

## CYTOGENETICAL EFFECTS OF ALUMINUM ON ROOT MERISTEM CELLS OF *HELIANTHUS ANNUUS* L.

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**Abstract:** Different aluminum concentrations (5, 50, and 100  $\mu\text{M}$ ), as well as different exposure times (24, 48, and 72 h), were applied to investigate cytogenetical alterations in sunflower (*Helianthus annuus* L.) meristem cells. Nucleolin was also examined under aluminum stress using silver-staining method and indirect immunofluorescent method. Results showed that the mitotic index decreased progressively when either aluminum concentration or exposure time increased. C-mitosis, anaphase bridges, and chromosome stickiness were observed in the aluminum treated root tip cells. Some particulates containing the argyrophilic NOR-associated proteins were distributed in the nucleus of the root-tip cells, and the amount of this particulate material progressively increased with increasing exposure time. Finally, the nucleolar material was extruded from the nucleus into the cytoplasm. The result also indicated the nucleolar protein nucleolin was translocated from nucleolus to nucleoplasm and cytoplasm, that presence of 50  $\mu\text{M}$  of aluminum for 48 h, suggesting that aluminum disturbed rRNA synthesis and induced the abnormal cellular location of nucleolar protein. The strongly toxic effect of aluminum on the nucleolus can inhibit or stop mitosis, and consequently inhibit root growth.

**Keywords:** aluminum, cytogenetical alterations, mitosis, nucleolin, sunflower.

**Resumen:** Diferentes concentraciones de aluminio (5, 50 y 100  $\mu\text{M}$ ) y diferentes tiempos de exposición (24, 48 y 72 h) se aplicaron a células meristemáticas de girasol (*Helianthus annuus* L.), para investigar las alteraciones citogenéticas, en particular, dilucidar sobre la nucleolina. La nucleolina fue estudiada bajo el estrés por aluminio, utilizando el método de tinción con plata y el método de inmunofluorescencia indirecta. Los resultados mostraron que el índice mitótico disminuyó progresivamente a medida que aumentó la concentración de aluminio o el tiempo de exposición. Se observaron C-mitosis, puentes en la anafase y rigidez en los cromosomas en todas las células de la punta de la raíz tratadas con aluminio. Algunas partículas contienen argirófilas NOR asociadas a proteínas distribuidas en el núcleo de las células del ápice de la raíz. La cantidad de este material granulado aumentó progresivamente a medida que aumentó el tiempo de exposición. Finalmente, el material nucleolar se expulsó desde el núcleo hacia el citoplasma. El resultado también indica que, en presencia de 50  $\mu\text{M}$  de aluminio por 48 h, la proteína nucleolina se translocó desde el nucleolo al nucleoplasma y citoplasma, lo que sugiere que el aluminio altera la síntesis de ARNr e induce la localización celular anormal de la proteína nucleolar. El fuerte efecto tóxico de aluminio en el nucléolo puede inhibir o detener la mitosis, y por consiguiente, inhibir el crecimiento de la raíz.

**Palabras clave:** alteraciones citogenéticas, aluminio, girasol, mitosis, nucleolina.

Aluminum (Al), a light metal, represents 7% of the Earth's crust and is the third most abundant element after oxygen and silicon (Ma *et al.*, 2001). Aluminum toxicity is a major growth-limiting factor for crop production in acid soils (Liu *et al.*, 2008). Aluminum is a major component of acid toxicity in soils (Foy, 1984).

It is well known that roots are the primary target of aluminum phytotoxic (Doncheva *et al.*, 2005). Clarkson (1965) indicated that cell division as a primary site could induce root growth inhibition under Al stress. Matsumoto *et al.* (1976) and Morimura *et al.* (1978) supported the view of Al-induced inhibition of root cell proliferation as a primary

target for Al-toxicity. Horst (1995) indicated that apoplastic Al could lead to the inhibition of root cell elongation. Kollmeier *et al.* (2000) found that Al caused inhibition of basipetal auxin transport in maize roots and inhibited root cell elongation. Limited information is available about the effects of Al on root cell in *Helianthus annuus* L. For the present investigation, the effects of Al on root growth, cell division and nucleoli in *Helianthus annuus* were investigated in order to deeply understand the toxic mechanism of Al on root growth.

