOUNIBOS. 2005. Ecology of invasive mosquitoes: Effects on resident species and on human health. *Ecology Letters* 8(5): 558-574.
- KARR, J. R. 1981. Assessment of Biotic Integrity Using Fish Communities. Fisheries 6(6): 21-27.
- KARR, J. R. & E. W. CHU. 1999. Restoring Life in Running Waters: Better Biological Monitoring. Island Press, Washington D.C. 220 p.
- KOHLMANN, B., D. VÁSQUEZ, A. ARROYO, & M. SPRINGER. 2021. Taxonomic and Functional Diversity of Aquatic Macroinvertebrate Assemblages and Water Quality in Rivers of the Dry Tropics of Costa Rica. *Frontiers in Environmental Science* 9:660260.
- LEAL-BASTIDAS, C., L. VARGAS-CHACOFF, N. SANDOVAL & P. FIERRO. 2021. Variabilidad temporal y espacial de los macroinvertebrados acuáticos y la calidad del agua en el río Palena, Patagonia Chilena. *Gayana* 85(2): 132-145.
- LI, F., N. CHUNG, M. J. BAE, Y. S. KWON & Y. S. PARK. 2012. Relationships between stream macroinvertebrates and environmental variables at multiple spatial scales. *Freshwater Biology* 57(10): 2107-2124.
- LIGEIRO, R., R. M. HUGHES, P. R. KAUFMANN, D. R. MACEDO, K. R. FIRMIANO, W. R. FERREIRA, D. OLIVEIRA, A. S. MELO & M. CALLISTO. 2013. Defining quantitative stream disturbance gradients and the additive role of habitat variation to explain macroinvertebrate taxa richness. *Ecological Indicators* 25: 45-57.
- LYONS, J., S. NAVARRO-PÉREZ, P. A. COCHRAN, E. C. SANTANA & M. GUZMÁN-ARROYO. 1995. Index of Biotic Integrity Based on Fish Assemblages for the Conservation of Streams and Rivers in West-Central Mexico. *Conservation Biology* 9(3): 569-584.

- LYTLE, D. A. 2015. Order Hemiptera. In: Thorp, J.H. & D.C. Rogers (eds.). Thorp and Covich's Freshwater Invertebrates. 4th ed. Elsevier, London, pp. 951-963.
- MACADAM, C. R. & J. A. STOCKAN. 2015. More than just fish food: Ecosystem services provided by freshwater insects. *Ecological Entomology* 40: 113-123.
- MACLAURIN, J. & K. STERELNY. 2008. What Is Biodiversity? 1st ed. University of Chicago Press, USA. 207 p.
- MAGURRAN, A. E. 2016. How ecosystems change. *Science* 351(6272): 448-449.
- MARSHALL, J. C., A. L. STEWARD & B. D. HARCH. 2006. Taxonomic Resolution and Quantification of Freshwater Macroinvertebrate Samples from an Australian Dryland River: The Benefits and Costs of Using Species Abundance Data. *Hydrobiologia* 572(1): 171-194.
- MARTÍNEZ-SANZ, C., F. GARCÍA-CRIADO, C. F. ALÁEZ & M. F. ALÁEZ. 2010. Assessment of richness estimation methods on macroinvertebrate communities of mountain ponds in Castilla y León (Spain). Annales de Limnologie International Journal of Limnology 46(02): 101-110.
- MD RAWI, C. S., S. A. AL-SHAMI, M. R. MADRUS & A. H. AHMAD. 2014. Biological and ecological diversity of aquatic macroinvertebrates in response to hydrological and physicochemical parameters in tropical forest streams of Gunung Tebu, Malaysia: Implications for ecohydrological assessment. *Ecohydrology* 7(2): 496-507.
- MERCADO-SILVA, N., J. LYONS, E. DÍAZ-PARDO, A. GUTIÉRREZ-HERNÁNDEZ, C. P. ORNELAS-GARCÍA, C. PEDRAZA-LARA & M. J. V. ZANDEN. 2006. Long-term changes in the fish assemblage of the Laja River, Guanajuato, central Mexico. *Aquatic Conservation: Marine and Freshwater Ecosystems* 16(5): 533-546.
- MERRITT, R. W., K. W. CUMMINS & M. B. BERG. 2008. An introduction to the aquatic insects of North America. 4th ed. Dubque, Iowa: Kendall/ Hunt Publishing Company. 1158 p.
- MESA, L. M. 2010. Hydraulic parameters and longitudinal distribution of macroinvertebrates in a subtropical andean basin. *Interciencia* 35(10):759-764
- MISERENDINO, M. L., R. CASAUX, M. ARCHANGELSKY, C. Y. DI PRINZIO, C. BRAND & A. M. KUTSCHKER. 2011. Assessing land-use effects on water quality, in-stream habitat, riparian ecosystems and biodiversity in Patagonian northwest streams. *Science of The Total Environment* 409(3): 612-624.
