





## Absence of concordance between polyembryony and apomixis in maize confirmed through DNA sequencing

### Ausencia de concordancia entre poliembrionía y apomixis en maíz confirmada mediante secuenciación de ADN

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**ABSTRACT.** Maize (*Zea mays* L.) polyembryony is a useful feature for genetic improvement of this specie, not only by its potential to generate multiple plants per seed, but also by its influence on increasing of fatty acids and amino acids content in the grain. It has been considered a possible association between apomixis and polyembryony in maize. With the objective to evidence the relation between apomixes and polyembryony, were used sequences of internal transcribed spacers (ITS), and intergenic spacers (IGS) and amplification of simple repeated sequences (SSR). The analyses were performed in 5 families derived from the IMM-UAAAN-BAP ("D") maize population. Within each of the families were analysed the female parent plant, and two types of progenies (individual and polyembryonic). Nucleotide sequences and genotypic class were compared and also a molecular variation analysis was performed. In these analyses only a close but not identical relationship between polyembryonic plants was found. With the use of these techniques, it was demonstrated that reproduction of the maize plants is of a sexual type, and that based on the molecular markers used, no evidence was obtained about the probable relationship of a common genetic basis between polyembryony and apomixis. Sequencing of the ITS and IGS regions, and use of SSR microsatellites of different chromosomes, was a practical and economical tool for the assessment of similarity between genotypes.

**Key words:** Genetic variation, IGS, ITS, molecular markers, polyembryony, *Zea mays*.

**RESUMEN.** La poliembrionia del maíz (*Zea mays* L.) es una característica útil para el mejoramiento genético, no solo por su potencial para generar múltiples plantas por semilla, sino también por su influencia en el aumento de ácidos grasos y aminoácidos en el grano. Se ha considerado una posible asociación entre apomixis y poliembrionía en maíz. Con el objetivo de evidenciar la relación entre la apomixis y la poliembrionia, se utilizaron las secuencias de espaciadores transcritos internos (ITS), y espaciadores intergénicos (IGS), y la amplificación de secuencias repetidas simples (SSR). Los análisis se realizaron en 5 familias generadas de la población de maíz IMM-UAAAN-BAP ("D"). Dentro de cada una de las familias, se analizaron la planta parental femenina y dos tipos de progenies (individual y poliembriónicas). Se compararon las secuencias de nucleótidos y la clase genotípica y también se realizó un análisis de variación molecular. En estos análisis solo se encontró una relación cercana, pero no idéntica entre las plantas poliembriónicas. Con el uso de estas técnicas se demostró que la reproducción de la planta de maíz es sexual y según los marcadores moleculares utilizados, no se tuvo evidencia sobre la posible relación de una base genética común entre la poliembrionía y la apomixis. La secuenciación de las regiones ITS e IGS, y el uso de microsatélites SSR de diferentes cromosomas, fue una herramienta práctica y económica para la evaluación de la similitud entre genotipos.

**Palabras clave:** IGS, ITS, marcadores moleculares, poliembrionia, variación genética, *Zea mays*.

## INTRODUCTION

Maize genetic diversity is very important and farmers use it to create new varieties (Kato *et al.* 2009). One maize variant that can be used for this purpose is the polyembryony phenomenon (PEm), which was generated through a natural mutation, because of its high productive potential and grain nutritional quality (Castro and Rodríguez 1979). These benefits are obtained under the hypothesis that two or more embryos per seed favors the increase of production, both in the number of plants and ears, allowing to increase grain storage of nutrients of quality because of its greater number of embryos (Espinoza *et al.* 1998, González-Vázquez *et al.* 2011). Polyembryony was discovered by Leewenhoek in 1719 (Batygina and Vinogradova 2007), who observed formation of two seedlings from the same seed in citrus. Moreover, formation of additional embryos may result from differentiation and development of various tissues, zygotes, synergies, antipodes, nucella or tegument (Peter 2009).

The Mexican Maize Institute at Universidad Autónoma Agraria Antonio Narro (IMM-UAAAN) has performed different researches on polyembryony since the 1970s development of two populations that have this characteristic with PEm on the average frequencies from 55 to 65% (González-Vázquez *et al.* 2011, Espinoza-Velázquez *et al.* 2012). The inheritance of this mutant has been identified as the one corresponding to two loci with epistatic interaction of the dominant double type, which segregates in F<sub>2</sub> in a proportion of 15 cases that germinate in individual plant: 1 case of double or higher number of plants (polyembryonic) associated to the phenomenon of incomplete penetrance (Rebolloza-Hernández *et al.* 2011). From this study, it can be assumed that the PEm population could contain some type of apomictic reproduction since polyploidy and polyembryony regularly accompany this type of asexual reproduction. In a study carried out by Avendaño *et al.* (2015) using the IMM-UAAAN polyembryonic populations, it was found similarity between DNA sequences of the internal transcribed spacers (ITS) of the female parent and one of the PEm progenies, which might suggest

a probable relationship between polyembryony and apomixis. If this hypothesis is corroborated in one of the PEm populations, genetic material could be available for studies and special reproductive applications in maize.