## Materials and methods

**Culture condition and aluminum treatment.** The seeds of *Helianthus annuus* were kindly provided by the Centre of Popularizing Agriculture Technique of Tianjin, China, and were used in the present investigation. The healthy and equal-sized seeds were chosen and soaked in aerated distilled water for 36 h before starting the experiments. They were germinated on wet gauze in containers at 25 °C, producing roots reaching about 1 cm length. After that, they were treated in Petri dishes with different concentrations of Al solutions (5, 50, and 100 µM; aluminum chloride, AlCl<sub>3</sub>) at pH 4.5 for 24, 48, and 72 h. Controls were grown on distilled water. The test liquids were changed regularly every 24 h. Seedlings were grown in a greenhouse equipped with supplementary 15 h light / 9 h dark at 18–20 °C diurnal cycle.

**Cytological study.** Fifteen root tips in each treatment group were washed with tap water and distilled water, and cut at 24, 48 and 72 h, respectively. They were cut and fixed in 3 parts 95% ethanol:2 parts 99.5% acetic acid for 2 h, and hydrolyzed in 5 parts 1 M hydrochloric acid:3 parts 95% ethanol:2 parts 99.8% acetic acid for 9 min at 60 °C. For the observation of changes in nucleolus, ten root tips were cut and squashed in 45% acetic acid, dried, and after 2 days stained with silver nitrate (Li *et al.*, 1990; Liu and Jiang, 1991). For the observation of chromosomal morphology, ten root tips selected from fifteen root tips were squashed in a carbol-fuchsin solution (Li, 1982). Mitotic index was used for the investigation; it means number of dividing cells per 500 observed cells (Fiskesjö, 1985). Data for root length were analyzed with standard statistical software (SigmaPlot 9.0).

**Indirect immunofluorescent microscopy.** Meristematic zones of root tips from control and seedlings treated with adding 50 µM Al for 48 h were cut and fixed with 4% (w/v) paraformaldehyde in phosphate-buffered saline (PBS, pH 7.0) for 2 h in darkness at room temperature and then they were washed with the same buffer. Meristematic cells were digested with a mixture of 2.5% cellulase and 2.5% pectolase at 37 °C, and then washed in PBS for three times. They were squashed on slides and extracted in freshly prepared 1% (v/v) Triton X-100 in PBS when slides dried. After

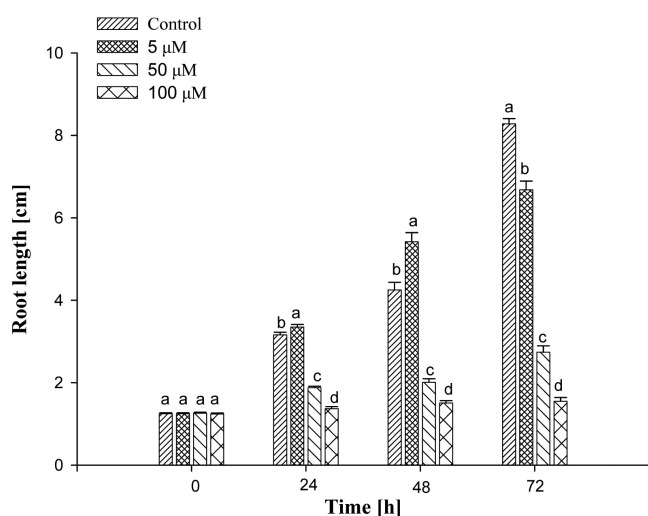
three washings in PBS, the cells were subsequently incubated with mouse primary antibodies respectively against nucleophosmin, nucleolin, or fibrillarin for 1 h at 37 °C or at 4 °C overnight in a moist, sealed chamber. After washing (3 × 10 min) in PBS, the cells were incubated with secondary antibodies for detection of the primary antibodies for 45 min in darkness at 37 °C. After repeated washing in PBS, nuclei were stained with 4', 6-diamidino-2-phenylindole (DAPI, Sigma) at a final concentration of 1 µg per 1 mL for 15 min at room temperature. After washing (3 × 10 min) in PBS, the cells were mounted in antifade mounting medium. The slides were stored at 4 °C in the dark until viewed. The immunofluorescent specimens were examined under a fluorescence microscope (Nikon, HB-10101AF) with the digital camera Pixera Pro 600CL, using violet (355–425 nm) and blue (450–490 nm) specific filters for proteins and nuclei respectively. Photographs were taken and images were processed with Adobe Photoshop 7.0 software.

