

# Bioaccumulation and changes in the photosynthetic apparatus of *Prosopis juliflora* exposed to copper

CLAUDIA YARED MICHEL-LOPEZ<sup>1</sup>, FRANCISCO ESPADA-Y GIL<sup>2</sup>, GABRIELA FUENTES ORTÍZ<sup>2</sup>, JORGE M. SANTAMARÍA<sup>2</sup> Y DANIEL GONZÁLEZ-MENDOZA<sup>1,3</sup>

Botanical Sciences  
94 (2): 323-330, 2016

DOI: 10.17129/botsci.507

## Abstract

The effect of copper toxicity on the photosynthetic activities and bioaccumulation in different tissues of *Prosopis juliflora* was investigated using three CuSO<sub>4</sub> concentrations (10, 50 and 100 mM) added under hydroponic conditions. Copper concentration and chlorophyll fluorescence were measured after 48 h of copper stress. The results obtained in this study show that increasing levels of 50 and 100 mM Cu<sup>2+</sup> resulted in a significant accumulation of this metal mainly in roots compared with control roots. On the other hand, our result showed a significant reduction of maximum photochemical efficiency of photosystem II ( $F_v/F_m$ ) and the activity of photosystem II ( $F_v/F_0$ ) ratios in *P. juliflora* leaf treated with 100 mM Cu<sup>2+</sup> with respect to control after 4h of exposure. These changes suggested that the photosynthetic apparatus of *P. juliflora* was the primary target of the Cu<sup>2+</sup> action. Therefore the information provided by this short-term (48 h) experiment in *P. juliflora* showed that several physiological processes are activated, in which the copper uptake by roots and their accumulation in tissues play a central role. In conclusion, the chlorophyll fluorescence parameters can be used as a useful physiological tool to assess early changes in photosynthetic performance of *P. juliflora* in response to copper pollution in short-term. Finally, the present study showed that *P. juliflora* is a promising prospect for heavy metals phytoremediation purposes occurring in arid and semi-arid climates in the northwest Mexico.

**Keywords:** bioaccumulation, chlorophyll fluorescence *a*, heavy metals, *Prosopis juliflora*

## Bioacumulación y cambios en aparato fotosintético de *Prosopis juliflora* expuesta a cobre

### Resumen

Los efectos de la toxicidad de cobre en la actividad fotosintética y bioacumulación en diferentes tejidos de *Prosopis juliflora* fue evaluada usando tres concentraciones de CuSO<sub>4</sub> (10, 50 y 100 mM) en condiciones hidropónicas por 48 h. Los resultados obtenidos indicaron que los niveles crecientes del metal generaron una acumulación significativa de Cu<sup>2+</sup>, principalmente en las raíces en comparación con las raíces del control. Por otro lado, nuestros resultados indicaron una reducción significativa de la máxima eficiencia fotoquímica del fotosistema II ( $F_v / F_m$ ) y la actividad de los coeficientes de fotosistema II ( $F_v / F_0$ ) en hojas *P. juliflora* tratada con 100 mM de Cu<sup>2+</sup> con respecto al control después de 4 h de la exposición. Estos cambios sugieren que el aparato fotosintético de *P. juliflora* es fuertemente afectado por el Cu<sup>2+</sup>. Por lo tanto, la información proporcionada por este experimento a corto plazo (48 h) en *P. juliflora* mostró que varios procesos fisiológicos se activan, en el que la absorción por las raíces del metal y su acumulación en los tejidos juegan un papel central. En conclusión, los parámetros de fluorescencia de clorofila pueden ser utilizados como una herramienta fisiológica útil para evaluar los cambios tempranos en la eficiencia fotosintética de *P. juliflora* en respuesta a la contaminación de cobre en periodos cortos de tiempo. Finalmente, el presente estudio muestra que *P. juliflora* es un prospecto promisorio para propósitos de fitoremediación en climas áridos y semiáridos en el noroeste de México.