- MONCAYO-ESTRADA, R., J. LYONS, J. P. RAMIREZ-HERREJON, C. ESCALERA-GALLAR-DO & O. CAMPOS-CAMPOS. 2015. Status and Trends in Biotic Integrity in a Sub-Tropical River Drainage: Analysis of the Fish Assemblage Over a Three Decade Period. *River Research and Applications* 31(7): 808-824
- MOSQUERA-RESTREPO, D. & E. J. PEÑA-SALAMANCA. 2019. "Ensamblaje" de macroinvertebrados acuáticos y su relación con variables fisicoquímicas en un río de montaña en Colombia. *Revista de Biología Tropical* 67(6): 1235-1246.
- NEL, J. L., D. J. ROUX, R. ABELL, P. J. ASHTON, R.M. COWLING, J. V. HIGGINS, M. THIEME, J. H. VIERS. 2009. Progress and challenges in freshwater conservation planning. *Aquatic Conservation: Marine and Freshwater Ecosystems* 19(4): 474-485.

- OMETO, J. P. H. B., L. A. MARTINELLI, M. V. BALLESTER, A. GESSNER, A. V. KRUSCHE, R. L. VICTORIA & M. WILLIAMS. 2000. Effects of land use on water chemistry and macroinvertebrates in two streams of the Piracicaba river basin, south-east Brazil. *Freshwater Biology* 44(2): 327-337.
- PANDOLFI, J. M. & C. E. LOVELOCK. 2014. Novelty Trumps Loss in Global Biodiversity. *Science* 344(6181): 266-267.
- PÉREZ-MUNGUÍA, R. M., R. F. PINEDA-LÓPEZ & M. MEDINA-NAVA. 2007. Integridad biótica de ambientes acuáticos. *In*: Herzig, M., E. P. Recagno, O. Sánchez, L. Zambrano & R. M. Huitzil (eds.). *Perspectivas de la conservación de ecosistemas acuáticos en México*. Instituto Nacional de Ecología, México, pp. 71-111.
- PIÑÓN-FLORES, M. A. P., R. M. PÉREZ-MUNGUÍA, U. TORRES-GARCÍA & M. MEDI-NA-NAVA. 2014. Integridad biótica de la microcuenca del Río Chiquito, Morelia, Michoacán, México, basada en la comunidad de macroinvertebrados acuáticos. *Revista De Biología Tropical* 62(2): 221-231.
- QUESADA-ALVARADO, F., G. UMAÑA VILLALOBOS, M. SPRINGER, & J. PICADO BARBOZA. 2020. Variación estacional y características fisicoquímicas e hidrológicas que influyen en los macroinvertebrados acuáticos, en un río tropical. *Revista de Biología Tropical* 68: 54-67.
- QUINN, G. P. & M. J. KEOUGH. 2002. Experimental Design and Data Analysis for Biologists. Cambridge University Press. 553 p.
- QUINN, J. M., A. B. COOPER, R. J. DAVIES-COLLEY, J. C. RUTHERFORD & R. B. WILLIAMSON. 1997. Land use effects on habitat, water quality, periphyton, and benthic invertebrates in Waikato, New Zealand, hillcountry streams. *New Zealand Journal of Marine and Freshwater Research* 31(5): 579-597.
- RAMIREZ, A. & P. E. GUTIÉRREZ-FONSECA. 2014a. Estudios sobre macroinvertebrados acuáticos en América Latina: avances recientes y direcciones futuras. *Revista de Biología Tropical* 62: 9-20.
- RAMIREZ, A. & P. E. GUTIÉRREZ-FONSECA. 2014b. Functional feeding groups of aquatic insect families in Latin America: A critical analysis and review of existing literature. *Revista de Biología Tropical* 62: 155-167.
- RAMIREZ, A., M. ARDÓN, M. DOUGLAS & M. GRAÇA. 2015. Tropical freshwater sciences: An overview of ongoing tropical research. *Freshwater Science* 34(2): 606-608.
- RIBEIRO, A. F., P. R. URBINATTI, A. M. R. DE CASTRO DUARTE, M. B. DE PAULA, D. M. PEREIRA, L. F. MUCCI, A. FERNANDES, M. H. S. H. DE MELLO, M. O. DE MATOS JÚNIOR, R. C. DE OLIVEIRA, D. NATAL & R. DOS SANTOS MALAFRONTE. 2012. Mosquitoes in degraded and preserved areas of the Atlantic Forest and potential for vector-borne disease risk in the municipality of São Paulo, Brazil. *Journal of Vector Ecology* 37(2): 316-324.