Antibodies used in this investigation were as follows, nucleolin/C23, primary antibody: a mouse monoclonal antibody to nucleolin (Santa, SC-8031) at dilution 1:100; secondary antibody: TRITC-conjugated goat anti-mouse IgGs (Sigma, T5393) at dilution 1:50. TRITC was used for the detection of signal.

**Statistical analysis.** Analysis of variance of the data was done with SigmaPlot 8.0 software. For statistical analysis, one-way analysis of variance (ANOVA) and *t*-test were used to determine the significance at *P* < 0.05.

## Results

**Effects of Al on root growth.** The effects of Al on root growth of *Helianthus annuus* varied with the concentration



**Figure 1.** Effects of different concentrations of Al on root length of *Helianthus annuus*. Values with different letters differ significantly from each other (*n* = 15, *P* < 0.05).

**Table 1.** Effects of Al on cell division in the root tip cells of *Helianthus annuus*.

Time (h)	Concentration (μM)	Mitotic Index (%)	Number of cells	Normal dividing cells (%)		Anomalous dividing cell (%)			
				Metaphases	Anaphases	C-mitosis	Chromosome bridge	Chromosome stickiness	Rate of anomalous cells
24	Control	284	500	52.4	45.4	0.8	1.0	0.4	2.2
	5	324	500	49.6	44.8	2.2	2.4	1.0	5.6
	10	160	500	45.8	44.2	4.2	3.8	2.0	10.0
	100	134	500	42.8	44.0	4.4	4.8	4.0	13.2
48	Control	261	500	53.4	44.6	1.0	1.0	0.0	2.0
	5	176	500	47.2	44.0	3.0	3.6	2.2	8.8
	10	142	500	44.4	45.6	3.2	4.0	4.8	10.0
	100	94	500	46.6	40.0	3.6	5.2	4.6	13.4
72	Control	216	500	45.6	52.4	2.4	1.2	0.0	3.6
	5	134	500	48.6	40.2	4.4	4.2	2.6	11.2
	10	90	500	39.8	46.3	5.6	1.4	6.8	13.8
	100	64	500	43.4	17.8	14.0	3.9	21.0	38.9

and treatment time. There is a significant difference between treatment and control groups after 72 h of treatment, demonstrating a dose response (Figure 1). At 5 μM Al there was stimulative effect on root growth ( $P < 0.05$ ) after 24 to 48 h, whereas 50 μM and 100 μM Al inhibited root growth significantly ( $P < 0.05$ ) after 24 to 72 h.

The effects of Al on the morphology of the roots also varied with the different concentrations of aluminum chloride in solution. At 5 μM Al, the morphology of the roots was similar to control during the whole treatment (72 h), but with the increase of the concentration, the color of the root tips became yellow and the color was deeper. At 100 μM Al, the root tips were curved, swollen, cracked, brownish and stubby after 24 h.

**Effects of Al on cell division in root tip cells.** Observations of roots treated with different levels of Al revealed structural damage that was not observed in the root cells of control plants.