**Palabras claves:** bioacumulación, fluorescencia de la clorofila *a*, metales pesados, *Prosopis juliflora*

<sup>1</sup> Instituto de Ciencias Agrícolas, Universidad Autónoma de Baja California (ICA-UABC), Carretera a Delta s/n Ejido Nuevo León, 21705, Baja California, México

<sup>2</sup> Unidad de Biotecnología, CICY, Mérida, Yucatán, México

<sup>3</sup> Corresponding author: daniel-asaf@gmail.com

The Mexicali Valley is located in northeastern Baja California, south of the Imperial Valley in California. Approximately 70 % of the cultivated land in Mexicali is irrigated by gravity flow with water from the Colorado River (Daesslé *et al.*, 2009). In recent years the Mexicali industrial unit such as piping steel, paint making, agriculture, paper mill, fish cultivation, electroplating industries drain their wastewater into the river (García-Hernández *et al.*, 2006). As a result of these emissions and the fact that metals are non-biodegradable, heavy metals accumulate in soils affecting the environment, leading to a potential toxic exposure of the population (Wong *et al.*, 2006). In recent years, a variety of techniques have been employed to clean up the soils and effluents. These methods include ion exchange, chemical precipitation, and disinfection, adsorption by activated carbons, reverse osmosis and nanofiltration (Machado *et al.*, 2010). However, these methods are expensive, require high energy and are not able to completely remove the heavy metals. In contrast phytoremediation is proposed as a cost effective alternative for the treatment of contaminated soils (Alkorta *et al.*, 2004). Many plant species are able to grow under heavy metals polluted environments (González-Mendoza and Zapata-Perez, 2008).

The genus *Prosopis* L. is characteristic of arid and semiarid zones, and its widespread distribution includes ecosystems in Asia, Africa, and the Americas (Pérez-García *et al.*, 2012). The species of this genus are characterized by high individual genetic variability at the population level, which influences the responsiveness of these species to extrinsic variables, such as climate, drought, alkalinity and salinity (Shiferaw *et al.*, 2004). In Mexico, the majority of the populations of the genus *Prosopis* are found in the northern and central of the country, where they have formed forest extensions that are adapted to desert climate.

Species such as *Prosopis glandulosa* Torr., *P. juliflora* (Sw.) DC., *P. pubescens* Benth. and *P. palmeri* S. Watson dominate the vegetation mosaic in Baja California Mexico, primarily in an ecosystem known as mezquites, which is characterized by a hyperarid climate with almost no rainfall (0.6 mm year<sup>-1</sup>) (Carevic, 2014). These populations of the genus *Prosopis* have been the focus of scientific interest mainly because of their physiological and ecological adaptations to their hyperarid environment. However, little research has analyzed the current physiological tolerance to heavy metals of *P. juliflora*. The few previous studies of *P. juliflora* have focused mainly on the evaluation of ability of these plants to germinate in different copper concentrations (Victor *et al.*, 2007). However, previous studies using *P. juliflora* foliar disc have demonstrated that the uptake of copper in excess by plants can initiate a disorder of numerous physiological functions causing damage at the cellular level (Michel-López *et al.*, 2014). Therefore a better understanding of the strategies that are used by individuals of *P. juliflora* for copper tolerance will help researchers to develop biotechnological alternatives as the phytoremediation in northwest of Mexico. In this context, the main objective of this research was evaluated the physiological responses of *P. juliflora* to copper an essential element. This assessment was accompanied by analyzing the influence of different concentrations of copper on the photosynthetic status and bioaccumulation of this metal in *P. juliflora* plants in a short time. We presume that individuals of *P. juliflora* denote a high accumulation of copper in response to stress caused by acute exposition to copper.

## Materials and methods

**Germination of *Prosopis juliflora*.** The seeds were donate for the National Forestry Commission of Mexico, in Mexicali, B.C. Seeds were surface-sterilized by soaking in 1 % NaOCl (Clorox) for 5 min and finally rinsed with deionized sterile water four times altogether (for ensuring pathogen-free seed) and dried at room temperature. After disinfection, the seeds were germinated in sterilized sand (grain size 6 to 9 mm), in a greenhouse under the following conditions: temperature range 30 to 34 °C during the day, 28 to 30 °C during the night, 12 h light: dark photoperiods and 60 % relative air humidity. Once the seedlings developed a few roots, they were transplanted to single pots (0.5 L) containing a commercial potting soil mix combined with quartz sand and peat moss (5 % soil, 20 % sand and 30 % peat moss) previously sterilized in hot water (100 °C), during 2 h. The plants were irrigated daily with water and every other week, fertilized with Hoagland solution without additional Cu<sup>2+</sup>.