Correlation of phenotypes with molecular markers (MM) provides reference points for elucidation of genetic variation in plants (Kumar *et al.* 2014). There are several MM that allow to determine structure, dynamics, relation and genetic variability of a population, and to estimate the degree of divergence between individuals. The most useful MM have been those based on the polymerase chain reaction (PCR) such as amplification of short-sequence DNA repeats (SSR) hypervariable regions, internal transcribed spacers (ITS) and intergenic spacers (IGS), among others (Rodríguez-Tovar *et al.* 2004, Kirst *et al.* 2005). The ITS and IGS regions have been used mainly in cases of biparental inheritance, and are suitable for comparisons between closely related species and genera (Álvarez and Wendel 2003). On the other hand, the SSR have applications in plant genetic studies, such as genetic traces of crops, diversity studies and germplasm selection. The aim of this work was to analyze and validate the genetic similarity, through the use of hypervariable regions and SSR microsatellites among members of maize families with PEm nature to obtain indicators of molecular variation and clustering relationship, with the purpose of obtaining evidence to corroborate the hypothesis of probable relation between apomixis and polyembryony.

## MATERIALS AND METHODS

### Plant material

Genetic material was obtained from four families of S1 inbred lines (D-S<sub>1</sub>) and a family of open pollination (D-PL), derived from the UAAAN-IMM-BAP maize population, which is dwarf (br-2 brachytic gene) with an average of polyembryony of 60% generated in the spring-summer/2010 cycle, in Buenavista, Saltillo, Mexico. Seed samples from three families (D-S<sub>1</sub>-03, D-S<sub>1</sub>-05 and D-PL-13) were germinated during spring/summer-2013 to obtain foliar tissue and DNA

samples. The other two families (D-S<sub>1</sub>-03\* and D-S<sub>1</sub>-07\*) correspond to the remaining DNA sample, which was extracted by Avendaño *et al.* (2015). Each family was composed of the female parent and three progeny plants (one individual, and one pair of polyembryonic PEm plants, these originated from a single seed). The families D-S<sub>1</sub>-03 and D-S<sub>1</sub>-03\* came from the same female parent, sampled at different time. For molecular variance and principal coordinate analyses, as reference material of the common corn (No-PEm), the high inbreeding line (AN-Tep-3) and the Tuxpeño-HOC population (high oil corn) were used because of their genetic distance in relation to the PEm genotypes.

#### DNA isolation and ITS amplification

DNA extraction was performed from leaf tissue of 17-day old PEm maize seedlings using the method described by Doyle and Doyle (1987) with some modifications, such as: addition of 0.5% (v/v) 2-mercaptoethanol to the CTAB extraction buffer. Concentration and quality of DNA was measured in and Epoch<sup>TM</sup> Micro-Volume System Spectrophotometer. Extracted DNA was visualized by electrophoresis in 1% (w/v) agarose gels and stained with ethidium bromide (0.5  $\mu\text{g mL}^{-1}$ ). The ITS region of the 18S-26S ribosomal DNA was amplified to determine genetic similarity using the primers pair: N18L-F (5'-AAGTCGTAACAAGGTTTCCGTAGGTG-3') and C26A-R (5'-TTTCTTTTCTCCGCT-3') (Eldenäs *et al.* 1998). Furthermore, the IGS region of ribosomal DNA was amplified with the primers pair: IGS-F (5'-CTGAACGCCTCTAAGTCAG-3') and IGS-R (5'-GAGACAAGCATATGACTACTG-3') (Kim *et al.* 2001).

Amplification of ITS and IGS regions of the four D-S<sub>1</sub> and one D-PL families was carried out in a total volume of 25  $\mu\text{L}$ , composed of 14.5 and 17.0  $\mu\text{L}$  of ddH<sub>2</sub>O, 1X and 0.4X reaction buffer added with MgCl<sub>2</sub>, respectively, 0.2 mM of dNTPs, 0.8  $\mu\text{M}$  of each primer and 2.0  $\mu\text{L}$  of DNA. PCR amplification was performed in a Px2 Thermal cycler using the following program: 5 min of initial denaturation at 94 °C; 35 cycles of 1 min at 94 °C, 1 min at 50.3 °C for ITS and 51.3 °C for IGS, and 1 min at 72 °C; and a final extension of 5 min at 72 °C. The ampli-

fied products were visualized in 1% (w/v) agarose gels and fragment size was estimated using a 100 pb DNA Ladder (Invitrogen<sup>®</sup>).