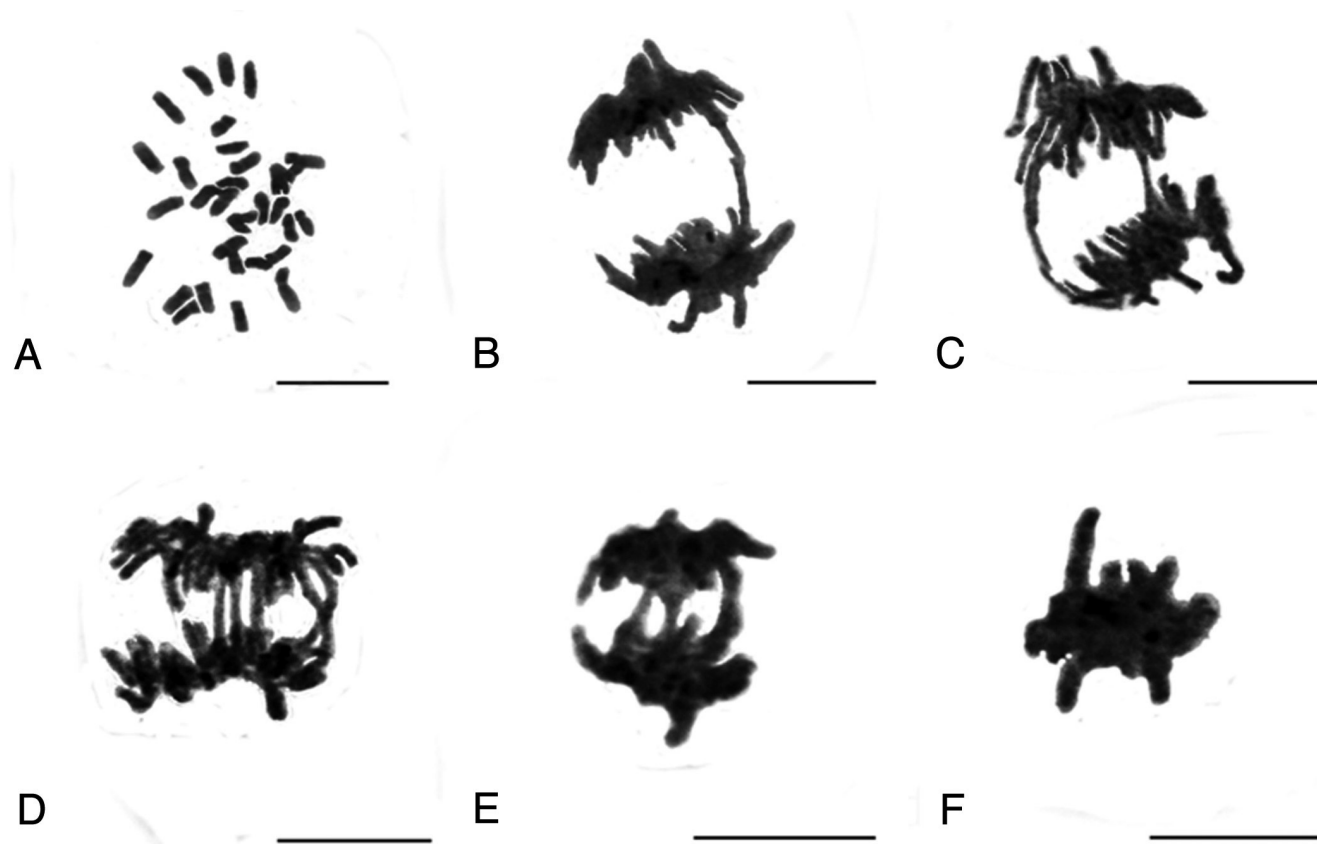
**Mitotic index.** The mitotic index reflects the frequency of cell division and it is regarded as an important parameter. The mitotic index decreased progressively as a function of increased Al concentration and exposure time (Table 1).