**Exposure to Cu<sup>2+</sup>.** Twenty two-months old plants showing similar leaf number and size were randomly allocated (*n* = 5) and transferred to individual plastic containers holding 500 mL of

deionized water solution with or without 10, 50 and 100 mM of copper sulfate ( $\text{CuSO}_4$ ) at pH: 5.5 for 48 h under hydroponic conditions. After 48 h plants were uprooted from the plastic containers and aerial parts (leaves and stems) and roots were collected for further analysis. These exposures were performed in quadruplicates.

**Copper analysis.** To remove residual copper solution, root samples were washed for 5 min in deionized water, then for 10 min in 20 mM EDTA, and again rinsed for 5 min in deionized water. Five gram (fresh weight) of leaves, stem, and roots was dried for 72 h at 70 °C. Sample of dried tissue (500 mg) was digested in 10 mL of nitric acid (85 % v/v) overnight according to González-Mendoza *et al.* (2007).

Resulting digests were diluted up to 10 mL with deionized water and  $\text{Cu}^{2+}$  concentration was determined for each sample by an inductively coupled plasma optical emission spectrophotometer (ICP-OES 400 Perkin-Elmer USA) using standards of 50, 20, 5, 0.5  $\text{mg L}^{-1}$  of  $\text{Cu}^{2+}$  (Sigma Chem., St. Louis, MO, USA) for calibration curves and readings were evaluated at 324.8 nm absorbance against 1 %  $\text{HNO}_3$  (Estrella-Gómez *et al.*, 2009). The concentration of  $\text{Cu}^{2+}$  in plant tissues was expressed in  $\mu\text{g g}^{-1}$  on a dry weight (dw) basis.

**Bioaccumulation factor and translocation factor.** The index of the ability of the plant to accumulate copper with respect to its concentration in the medium, bioaccumulation factor (BAF), was calculated according to Ghosh and Singh, (2005). The relative translocation of copper from roots to other parts of the plants, the translocation index ( $T_i$ ), was calculated according to Singh *et al.* (2011).

**Chlorophyll fluorescence measurement.** The chlorophyll fluorescence was measured using a Plant Efficiency Analyser (PEA, Hansatech Instruments Ltd., King's Lynn Norfolk PE32 1JL, UK) on completely expanded leaves. The leaves were subjected to a 5 min period of adaptation to darkness under to induce the complete oxidation of the reaction centers using light exclusion clips (González-Mendoza *et al.*, 2013).

The maximum photochemical efficiency of photosystem II ( $F_v/F_m$ ), the activity of photosystem II ( $F_v/F_o$ ) and the efficiency of the water splitting apparatus represent by ( $F_o/F_v$ ) were measurements of five plants for each treatment, at 8, 12, 24, and 48 h after exposure to copper levels using a leaf one per plant analyzed. The plants exposed to the copper were arranged in a complete randomized design, with four replications and five plants per plot.

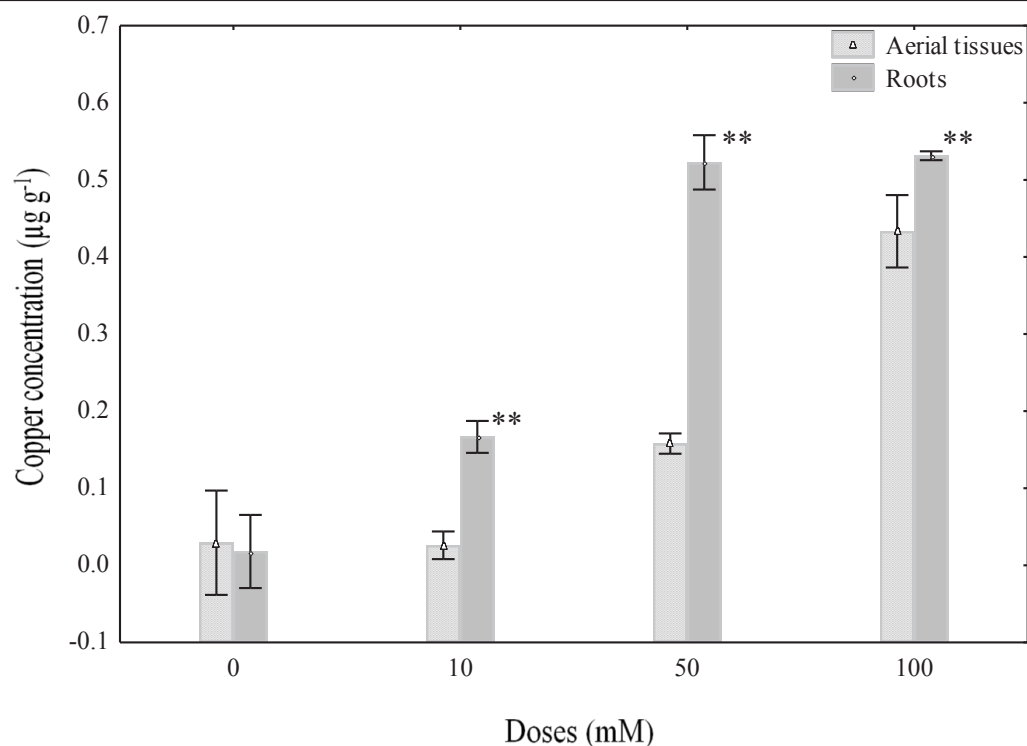
**Statistical analysis.** Data were analyzed with analyses of variance (ANOVA), and mean were comparison test (Tukey's  $\alpha = 0.05$ ) was performed (Statistical Package version 5.5, Statsoft, USA). Significant differences were accepted if  $P < 0.05$  and data was expressed as mean  $\pm$  Standard error.