Sequence analysis. Sequencing of PCR products were performed on a Perkin Elmer Applied Biosystem Model 3730, using the fluorescence terminator method (Taq FS Dye Terminator Cycle Sequencing Fluorescence-Based Sequencing). Alignments were carried out in BioEdit version 7.1.3 software, and the percentage of invariant sites (monomorphic) and G+C content were determined using DnaSP (DNA sequence polymorphism version 5.10.01) (Librado and Rozas 2009), by pair comparison of individuals in each family including the female parent plant and three descendents (one individual and a pair of polyembryonic plants PEm).

#### SSR analysis

In the SSR analysis, ten maize microsatellite loci, one of each chromosome, were obtained from the MaizeGDB database (Table 1). Each locus was selected because of the high PIC (polymorphism information content) presented in a study by Warburton *et al.* (2002). Reactions were performed in a cocktail composed as following: 1.5 to 3X buffer with MgCl<sub>2</sub>, 0.25 mM of dNTPs, 0.66 to 2  $\mu\text{M}$  of each primer, 6.6% (v/v) of DMSO. 0.1 U of PaqDNA Polymerase, 1.5  $\mu\text{L}$  of DNA and ddH<sub>2</sub>O up to a final volume of 15  $\mu\text{L}$ . Warburton *et al.* 2002 described the reference amplification protocol with an initial denaturation step of 2 min at 94 °C; 30 cycles of 30 s at 94 °C, 1 min of annealing (Table 1) and 1 min of polymeration at 72 °C; followed of final extension step at 72 °C for 5 min.

#### Data analyses

The amplification pattern of PCR products was transformed into a binary matrix, where 1 indicates presence and 0 = absence. From these results, the homozygous and heterozygous condition of each loci was determined. Pattern of similarity was sought between the female parent plant and each progenie seedling, one of the individual type and the polyembryonic pair (PEm). An analysis of molecular variance (AMOVA) was performed, where five groups were considered, four belonging to each of the PEm fami-

**Table 1.** Simple Sequence Repeats (SSR) markers used in present study, with their genomic position, annealing temperature and sequence.

Primer name	Bin†	PIC††	Alignment temperature (°C)	Sequence (5' → 3')
phi064 f				CCGAATTGAAATAGCTGCGAGAACCT
phi064 r	1.11	0.79	56.0	ACAATGAACGGTGGTTATCAACACGC
phi96100 f				AGGAGGACCCCAACTCCTG
phi96100 r	2.0	0.75	50.0	TTGCACGAGCCATCGTAT
phi053 f				CTGCCTCTCAGATTCAGAGATTGAC
phi053 r	3.05	0.70	60.0	AACCCAACGTACTCCGGCAG
phi072 f				ACCGTGCATGATTAATTTCTCCAGCCTT
phi072 r	4.01	0.59	56.0	GACAGCGCGCAAATGGATTGAACT
zct118 f				CTTCCAGCCGCAACCCTC
zct118 r	5.07	0.76	56.0	CCAACAACGCGGACGTGA
phi452693 f				CAAGTGCTCCGAGATCTTCCA
phi452693 r	6.06	0.57	52.4	CGCGAACATATTGAGAAGTTTG
phi328175 f				GGGAAGTGCTCCTTGACG
phi328175 r	7.04	0.67	60.0	CGGTAGGTGAACGCGGTA
phi420701 f				ATGTTTCAAACCACCCAGA
phi420701 r	8.01	0.56	60.0	ATGGCAGCAATAGCAACAGG
phi032 f				CTCCAGCAAGTGATGCGTGAC
phi032 r	9.04	0.48	56.0	GACACCCGGATCAATGATGGAAC
umc1152 f				CCGAAGATAACCAACAATAATAGTAGG
umc1152 r	10.01	0.74	60.0	ACTGTACGCCTCCCCTTCTC

†Bin indicates the chromosomal location.

††PIC is polymorphic information content.

lies (D-S<sub>1</sub>-03, D-PL-13, D-S<sub>1</sub>-03\* and D-S<sub>1</sub>-07\*) and the fifth group corresponding to the reference material. A Principal Coordinate Analysis (PCoA) and a dendrogram were performed in order to observe the variability and to group the studied genotypes. All the statistical analyzes were carried out using INFOGEN software (v. 2011).