**Chromosomal aberrations.** The results showed that significant cytotoxic effects emerged after that the root tips of *Helianthus annuus* were treated with Al. Aluminum ion-induced mitotic aberration types included C-mitosis, chromosome bridges, and chromosome stickiness. C-mitosis was observed in the root tip cells of all treated groups after treatment with Al (Figure 2A). The frequency of cells with C-mitosis increased with increasing Al concentration and with prolonging duration of treatment (Table 1). Anaphase

bridges involving one or more chromosomes (Figure 2B-E) were found in the Al treated groups. The frequency of cells with chromosome bridges increased with increasing Al concentration during 48 h of treatment. Al could induce more chromosome stickiness bridges at 100 μM Al treatment group (Figure 2E). Chromosome stickiness reflects highly toxic effect, probably leading to cell death (Figure 2F). The frequency of cells with chromosome stickiness increased with increasing Al concentration (Table 1).

**Effects of Al on nucleolus.** Normally, the diploid nucleus of *Helianthus annuus* contains one or two nucleoli under normal circumstances (Figure 3A). The effects of Al on nucleoli varied with the different concentrations of aluminum chloride used. Three phenomena were observed after these treatments. Firstly, irregular nucleolus was found in the root tips exposed to 50 μM Al for 24 h (Figure 3B). Secondly, some tiny particles containing the argyrophilic NOR-associated (AgNOR) proteins gradually together with the main nucleolus/nucleoli in the nucleus of some root-tip cells increased with increasing of Al concentration and prolonging the duration of treatment (Figure 3C-F). Thirdly, in concentrations of 100 μM Al, the AgNOR protein was extruded from the nucleus into the cytoplasm (Figure 3G-H).

**Translocation of nucleolar proteins.** The toxic effects of Al on nucleolar protein nucleolin in the root tip cells were observed when compared with control. Using the red fluorescent signal to observe it, the study proved that small immunofluorescence spot of nucleolin was located in nucleolus exclusively in control cells (Figure 4A1-A3). In comparison with control nucleolin in some cells it was located in cytoplasm after the exposure of 50 μM Al for 48 h (Figure 4B3-C3). The nucleolar protein in the cytoplasm



**Figure 2.** Effects of Al on root tip cells division of *Helianthus annuus*. A) C-mitosis (5  $\mu$ M, 48 h). B-D) Chromosome bridges; B) 50  $\mu$ M, 24 h; C) 50  $\mu$ M, 24 h; D) 50  $\mu$ M, 48 h. E) Chromosome stickiness bridges (100  $\mu$ M, 24 h) F) Chromosome stickiness (100  $\mu$ M, 24 h). Scale bar = 10  $\mu$ m.

increases progressively with longer duration of the treatment (Figure 4E3).

## Discussion

Root growth inhibition upon exposure to Al has been used extensively as one of the most distinct and earliest symptoms of Al toxicity (Samuels *et al.*, 1997). Some reports revealed that Al caused the inhibition of root elongation by metal interference with cell division, including inducement of chromosomal aberrations and abnormal mitosis (Zhang, 1995; Qin *et al.*, 2010; Vardar *et al.*, 2011). The results in the present investigation indicated that significant root growth inhibition in *Helianthus annuus* seedlings exposed to 50  $\mu$ M and 100  $\mu$ M Al, which was in agreement with the early findings in *H. annuus* (Chakravarty and Srivastava, 1992; Kumar and Srivastava, 2006), *Vicia faba* L. (Zhang, 1995) and *Allium cepa* L. (Qin *et al.*, 2010).