## Results

**$\text{Cu}^{2+}$  concentration in different tissues of *Prosopis juliflora*.** In the present study *P. juliflora* plants grew well in the presence of 10, 50 and 100 mM  $\text{Cu}^{+2}$ , and no showed physiological changes (eg., discoloration of leaves) associated at heavy metal exposure. However, the  $\text{Cu}^{2+}$  accumulation on roots and aerial parts of *P. juliflora* varied with the different concentrations of copper sulphate solutions used in this experiment (Figure 1). In this sense 10, 50 and 100 mM  $\text{Cu}^{2+}$  doses caused significant changes in  $\text{Cu}^{2+}$  accumulation on roots and aerial parts compared with control roots. For example, the accumulation of  $\text{Cu}^{+2}$  in roots plants treated with 50 mM  $\text{Cu}^{2+}$  ( $0.522 \mu\text{g g}^{-1}$ ) did not show significantly change with plants treated with 100 mM  $\text{Cu}^{2+}$  ( $0.531 \mu\text{g g}^{-1}$ ) after 48 h of exposure. However, in case of aerial parts, the exposure at 10, 50 and 100 mM  $\text{Cu}^{2+}$  showed a gradual accumulation of copper in *P. juliflora* with respect to doses used. In this regard, the figure 1 show that the ranges of  $\text{Cu}^{+2}$  accumulation were 0.026, 0.158 and  $0.433 \mu\text{g g}^{-1}$  in aerial parts, exposed to 10, 50 and 100 mM, respectively after exposure.

**Bioaccumulation factor and translocation index of  $\text{Cu}^{+2}$ .** Bioaccumulation factor (BAF) and translocation index ( $T_i$ ) was significantly higher in root than that of aerial parts (stem and leaves) ( $P < 0.05$ ; Table 1). However, its BAF was significantly less in roots of plant grown to 100 mM ( $66.41 \pm 0.28$ ) with respect to 10 ( $208.33 \pm 9.01$ ) and 50 mM ( $130.66 \pm 3.54$ ), respectively (Table 1).

**Figure 1.** Copper concentration ( $\mu\text{g g}^{-1}$ ) in aerial parts and roots of *Prosopis juliflora* plants exposed to different concentrations (0, 10, 50, and 100 mM) under hydroponic conditions during 48 h. Values are mean  $\pm$  SD ( $n = 4$ ). Significant differences among treatments are indicated by: \*  $P < 0.05$ ; \*\*  $P < 0.01$ .



**Table 1.** Bioaccumulation Factor (BAF) and Transportation Index ( $Ti$ ) of copper in *Prosopis juliflora* (SW) during 48 h exposure.