## RESULTS

### Comparison of the hypervariable regions nucleotide sequences in families D-S<sub>1</sub> (first self-fertilization), and D-PL (open pollination)

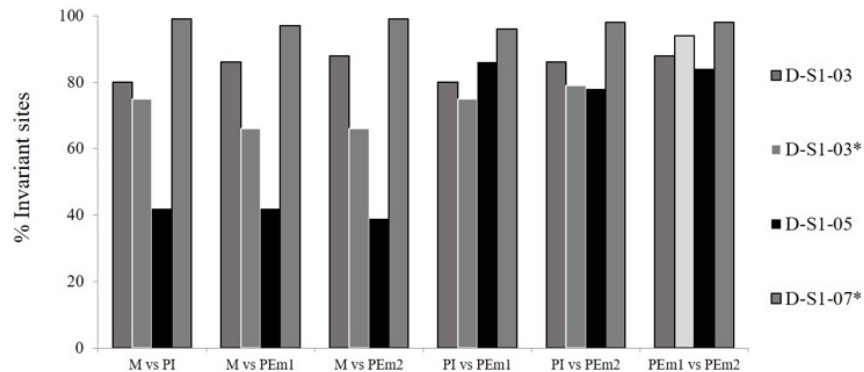
Frequencies invariable sites in the ITS region of D-S<sub>1</sub> families did not show cases of identical sequences in the six possible comparisons per maize family, but the D-S<sub>1</sub>-07\* family had very high similarity values (96 to 99%). D-S<sub>1</sub>-05 Family showed the highest variability in the sites, indicating absence of identical sequences among its members, being discarded for the rest of analyzes. In the series of comparisons of D-S<sub>1</sub>-03 and D-S<sub>1</sub>-03\* families, the highest percentage of similarity was showed between the pair of polyembryonic sister plants (PEm1 and PEm2). In general, the families analyzed had widely

divergent values, from 39 to 99%, where the highest values were mostly for the comparison between PEm sisters plants, and not for the female parent (M) with one of the PEm daughters (Figure 1), which is a contrary condition from a previous report. As in the ITS region of the D-S<sub>1</sub>-03 and D-S<sub>1</sub>-03\* families, the highest value of monomorphic sites in the IGS region of the D-S<sub>1</sub>-07\* family was presented in comparison between the pair of polyembryonic sister plants (PEm1 and PEm2) (Figure 2).

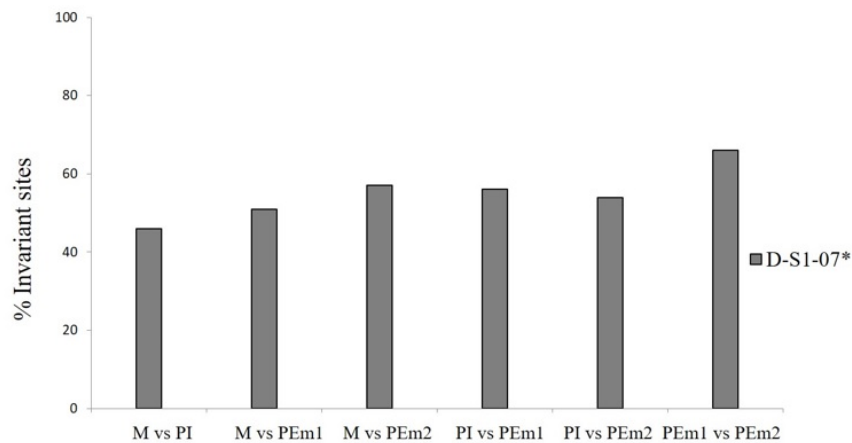
The D-PL-13 family, generated by open-pollination (PL) showed high percentages of monomorphic sites in the six comparisons of the ITS region (Figure 3). However, the comparison analysis of the IGS region in the genotypes of this family were contrary to those obtained in the ITS analysis (Figure 4). Morphology (phenotype) of the polyembryonic seedlings corresponding to the D-PL-13 family consisted of two mesocotyls and two completely separate radicles.

### SSR analysis

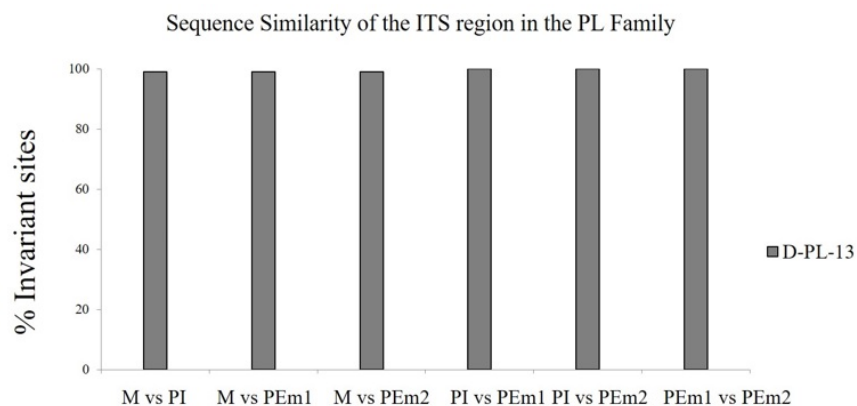
The genotypes included in this study were the two families that showed high sequence similarity (99-100%) in the ITS region. Results of the similarity ana-



**Figure 1.** Percentage of invariant sites in ITS region sequence in D-S<sub>1</sub> families. M: female parent plant, PEm: each of the two polyembryonic plant, PI: individual plant.