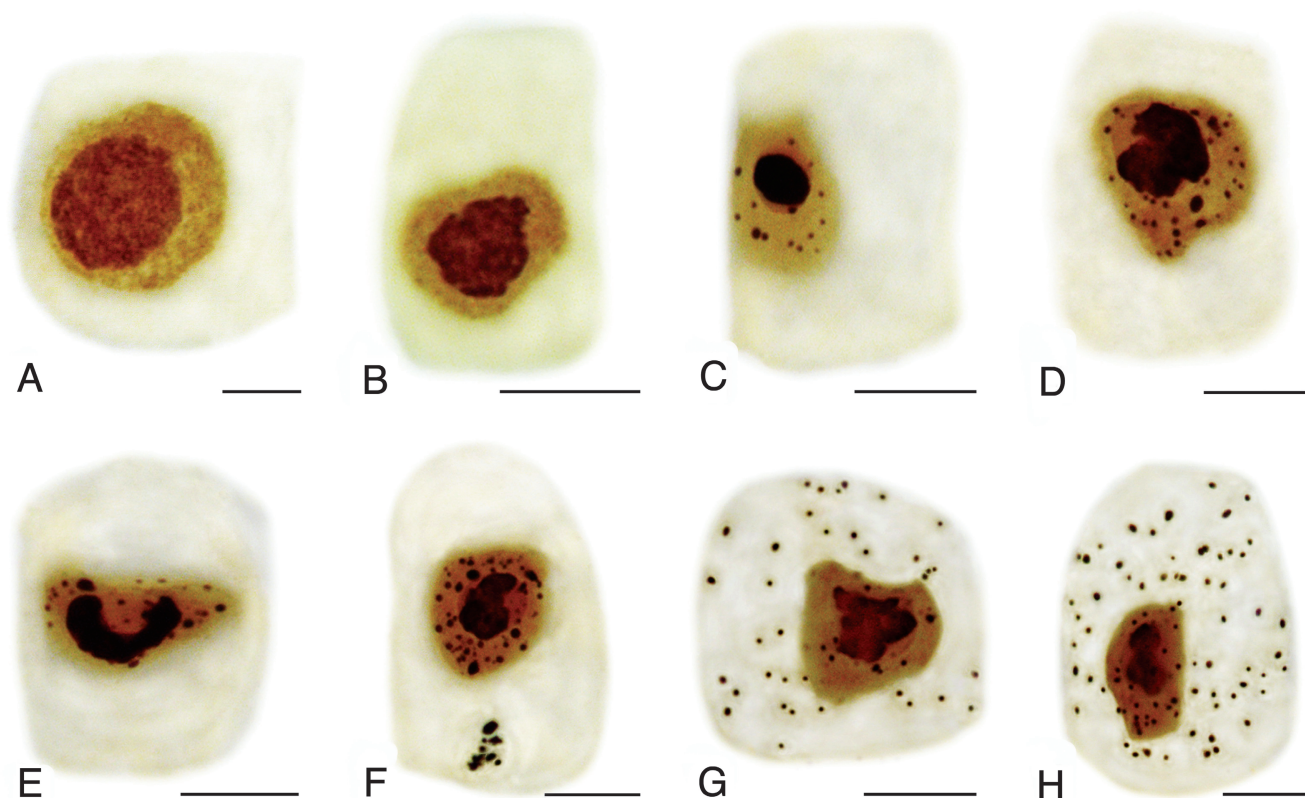
The silver staining technique has been widely applied in cytological studies aimed at understanding the nucleolar cycle and nucleolar organization in both animals and plants. Nucleolus is well known as the site of transcription of ribosomal genes and further transcript process (Shaw and Jordan, 1995), which contains a set of acidic, nonhistone

proteins that bind silver ions and are selectively visualized by silver method. In the present investigation, Al induced irregular nucleolus, fragmentation of particles of argyrophilic proteins scattered in the nuclei, and the material extruded from the nucleus into the cytoplasm. However, the results were, with a few differences, when comparison with the findings of Qin *et al.* (2010) and Zhang *et al.* (2009) for the effect of Al on *Allium cepa* and *Vicia faba*. For instance, (1) there is not so much nucleolar material released from nuclei into cytoplasm in *Helianthus annuus*, and (2) Al toxicity on the nucleoli in root tip cells of *A. cepa* and *V. faba* is stronger than one in *H. annuus*.

Amenós *et al.* (2009) indicated that Al was localized inside the nucleoli of root tip cells of Al sensitive maize. The evidence from the present work shows that once many particles of argyrophilic proteins are scattered in the nuclei, root growth in *H. annuus* can be inhibited, and once a large amount of nucleolar material is extruded from the nucleus into the cytoplasm, root growth is seriously inhibited or the seedlings died. The strongly toxic effect of Al on the nucleolus often inhibited or stopped mitosis.