Doses (mM)	BAF		$Ti$
	Roots	Aerial part	Aerial part/Roots
10	208.33 $\pm$ 10.40 <sup>a</sup>	32.5 $\pm$ 9.01 <sup>a</sup>	0.16 $\pm$ 0.036 <sup>a</sup>
50	130.66 $\pm$ 3.54 <sup>b</sup>	39.5 $\pm$ 1.32 <sup>a</sup>	0.30 $\pm$ 0.002 <sup>b</sup>
100	66.41 $\pm$ 0.28 <sup>c</sup>	54.16 $\pm$ 2.36 <sup>c</sup>	0.81 $\pm$ 0.04 <sup>c</sup>

Data are means  $\pm$  S.D. ( $n = 4$ ). Those with different superscript letter (a, b and c) in the same column are significantly different ( $p < 0.05$ , turkey multiple comparison).

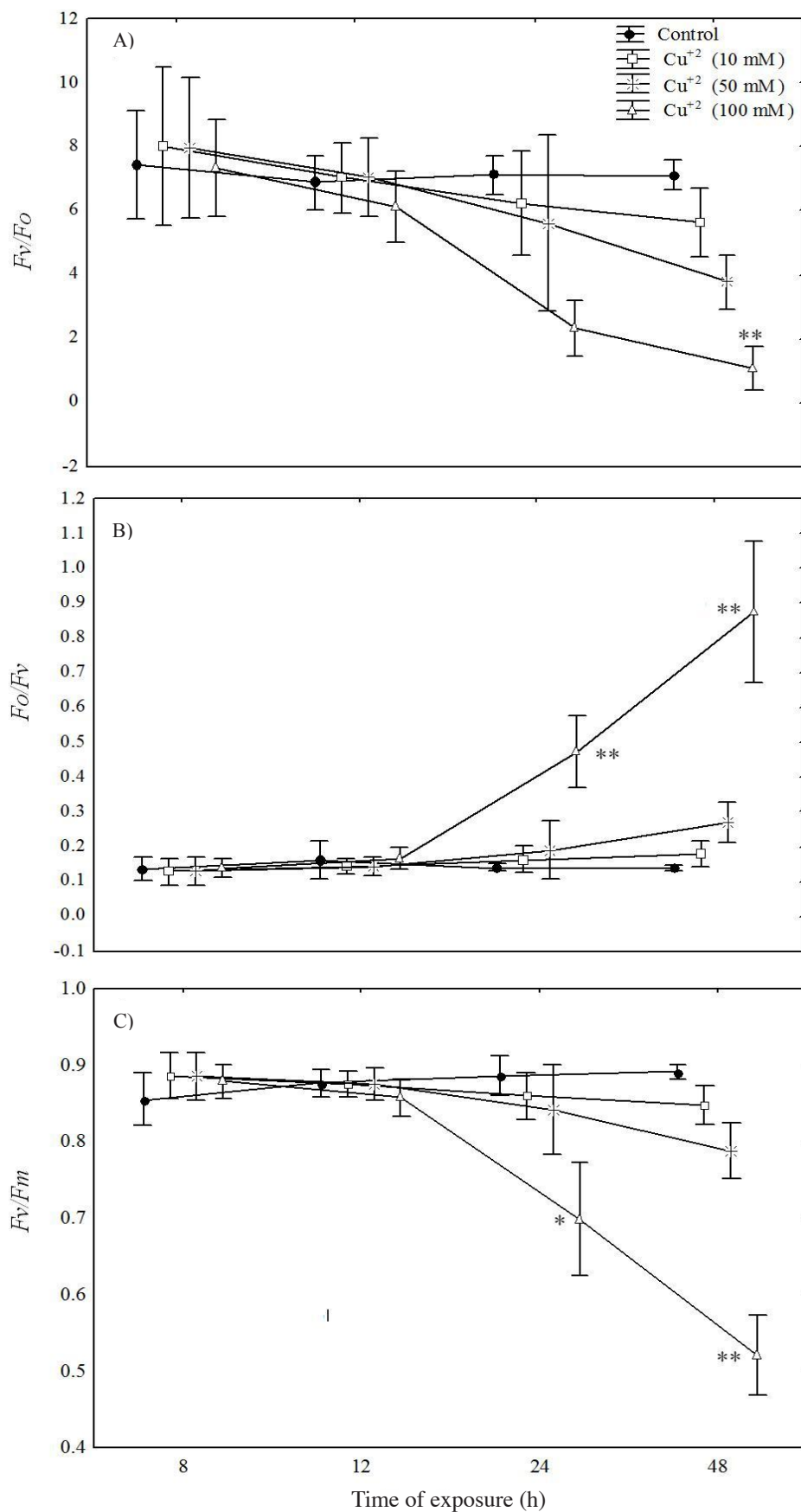
However, the aerial parts/roots  $Ti$  for plants exposed to 10 and 50 mM  $\text{Cu}^{+2}$  ( $0.16 \pm 0.036$ ;  $0.306 \pm 0.002$ , respectively) was very low, compared with the value showed by 100 mM  $\text{Cu}^{+2}$  ( $0.81 \pm 0.04$ ; Table 1).