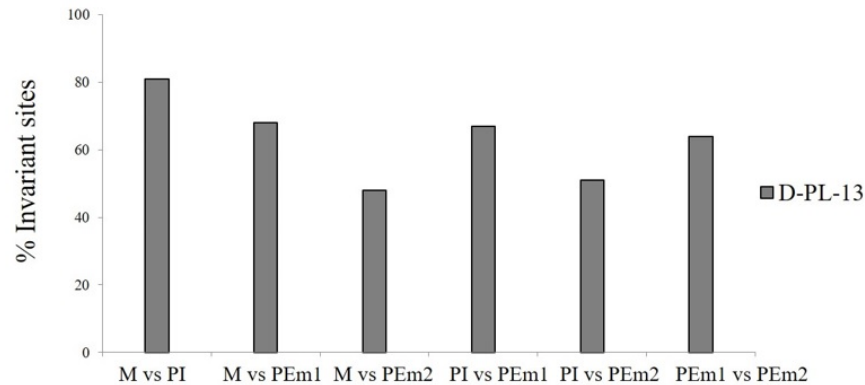


**Figure 2.** Percentage of invariant sites of IGS region sequence in D-S<sub>1</sub>-07\* family. M: female parent plant, PEm: polyembryonic plant, PI: individual plant.



**Figure 3.** Percentage of invariant sites in ITS region sequence of D-PL-13 family. M: female parent plant, PEm: polyembryonic plant.

### Sequence Similarity of IGS Region in the PL Family



**Figure 4.** Percentage of invariant sites in IGS region sequence of D-PL-13 family. M: female parent plant, PEm: polyembryonic plant, PI: individual plant.

lysis in those families are presented in Table 2. Families with the highest loci similarity were D-S<sub>1</sub>-03, D-S<sub>1</sub>-03\* and D-PL-13. In the D-S<sub>1</sub>-03 family, the female parent plant and the individual daughter showed 100% homozygous loci, likewise in the D-S<sub>1</sub>-03\* family, comparisons among the three sister plants (the possible comparisons among PI, PEm1 and PEm2) presented 100% homozygous loci, and the female parent plant only 67%. Similarly, the D-PL-13 family showed high similarity (100%) in only the two polyembryonic sister plants (PEm1 and PEm2), and the percentage of homozygous loci of the female parent plant was equal or less to 50%.

**Table 2.** Similarity analysis of homozygous and heterozygous loci in maize families, within the context of polyembryony.

Comparisons	D-S <sub>1</sub> -03 (%)	D-PL-13 (%)	(Avendaño <i>et al</i> 2015)	
			D-S <sub>1</sub> 03* (%)	D-S <sub>1</sub> -07* (%)
M vs PI	100	25	67	50
M vs PEm1	83	50	67	67
M vs PEm2	83	50	67	83
PI vs PEm1	83	50	100	83
PI vs PEm2	83	50	100	67
PEm1 vs PEm2	83	100	100	83

M: mother plant, PEm: polyembryonic plant, PI: individual plant.

Results of the molecular variance analysis (AMOVA) showed significant differences ( $p \leq 0.01$ ) among the five analyzed groups (Table 3). The highest percentage of genetic variation was due to

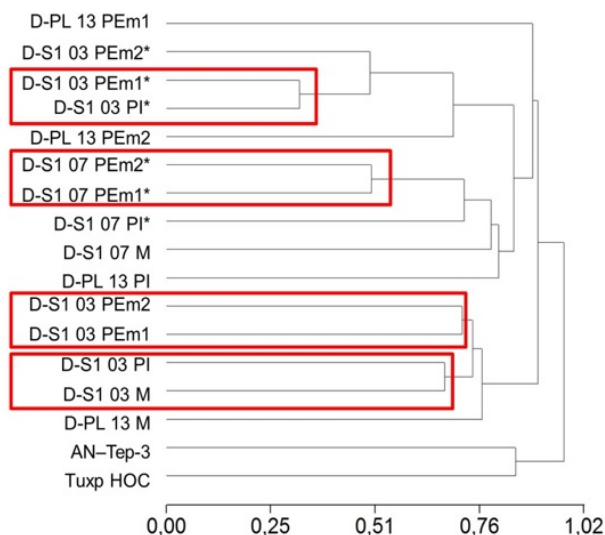
difference within members of each group, and the rest was due to differences among groups. The ordering of polyembryonic and exotic maize samples that yielded the Principal Coordinate Analysis (PCoA), performed with the Anderberg distance (cophenetic correlation of 0.966), is presented in Figure 5. The ordering consisted of formation of four groups, one containing the AN-Tep-3 and Tuxpeño HOC genotypes, both representing common or normal maize genotypes, and genetically distant from PEm-like families. The second group consisted of the D-S<sub>1</sub>-03 family and a member of the D-PL-13; the third group included the D-S<sub>1</sub>-07\* family and a member of DPL-13 family and finally, the fourth group consisted of the D-S<sub>1</sub>-03\* family and the rest of the individuals in the D-PL-13 family.