Nucleolus is a multifunctional subnuclear territory that plays several important roles in cellular processes (Sirri *et al.*, 2008). Under stress conditions, many nucleolar proteins





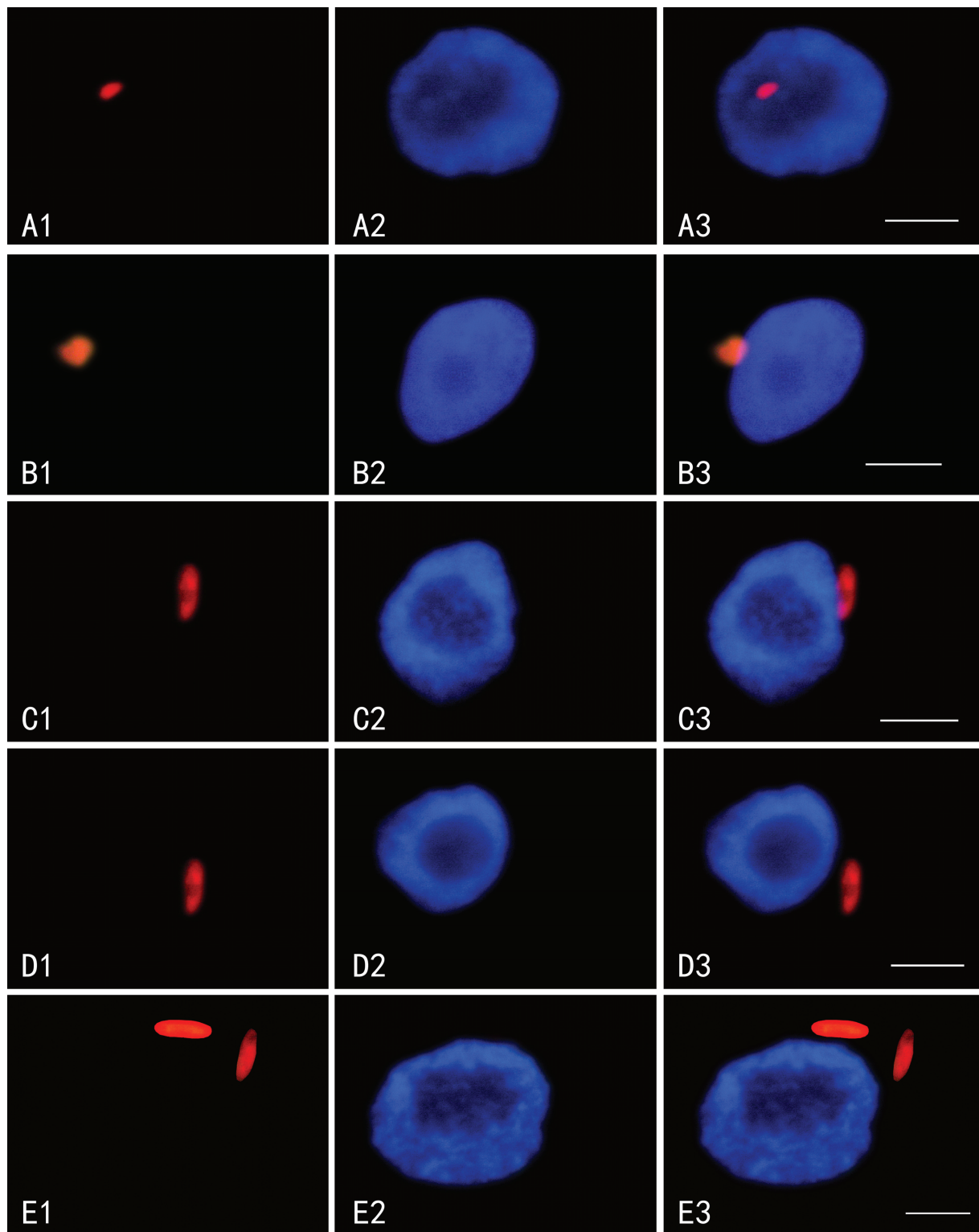
**Figure 3.** Effects of different concentrations of Al on nucleoli in root tip cells of *Helianthus annuus*. A) control cells, B) irregular nucleolus (50  $\mu$ M Al, 24 h), C) some tiny particles of silver-stained materials in nucleus (50  $\mu$ M Al, 24 h), D-F) Large amounts of silver stained materials in nucleus with increasing Al concentration and prolonging treatment time (50  $\mu$ M Al, 72 h), G-H) Silver-stained materials extruded from the nucleus into the cytoplasm (100  $\mu$ M Al, 48 h). Scale bar = 10  $\mu$ m.