**Chlorophyll fluorescence measurement.** The biophysical analysis of the leaf blades demonstrated that the size and number of active reaction center ( $Fv/Fo$ ) and the maximum photochemical efficiency of photosystem II ( $Fv/Fm$ ) showed a progressive drop with the greatest differences (relative to the control) being observed at the highest metal concentration at 24 h after exposure to metal (Figure 2A, C). On the other hand, the water splitting apparatus represented by ( $Fo/Fv$ ) was significantly higher only in plants treated with 100 mM  $\text{Cu}^{+2}$  during the 24 and 48 h after metal exposure (Figure 2B).

## Discussion

In order to maintain the absorption of essential metals within physiological limits and minimize their negative effects, many mesquite species have evolved a complex network of homeostatic mechanisms in roots that serve to control the uptake, accumulation, trafficking and detoxification of metals (Usha *et al.*, 2009; Zappala *et al.*, 2013). In this sense, our result revealed that the amount of copper accumulated in the roots of *Prosopis juliflora* indicated that this species have the ability to take up copper from the solution and to accumulate the metal in their roots, so they

**Figure 2.** Fluctuations of various fluorescent parameters for *Prosopis juliflora* after 48 h exposure to 10, 50 and 100 mM  $\text{Cu}^{2+}$ : **A)** the activity of photosystem II ( $F_v/F_o$ ); **B)** water splitting apparatus of PSII ( $F_o/F_v$ ), and **C)** the maximal photochemical yield of PSII in dark adapted leaves ( $F_v/F_m$ ). Significant differences among treatments are indicated by: \*  $P < 0.05$ ; \*\*  $P < 0.01$ .



could be used to phytostabilize metal-polluted groundwater (rhizofiltration). This result might be related to the presence of exclusion and sequestering processes that moderate metal uptake by roots and induce their accumulation in tissues (González-Mendoza *et al.*, 2007). The presence of higher values of  $\text{Cu}^{2+}$  found in roots and the low value for aerial parts/root translocation index found in *P. juliflora* exposure during 48 h, suggest a metal exclusion mechanism might be operating, as described by Baker (1981) and Boularbah *et al.* (2006).

In this context, *Prosopis juliflora* could not be considered as hyperaccumulator in view of less accumulation of Cu in roots and aerial parts. Because has lower concentrations of  $\text{Cu}^{+2}$  ( $0.531 \mu\text{g g}^{-1}$ ) than  $1000 \mu\text{g g}^{-1}$ , which are the limits prescribed for a hyperaccumulator (Boularbah *et al.*, 2006; Zappala *et al.*, 2014).

For evaluation of *Prosopis juliflora* plants as potential phytoremediators, low-cost, easy to carry and high throughput techniques are essential. In this sense, chlorophyll *a* fluorescence represent an excellent screening tool for evaluation of heavy metal stress in aerial parts of plants (Kummerová *et al.*, 2010; Hussain *et al.*, 2011). In the present study the changes observed in *Fv/Fm*, *Fv/Fo* and *Fo/Fv* (chlorophyll fluorescence parameters) as result of exposition with  $\text{Cu}^{+2}$  demonstrated a rapid inactivation of photosystem II (PSII) in *P. juliflora* leaves. The fluctuations in these parameters can be the result of  $\text{Ca}^{+2}$  substitution by  $\text{Cu}^{+2}$  in the catalytic center of PSII during photoactivation. Another possibility is that  $\text{Mn}^{+2}$  is replaced by  $\text{Cu}^{+2}$  from the water-splitting apparatus at the oxidizing side, resulting in a disruption of photosynthetic reactions (Küpper *et al.*, 2009). Additionally, the decline in the  $F_v/F_o$  ratio is indicative of a decline in the rate of photochemistry as the primary electron acceptor pool ( $Q_n$ ) became increasingly oxidized, or a reduction of the pool size of the primary electron acceptors associated with PSII activity (Krause and Weiss, 1991). It may indicate that *Fv/Fo* and *Fo/Fv* parameters are the most sensitive components in the photosynthetic electron transport chain in the presence of high copper concentration. In agreement to these results, it has been previously confirmed that the *Fv/Fo* and *Fo/Fv* parameter are the most sensitive component of PSII under undesirable conditions (González-Mendoza *et al.*, 2013). Similar results were observed by and Singh *et al.* (2013) who reported that higher concentrations of heavy metals induced one rapid inactivation of PSII in *Luffa acutangula* under hydroponics conditions.