The dendrogram to identify the grouping of the evaluated genotypes was obtained with the UP-GMA method (Unweighted Pair-Group Methods with Arithmetic Mean), which is shown in Figure 6. The most remote group corresponded to the AN-Tep-3 and Tuxpeño HOC genotypes. The second subgroup was formed with the two polyembryonic plants of the same family (D-S<sub>1</sub>-03 PEm1 and D-S<sub>1</sub>-03 PEm2). In the third group, two polyembryonic plants of D-S<sub>1</sub>-07 family were found to form a subgroup (D-S<sub>1</sub>-07 PEm1\* and D-S<sub>1</sub>-07 PEm2\*), with a distance of 0.50 and in the fourth group, a subgroup with a distance of

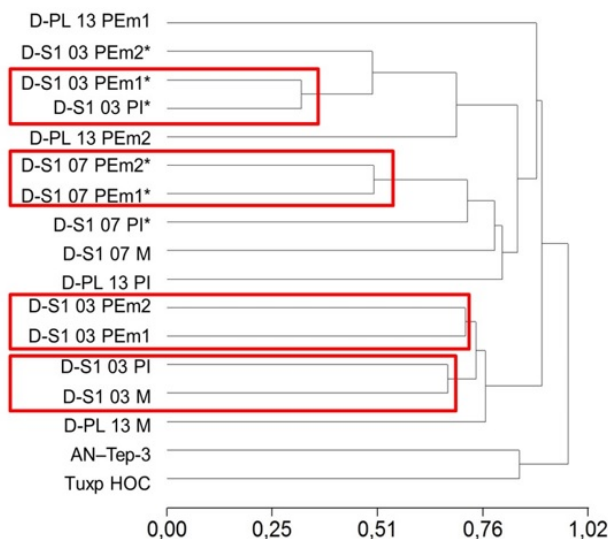


**Table 3.** Analysis of Molecular Variance (AMOVA) within and between polyembryonic and exotic families.

Variation source	DF	SS	MS	Variation Percentage	p-value	Number of Iterations
Groups	4	95.1	23.78	26.2	0.0075	400
Within groups	13	138.0	10.6	73.8	0.0025	400
Total	17	233.1	13.71			



**Figure 5.** Principal Coordinate Analysis of polyembryonic and exotic genotypes. M: female parent plant, PEm: polyembryonic plant, PI: individual plant, \*: material generated by Avendaño *et al.* (2015), AN-Tep-3 y Tuxp HOC: exotic materials Tepalcingo and Tuxpeño, respectively.



**Figure 6.** Dendrogram of polyembryonic and exotic maize genotypes based on Anderberg distance measurements. M: female parent plant, PEm: polyembryonic plant, PI: individual plant, \*: material generated by Avendaño *et al.* (2015), AN-Tep-3 and Tuxp HOC: exotic materials Tepalcingo and Tuxpeño, respectively.

0.32 between a single plant and a polyembryonic (D-S<sub>1</sub>-03 PI\* and D-S<sub>1</sub>-03 PEm1\*) was formed, the genotypes being considered more closely related.

## DISCUSSION

The high levels of similarity among members comparison of the D-S<sub>1</sub>-07\* family in the ITS region suggested the possibility of corroborating the results with an analysis of the IGS region; however, application of the analysis showed significant divergences (Figure 2), with values well below those obtained in the ITS region. A possible explanation is the rapid evolution of these regions, where it is common manifestation of polymorphism (Zhou *et al.* 1996). In a review by Willson and Burley (1983), referred to simple polyembryony as the ability of the female gametophyte to present more than one egg cell with the potential to produce an embryo after being fertilized by the same or different male, that will depend on the source of pollen grains (Tudge 2006). With the values obtained in the present research for monomorphic sites in the IGS region, it can be supported the fact that PEm1 and PEm2 plants are not considered as identical, and therefore not come from the same origin, even in the case when pollination is controlled and the degree of similarity could come from the accumulation of homozygous genes generated by self-fertilization in an expected proportion of 50%, as is the case of the S<sub>1</sub> families.

