Ecology / Ecología

CHECKLIST OF THE ARBUSCULAR MYCORRHIZAL FUNGI OF OAXACA, AN IMPORTANT HOTSPOT OF BIODIVERSITY IN MEXICO

Listado de los hongos micorrízico arbusculares de Oaxaca, un punto caliente de biodiversidad importante en México

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Abstract

Background: Oaxaca, a southern State in Mexico, belongs to the Mesoamerican hotspot of biodiversity. The taxonomic and diversity knowledge of arbuscular mycorrhizal fungi (AMF) in Oaxaca State is scattered and accessibility to it is difficult.

Questions: Which AMF species have been identified in Oaxaca and which is their distribution?

Studied species: Arbuscular mycorrhizal fungi.

Study site: State of Oaxaca, Mexico.

Methods: We prepared an updated and annotated checklist of the AMF species with frequency of occurrence and the sampling sites where the species had been registered. All studies carried out previously (to date, 2022) were considered. Information on AMF taxa was available in scattered form in different publications in physical and electronic media, theses, projects, technical reports, and scientific papers.

Results: The checklist includes 78 species, distributed among 23 genera, 10 families, and four orders. The most diverse families were Glomeraceae (26 species), followed by Gigasporaceae (20) and Acaulosporaceae (15), whereas the most diverse genera were *Acaulospora* and *Glomus* (15 and eight species, respectively). The most frequent species were *Funneliformis geosporus*, *Acaulospora scrobiculata*, and *A. spinosa*. The highest species diversity has been reported under the rhizosphere of *Agave angustifolia*, *A. karwinskii*, and *Zea mays* (39, 36, and 34 species, respectively). A new AMF species was recently isolated from the rhizosphere of a microendemic agave.

Conclusions: The recorded AMF species of Oaxaca State confirms its position as one of the states with the highest AMF species diversity of Mexico.

Key words: Biodiversity, distribution, Glomeromycota, soil ecology, species richness, taxonomy.

Resumen

Antecedentes: Oaxaca, un estado sureño de México, pertenece al punto caliente Mesoamericano de biodiversidad. El conocimiento taxonómico y de diversidad de los hongos micorrízico arbusculares (HMA) en el estado de Oaxaca se encuentra disperso y poco accesible.

Preguntas: ¿Qué especies de HMA se han identificado en Oaxaca y cuál es su distribución?

Especies en estudio: Hongos micorrízico arbusculares.

Sitio de estudio: Estado de Oaxaca, México.

Métodos: Se elaboró un listado de las especies de HMA con la frecuencia de ocurrencia y los sitios de registro. Se consideraron los estudios realizados hasta 2022. La información estuvo disponible en forma dispersa en publicaciones en medios físicos y electrónicos: tesis, proyectos, reportes técnicos y artículos científicos.

Resultados: La lista incluye 78 especies, distribuidas en 23 géneros, 10 familias y cuatro órdenes. La familia más diversa fue Glomeraceae (26 especies), seguida por Gigasporaceae (20) y Acaulosporaceae (15), mientras que los géneros más diversos fueron *Acaulospora y Glomus* (15 y ocho especies, respectivamente). Las especies más frecuentes fueron *Funneliformis geosporus, Acaulospora scrobiculata y A. spinosa*. La diversidad más elevada de especies se ha reportado en la rizosfera de *Agave angustifolia, A. karwinskii y Zea mays* (39, 36, y 34 especies, respectivamente). Una nueva especie de HMA fue recientemente aislada en la rizosfera de un agave microendémico.

Conclusiones: Las especies de HMA registradas en Oaxaca confirman la posición de este estado como uno de los más altos en diversidad micorrízico arbuscular en México.

Palabras clave: Biodiversidad, distribución, Glomeromycota, ecología del suelo, riqueza de especies, taxonomía.

rbuscular mycorrhizal fungi (AMF) are mutualistic organisms that are associated with about 80 % of terrestrial plant species with respect to the arbuscular mycorrhiza interaction (Smith & Read 2008). Following Redecker *et al.* (2013), AMF belong to the Phylum Glomeromycota. Three hundred and forty-six AMF species have been described worldwide, and have been classified into five orders, 16 families, and 48 genera (Oehl *et al.* 2011b, Błaszkowski *et al.* 2015, Wijayawardene *et al.* 2020, Schüßler 2022). However, it is estimated that there are perhaps up to 1,000 AMF species according to modern molecular techniques (Kivlin *et al.* 2011, Öpik *et al.* 2013).