Therefore the information provided by this short-term experiment in *Prosopis juliflora* showed that several physiological processes are activated, in which the copper uptake by roots and their accumulation in tissues play a central role. In this sense during short-term exposure to copper (48 h), sublethal effects such as alteration of the photosystem II, can be observed. It can thus be concluded that chlorophyll fluorescence parameters are sensitive as biomarkers of short-term copper exposures.

## Conclusion

The results presented in this research demonstrate that exposure of *Prosopis juliflora* plants to excess  $\text{Cu}^{+2}$  causes specific negative effects on the photosystem II. Such alteration may have critical physiological implications under conditions of  $\text{Cu}^{+2}$  toxicity in this mesquite species. Finally, the present study showed that *P. juliflora* is a promising prospect for heavy metals phytoremediation purposes occurring in arid and semi-arid climates in the northwest Mexico.

## Acknowledgements

The authors are particularly gratefully to E. Sauri (ITM, Mérida) for kindly allowing us to use ICP-OES and Conacyt Ciencia Basica 2013 (219750).

## Literature cited

- Alkorta I., Hernández-Allica J., Becerrill J.M., Amezcua I., Albizu I., Onaindia M. and Garbisu C. 2004. Chelate-enhanced phytoremediation of soils polluted with heavy metals. *Reviews in Environmental Science Biotechnology* 3:55-70.