High percentages of monomorphic sites in the ITS region of an open-pollination (PL) generated family were found. That is to say, as if the nucleotide sequence of each of the members, female parent plant and its progenies, were the same and some type of asexual reproduction (apomixis) could exist since the progeny, product of open pollination does not contain the forced accumulation of gene pairs in homozygous by effect of self-fertilization, a condition that, when observed, would increase the likelihood of similarity, at least among the DNAs of sister plants. But, it can be concluded that existence of some type of apomictic reproduction is not confirmed according to comparison analysis of the IGS region.

Morphology of polyembryonic seedling was

similar to one of the three polyembryonic origin versions described by Erdelská (1996), where multiple individual embryos were produced closely together, but separated by an epidermis, with independent plumules and radicles, generated by the phenomenon called polyspermy, where the embryos originate from the fertilization of multiple egg cells. One of the probable causes that may inflict the sequence similarity in the ITS region in this study could be due to similar genetic condition of the pollen source, with common genetic basis. In addition, the percentage of G+C content in all the studied families was presented in a range of 46 to 76%, which coincides with that reported for maize composition, in a range from 45 to 75% (Carels and Bernardi 2000).

Regarding SSR analysis, homozygous *loci* increased by the reduction of heterozygous sites because of self-fertilization, as was in the case of three S<sub>1</sub> families. On the other hand, it was expected that in the genotypes with open pollination (PL), the progeny would present the majority of *loci* in heterozygous condition with respect to the female parent plant. However, this was not observed in the D-PL-13 family, which may be explained because families derived from the D population contained a common genetic base. Moreover, asexual reproduction is strongly correlated with hybridization, since most species that are apomictic emerged from the hybridization of two genomes, having heterozygous *loci* and, because of epigenetic changes, heterozygous was perpetuated (Beck *et al.* 2012, Lovell *et al.* 2013).

According to the molecular variance analysis, higher genetic variation was found within members of the five analysed groups. In a study by Ramakrishnan *et al.* (2014) in bud explants of 17 maize genotypes, they determined an amount of variation of 93% within genotypes and 3% between genotypes, concluding that the female parent and the seedlings generated *in vitro* have a 100% genetic similarity. Based on the results of this study, it is possible to indicate that there was variation among members of the families, and even more in the group that corresponded to the exotic genotypes, corresponding to the open-pollinated Tuxpeño HOC population and the inbred line AN-Tep-3, which correspond to the



common maize type, or Non-PEm.

In the Principal Coordinate Analysis, the D-S<sub>1</sub>-03 and D-S<sub>1</sub>-07\* families had a better ordering, which was consistent with the close relationship of the percentages of invariant sites in the ITS region. On the other hand, the individuals that were less ordered were those of the D-PL-13 family, which was originated by open pollination. Results of the three performed analyzes in the D-S<sub>1</sub>-03 and D-S<sub>1</sub>-07 families, either PCoA, dendrogram or sequence comparison of the ITS region, coincided in detecting the close relationship that exists between the members of the families and even more so between the two polyembryonic sister plants (PEm1 and PEm2). On the other hand, the D-PL-13 family corroborated the divergence among the members that formed it, even between the two PEm plants generated in the same seed, a result that suggests that this pair of plants, initially considered as “twins”, differ in some degree in their genetic condition, that is, they are non-identical sisters.

## CONCLUSIONS

The use of molecular techniques applied in this research to genotypes derived from UAAAN-IMM-BAP maize population allowed to demonstrate that reproduction of the studied maize plants is of a sexual

type, and that based on the molecular markers used, no evidence was obtained about the probable relationship of a common genetic basis between polyembryony and apomixis. The high similarity detected in the analysis of the ITS hyper-variable regions is not sufficient evidence to support an identical genetic condition between the female parent plant and one of the PEm progenies, and even among the PEm sister plants pair. However, the highest but not complete similarity was observed in all cases between the pair of polyembryonic plants, emerged from a single seed. Sequencing of the ITS and IGS regions, as well as the use of SSR microsatellites of different chromosomes, was a practical and economical tool for the assessment of similarity between genotypes.

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## LITERATURE CITED

- Álvarez I, Wendel JF (2003) Ribosomal ITS sequences and plant phylogenetic inference. *Molecular Phylogenetic and Evolution* 29: 417-434
- Avendaño M, Espinoza J, Gutiérrez A, Flores A, Rodríguez R (2015) Secuencias nucleotídicas de la región ITS en familias S 1 y PL de maíces poliembriónicos. *Revista Mexicana de Ciencias Agrícolas* 6: 509-521
- Batygina TB, Vinogradova GY (2007) Phenomenon of polyembryony. Genetic heterogeneity of seeds. *Russian Journal of Developmental Biology* 38: 126-151
- Beck JB, Alexander PJ, Allphin L, Al-Shehbaz IA, Rushworth C, Bailey CD, *et al.* (2012) Does hybridization drive the transition to asexuality in diploid *Boechera*? *Evolution* 66: 985-995
- Carels N, Bernardi G (2000) Two classes of genes in plants. *Genetics* 154: 1819-1825
- Castro GM, Rodríguez HS (1979) Estudio preliminar del potencial de plantas de maíz con tallos gemelos. *Fitotecnia* 2: 39-49.
- Doyle JJ, Doyle JL (1987) A rapid DNA isolation procedure for small quantities of fresh leaf tissue. *Phytochemical Bulletin* 19: 11-15