- RICO-SÁNCHEZ, A. E., A. J. RODRÍGUEZ-ROMERO, E. LÓPEZ-LÓPEZ & J. E. SEDE-ÑO-DÍAZ. 2014. Patrones de variación espacial y temporal de los macroinvertebrados acuáticos en la Laguna de Tecocomulco, Hidalgo (México). *Revista de Biología Tropical* 62: 81-96.
- RICO-SÁNCHEZ, A. E., A. J. RODRÍGUEZ-ROMERO, J. E. SEDEÑO-DÍAZ, E. LÓPEZ-LÓPEZ & A. SUNDERMANN. 2022. Aquatic macroinvertebrate assemblages in rivers influenced by mining activities. *Scientific Reports* 12(1): 3209.
- SACHS, J. D. 2001. *Tropical underdeveloped*. National Bureau of economic research, Cambridge. 64 p.
- SECRETARÍA DE SALUD. 2016. Infección por virus Zika en México. Disponioble en línea en: https://www.gob.mx/salud/acciones-y-programas/infeccion-por-virus-zika-21776#:~:text=La%20in-

fecci%C3%B3n%20por%20virus%20Zika%20es%20una%20 enfermedad,%28ZIKV%29%20pertenece%20a%20la%20familia%20Flaviviridae%2C%20g%C3%A9nero%20Flavivirus. (consultado el 22 Febrero 2018).

- SECRETARÍA DE SALUD. 2001. Programa de Acción: Enfermedades Transmitidas por vector. Disponible en línea en: http://www.salud.gob.mx/unidades/ cdi/documentos/vectores.pdf (consultado el 22 Febrero 2018).
- SERRANO-BALDERAS, E. C., C. GRAC, L. BERTI-EQUILLE & MA. A. A. HERNÁNDEZ. 2016. Potential application of macroinvertebrates indices in bioassessment of Mexican streams. *Ecological Indicators* 61: 558-567.
- SPRINGER, M., A. RAMÍREZ & P. HANSON. 2010. Macroinvertebrados de Agua Dulce de Costa Rica I. *Revista de Biología Tropical* 58(4).
- STAPLES, J. E. & M. FISCHER. 2014. Chikungunya Virus in the Americas, What a Vectorborne Pathogen Can Do. *The New England Journal of Medicine* 371(10): 887-889.
- STRAYER, D. L. & D. DUDGEON. 2010. Freshwater biodiversity conservation: Recent progress and future challenges. *Journal of the North American Benthological Society* 29(1): 344-358.
- STRAYER, D. L., R. E. BEIGHLEY, L. C. THOMPSON, S. BROOKS, C. NILSSON, G. PINAY & R. J. NAIMAN. 2003. Effects of Land Cover on Stream Ecosystems: Roles of Empirical Models and Scaling Issues. *Ecosystems* 6(5): 407-423.
- SUHLING, F., G. SAHLÉN, S. GORB, V. KALKMAN, K. D. DIJKSTRA & J. VAN TOL. 2015. Order Odonata. *In:* Thorp, J.H & D. C. Rogers (eds.). *Thorp* and Covich's Freshwater Invertebrates. 4th ed. Elsevier, London, pp. 893-932.
- TOWNSEND, C. R., A. G. HILDREW & J. FRANCIS. 1983. Community structure in some southern English streams: The influence of physicochemical factors. *Freshwater Biology* 13(6): 521-544.
- TOWNSEND, C. R., S. DOLÉDEC, R. NORRIS, K. PEACOCK & C. ARBUCKLE. 2003. The influence of scale and geography on relationships between stream community composition and landscape variables: Description and prediction. *Freshwater Biology* 48(5): 768-785.
- WALLACE, J. R. & E.D. WALKER. 2008. Culicidae. *In*: Merritt, R.W., K. W. Cummins & M. B. Berg (eds.). *An introduction to the aquatic insects of North America*. Kendall/Hunt Publishing Company, Dubque, Iowa, pp. 801-823.
- WALSH, C. J., A. H. ROY, J. W. FEMINELLA, P. D. COTTINGHAM, P. M. GROFFMAN & R. P. MORGAN. 2005. The urban stream syndrome: Current knowledge and the search for a cure. *Journal of the North American Benthological Society* 24(3): 706-723.
- WOOD, P. J. & P. D. ARMITAGE. 1997. Biological Effects of Fine Sediment in the Lotic Environment. *Environmental Management* 21(2): 203-217.
- WRIGHT, I. A. & M. M. RYAN. 2016. Impact of mining and industrial pollution on stream macroinvertebrates: Importance of taxonomic resolution, water geochemistry and EPT indices for impact detection. *Hydrobiologia* 772(1): 103-115.
- YAMADA, H. & F. NAKAMURA. 2002. Effect of fine sediment deposition and channel works on periphyton biomass in the Makomanai River, northern Japan. *River Research and Applications* 18(5): 481-493.
- Zar, J. H. 2014. *Biostatistical Analysis.* 5th ed. Pearson, United States of America. 960 p.