- Baker A.J.M. 1981. Accumulators and excluders strategies in the response of seedlings to heavy metals. *Journal Plant Nutrition* **3**:643-654.
- Boularbah A., Schwartz C., Bitton G., Abouddar W., Ouhammou A. and Morel J.L. 2006. Heavy metal contamination from mining sites in South Morocco: 2. Assessment of metal accumulation and toxicity in plants. *Chemosphere* **63**:811-817.
- Carevic F.S. 2014. The role of ecophysiological studies in the genus *Prosopis*: implications for the conservation of drought-prone species. *IDESIA* **32**:77-81.
- Daesslé L.W., Lugo-Ibarra K.C., Tobschall H.J., Melo M., Gutiérrez-Galindo E.A., García-Hernández J. and Álvarez L. 2009. Accumulation of As, Pb, and Cu associated with the recent sedimentary processes in the Colorado Delta G. South of the United States Mexico boundary. *Archives Environmental Contamination Toxicology* **56**:680-692.
- Estrella-Gómez N., Mendoza-Cózatl D., Moreno-Sánchez R., González-Mendoza D., Zapata-Pérez O., Martínez-Hernández A. and Santamaría J.M. 2009. The Pb-hyperaccumulator aquatic fern *Salvinia minima* baker, responds to Pb<sup>2+</sup> by increasing phytochelatin synthesis via changes in smcps expression and in phytochelatin synthase activity. *Aquatic Toxicology* **91**:320-328.
- García-Hernández J., Sapozhnikova Y.V., Schlenk D., Mason A.Z., Hinojosa-Huerta O., Rivera-Díaz J.J., Ramos-Delgado N.A. and Sánchez-Bon G. 2006. Concentration of contaminants in breeding bird eggs from the Colorado River Delta, Mexico. *Environmental Toxicology Chemistry* **25**:1640-1647.
- Ghosh M. and Singh S.P. 2005. A review on phytoremediation of heavy metals and utilization of its by-products. *Applied Ecology and Environmental Research* **3**:1-18.
- González-Mendoza D. and Zapata-Pérez O. 2008. Mecanismos de tolerancia a elementos potencialmente tóxicos en plantas. *Boletín de la Sociedad Botánica de México* **82**:53-61.
- González-Mendoza D., Ceja-Moreno V., Gold-Bouchot G., Escobedo-Gracia Medrano R. M., Del-Rio M., Valdés-Lozano D. and Zapata-Pérez O. 2007. The influence of radical architecture on cadmium bioaccumulation in the black mangrove, *Avicennia germinans* L. *Chemosphere* **67**:330-334.
- González-Mendoza D., Espadas y Gil F., Escoboza-García F., Santamaría J.M. and Zapata-Pérez O. 2013. Copper stress on photosynthesis of black mangle (*Avicennia germinans*). *Anais da Academia Brasileira de Ciências* **85**:665-670.
- Hussain M.I. and Reigosa, M.J. 2011. Allelochemical stress inhibits growth, leaf water relations, PSII photochemistry, non-photochemical fluorescence quenching, and heat energy dissipation in three C3 perennial species. *Journal Experimental Botany* **62**:4533-4545.
- Krause G.H. and Weis E. 1991. Chlorophyll fluorescence and photosynthesis: the basics. *Annual Review of Plant Physiology and Plant Molecular Biology* **42**:313-349.
- Kummerová M., Zezulka Š., Kráľová K. and Masarovičová E. 2010. Effect of zinc and cadmium on physiological and production characteristics in *Matricaria recutita*. *Biologia Plantarum* **54**:308-314.
- Küpper H., Götz B., Mijovilovich A., Küpper F.C. and Meyer-Klaucke W. 2009. Complexation and toxicity of copper in higher plants I. Characterization of copper accumulations, and toxicity in *Crassula helmsii* as a new copper accumulator. *Plant Physiology* **151**:702-714.
- Machado M.D., Soares H.M.V.M. and Soares V.E. 2010. Removal of chromium, copper, and nickel from an electroplating effluent using a flocculent brewer's yeast strain of *Saccharomyces cerevisiae*. *Water Air and Soil Pollution* **212**:199-204.
- Michel-López C.Y., González-Mendoza D., Ruiz-Sánchez E. and Zamora-Bustillos R. 2014. Modifications of photochemical efficiency, cellular viability and total phenolic content of *Prosopis glandulosa* leaves exposed to copper. *Chemistry and Ecology* **30**:227-232.
- Pérez-García E.A., Meave J.A., and Cevallos-Ferriz. S.R.S. 2012. Flora and vegetation of the seasonally dry tropics in Mexico: origin and biogeographical implications. *Acta Botánica Mexicana* **100**:149-193.
- Shiferaw H., Teketay D., Nemomissa S. and Assefa F. 2004. Some biological characteristics that foster the invasion of *Prosopis juliflora* (Sw.) DC. at Middle Awash Rift Valley Area, north-eastern Ethiopia. *Journal of Arid Environmental* **58**:135-154.
- Singh R., Gautam N., Mishra A. and Gupta R. 2011. Heavy metals and living systems: An overview. *Indian Journal Pharmacology* **43**:246-253.
- Singh V.P., Srivastava P.K. and Prasad S.M. 2013. Nitric oxide alleviates arsenic-induced toxic effects in ridged Luffa seedlings. *Plant Physiology and Biochemistry* **71**:155-163.
- Usha B., Venkataraman G. and Parida A. 2009. Heavy metal and abiotic stress inducible metallothionein isoforms from *Prosopis juliflora* (SW) D.C. show differences in binding to heavy metals in vitro. *Molecular Genetics Genomics* **281**:99-108.
- Victor R., Pillay A. and Al-Minji S. 2007. Copper tolerance to germination in mesquite, a potential tree species for restoring mined-lands in Oman. *Journal of Agricultural Food and Environmental Science* **1**:1-8.
- Wong J.W.C. and Selvam A. 2006. Speciation of heavy metals during co-composting of sewage sludge with lime. *Chemosphere* **63**:980-986.

Received:  
December 23rd, 2014

Accepted:  
April 9th, 2015

- Zappala M.N., Ellzey J.T., Bader J., Peralta-Videa J.R. and Gardea Torresdey J. 2013. *Prosopis pubescens* (Screw Bean Mesquite) seedlings are hyperaccumulators of copper. *Archives Environmental Contamination Toxicology* **65**:212-223.
- Zappala M.N., Ellzey J.T., Bader J., Peralta-Videa J.R. and Gardea Torresdey J. 2014. Effects of Copper Sulfate on Seedlings of *Prosopis pubescens* (Screwbean Mesquite). *International Journal of Phytoremediation* **16**:1031-1041.