- Eldenäs P, Anderberg AA, Källersjö M (1998) Molecular phylogenetics of the tribe Inuleae s. str. (Asteraceae), based on ITS sequences of nuclear ribosomal DNA. *Plant System Evolution* 210: 159-173
- Erdelská O (1996) Polyembryony in maize: histological analysis. *Acta Societatis Botanicorum Poloniae* 65: 123-125
- Espinoza J, Vega MC, Navarro E, Burciaga G (1998) Poliembrionía en maíces de porte normal y enano. *Agronomía Mesoamericana* 9: 83-88
- Espinoza-Velázquez J, Valdés-Reyna J, Alcalá-Rodríguez JM (2012) Morfología y anatomía de radículas múltiples en plántulas de maíz derivadas de cariopsis con poliembrionía. *Polibotánica* 33: 207-221
- González-Vázquez V, Espinoza-Velázquez J, Mendoza-Villareal R, De León-Castillo H, Torres-Tapia MA (2011) Caracterización de germoplasma de maíz que combina un alto contenido de aceite y poliembrionía. *Universidad y Ciencia* 27: 157-167
- Kato TA, Mapes C, Mera LM, Serratos JA, Bye RA (2009) Origen y diversificación del maíz: una revisión analítica. Universidad Nacional Autónoma de México, Comisión Nacional para el Conocimiento y Uso de la Biodiversidad. México. 116p.
- Kim HJ, Choi YK, Min B (2001) Variation of the Intergenic Spacer (IGS) Region of Ribosomal DNA among *Fusarium oxysporum* formae speciales. *Journal of Microbiology* 39: 265-272
- Kirst M, Cordeiro CM, Rezende GDSP, Grattapaglia D (2005) Power of Microsatellite Markers for Fingerprinting and Parentage Analysis in *Eucalyptus grandis* Breeding Populations. *Journal of Heredity* 96: 161-166
- Kumar V, Malik SK, Pal D, Srinivasan R, Bhat SR (2014) Comparative transcriptome analysis of ovules reveals stress related genes associated with nucellar polyembryony in citrus. *Tree Genetics and Genomes* 10: 449-464
- Librado P, Rozas J (2009) DnaSP v5: a software for comprehensive analysis of DNA polymorphism data. *BMC Bioinformatics* 25: 1451-1452
- Lovell JT, Aliyu OM, Mau M, Schranz ME, Koch M, Kiefer C, et al. (2013) On the origin and evolution of apomixis in *Boechera*. *Plant Reproduction* 26: 309-315
- Peter KV (2009) Basics of horticulture. New India Publishing Agency. Pitan Pura, New Delhi, India. 762p.
- Ramakrishnan M, Ceasar SA, Duraipandiyar V, Ignacimuthu S (2014) Efficient plant regeneration from shoot apex explants of maize (*Zea mays*) and analysis of genetic fidelity of regenerated plants by ISSR markers. *Plant Cell Tissue and Organ Culture* 119: 183-196
- Rebolloza-Hernández H, Espinoza-Velázquez J, Sámano-Garduño D, Zamora-Villa VM (2011) Herencia de la poliembrionía en dos poblaciones experimentales de maíz. *Revista Fitotecnia Mexicana* 34: 27-33
- Rodríguez-Tovar A, Xoconostle-Cásarez B, Valdés M (2004) Ecología molecular de los hongos ectomicorrízicos. *Revista Fitotecnia Mexicana* 27: 267-278
- Tudge C (2006) The secret life of trees: How they live and why they matter. Penguin Books. London England 464p.
- Warburton ML, Xianchun X, Crossa J, Franco J, Melchinger AE, Frisch M, et al. (2002) Genetic characterization of CIMMYT inbred maize lines and open pollinated populations using large scale fingerprinting methods. *Crop Science* 42: 1832-1840
- Willson M, Burley N (1983) Mate choice in plants: Tactics, mechanisms, and consequences. Princeton University Press, Princeton, USA. 175p.

Zhou Y, Zou YP, Hong DY, Zhou J, Chen SY (1996) ITS1 Sequences of Nuclear Ribosomal DNA in Wild Rices and Cultivated Rices of China and Their Phylogenetic Implications. *Acta Botanica Sinica* 38: 785-791