change their locations and distribute disorderly in and out of nucleolus, furthermore alter their expressions and functions (Mayer *et al.*, 2005; Sun and Yi, 2008). Nucleolus is thought to be a sensor for cellular stress signals (Lewinska *et al.*, 2010).

Data from the present investigation showed that Al had toxic effects on nucleolus and nucleolar proteins. Nucleolin (C23) is a major Ag-NORs protein (Ginisty *et al.*, 1999). C23, the most abundant nucleolar phosphoproteins, is presented in the dense fibrillar component (DFC) and granular component (GC) of the nucleolus and involves in multiple steps of ribosome biogenesis (Chathoth *et al.*, 2009). Data from this work showed that C23 was detected exclusively at the nucleolar territory in control cells and in the treatment group. C23 was observed to be localized to the nucleoplasm and cytoplasm. C23 was demonstrated to be one of the main proteins in nucleolus and oxidative stress could induce the cleavage of it (Wang *et al.*, 2004). van der Aa *et al.* (2006) found that the nuclear pore complex (NPC) was the most important channel for nuclear material. The phenomenon that the nucleolar material was extruded from the nucleus into the cytoplasm might be explained that the proteins were affected after Al treatment, causing the NPC

to lose selectivity. C23 is an important multifunctional protein that has been implicated directly or indirectly in many metabolic processes and is one of the most abundant non-ribosomal proteins of the nucleolus (Bugler *et al.*, 1982). Sripinyowanich *et al.* (2013) suggest that C23 and ribosomal proteins affect ribosome biogenesis independently or that C23 may interact directly to promote plant growth. It was proved that the retention of nucleolar proteins within the nucleolus was closely related to active rRNA synthesis (Martin *et al.*, 2007; Chen and Jiang, 2004). Rubbi and Milner (2003) reported that inhibition of rRNA synthesis could affect interaction between nucleolar proteins and rRNA, which induced the relocation of nucleolar proteins. Therefore, we supposed that Al disturbed rRNA synthesis and induced the abnormal cellular location of nucleolar protein. The strongly toxic effect of Al on the nucleolus can inhibit or stop mitosis, and consequently inhibit root growth. The results can further confirm our previous observations (Liu and Jiang, 1991), and firstly identifies 'Al-structure' named by Fiskesjö (1983, 1990).

The data from these biomarkers can provide valuable information for monitoring and forecasting earlier effects of exposure to Al in real scenarios conditions.



**Figure 4.** Simultaneous location of nucleolin after the reaction with primary anti-nucleolin antibody and secondary antibody conjugated with TRITC (red) and DNA after the reaction with DAPI (blue) in the same single optical section, obtained with the fluorescence microscope. A1, B1, C1, D1, E1) nucleolin detection; A2, B2, C2, D2, E2) DNA detection; A3, B3, C3, D3, E3) merged image of “Nucleolin detection” and “DNA detection”. A1-A3) Showing that nucleolin was located in nucleolus in control cells (48 h). B1-B3) Showing that nucleolin was migrated from nucleolus to nucleoplasm in the cells exposed to 50  $\mu$ M Al for 48 h. C3-E3) Showing that in some cells larger amount of nucleolin appeared and was extruded from nucleolus into cytoplasm after the exposure of Al for 48 h. Scale bar = 10  $\mu$ m.

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