AMF are an essential component of rhizosphere and soil in the ecosystems, where they play active roles in the acquisition of water and nutrients (macro- and microelements) for their plant hosts, especially phosphorus, often considered the most limiting nutrient for plants in lowland tropical soils (Hodge & Fitter 2010, Smith & Smith 2011). AMF modulate the populations of several microorganisms, many of which are beneficial for plants (Artursson *et al.* 2006). AMF supply the plants with up to 80 % of nitrogen and 90 % of the phosphorus required (van der Heijden *et al.* 1998); AMF also improve survival, growth, reproduction, and resistance to biotic and abiotic stress in plants (Wagg *et al.* 2014, Van der Heijden *et al.* 2015).

The first list, to our knowledge, of AMF species tailored for Mexico reported the presence of only 44 species (Varela & Trejo 2001). Years later, in a second approach on AMF species diversity in the country, 95 species were registered (Montaño *et al.* 2012). The most current studies report the presence of about 160 species (*e.g.*, Chimal-Sánchez *et al.* 2018, Varela *et al.* 2019, Polo-Marcial *et al.* 2021), that is, one half (50 %) of AMF species described throughout the world have been reported in Mexico. In this context, Oaxaca State belongs to the Mesoamerican hotspot of biodiversity of vascular plants (Myers *et al.* 2000, García-Mendoza *et al.* 2004), in different ecosystems and agroecosystems including biosphere reserves and natural protected areas for preserving biodiversity and ecosystem services. However, Oaxaca, the most biodiverse state in the country (García-Mendoza *et al.* 2004) at present lacks an updated checklist of the AMF species it harbors. It is because most of the information generated is scattered and difficult to access (in theses and technical reports), and the vast majority has not been published in scientific journals, hindering access to information.

Considering that Oaxaca State in Mexico is a hotspot zone possessing an important plant species diversity (García-Mendoza *et al.* 2004), this contribution aimed to investigate the state-of-the-art of AMF species diversity and AMF distribution in the Oaxacan region in order to identify needs and future tasks. In this work, the AMF species richness was considered as the sole index of diversity. This report increases knowledge by providing information regarding trends in geographic and ecological distribution, and the association with ecosystems, crops, or particular plant species. This can allow for the comparison and contrasting of the knowledge concerning this fungal group in other geographical regions and under other conditions, which may provide elements to corroborate or refute different hypotheses. This checklist will continue to be updated when new AMF species registries are added. Additionally, even new species being described from this region will be included, while the information presented herein provides a state-of-the-art panorama of knowledge to date. These data are essential to establish conservation strategies and to better the use of natural resources, through the knowledge of what these species are and where they are located.

Materials and methods

Study area. Oaxaca is located at the southern part of Mexico between 15° 39'-18° 09' N latitude, and 93° 52'-98° 32' W longitude (Figure 1), and covers an area of 95,364 km², representing 4.8 % of the Mexican territory (García-Mendoza *et al.* 2004). The topography is exceptionally irregular because of constant tectonic movements. Elevation ranges from sea level along the coastal plains up to 3,000 m asl. The climatic and physiographic variation is reflected in the diversity of soils and vegetation communities that exist. According to economic activities and climate, eight regions are distinguished in Oaxaca: Cañada, Costa, Istmo, Mixteca, Papaloapan, Sierra Norte, Sierra Sur, and Valles Centrales (García-Mendoza *et al.* 2004). Nearly all vegetation types reported for Mexico



Figure 1. Location of different vegetation types in Oaxaca State, Mexico. The black circles represent the localities where the studies of arbuscular mycorrhizal fungi diversity have been carried out.

are represented in the state, ranging from tropical rainforest to xerophytic vegetation (Rzedowski 1978, García-Mendoza *et al.* 2004).

Database. We prepared an updated and annotated checklist of the AMF species with frequency (number of times the species has been recorded) of occurrence (%) and the sampling sites where the species had been registered. The AMF species richness was considered as the sole index of diversity. For this checklist, all studies carried out previously (to date, 2022) were considered. Information on AMF taxa was available in scattered form in different publications in physical and electronic media, theses (undergraduate degrees, M.Sc. degrees, and Ph.D. dissertations), projects, technical reports, and scientific papers (Varela & Trejo 2001, López-Guerra 2003, 2006, Guadarrama-Chávez et al. 2007, Carballar-Hernández 2009, Carballar-Hernández et al. 2013, Bautista-Cruz et al. 2014, Robles-Martínez et al. 2015, García-Velasco 2016, Cruz-Luna 2017, Hernández-Morales et al. 2017, Álvarez-Lopeztello et al. 2018, Chimal-Sánchez et al. 2018, Álvarez-Lopeztello et al. 2019a, b, Reyes-Jaramillo et al. 2019, de la Cruz-Ortiz et al. 2020, Chimal-Sánchez et al. 2021, Méndez-Matias et al. 2021). Prior to the elaboration of the checklist from all the sources consulted, only taxa determined at a specific level (with a binomial name) were employed, that is, aff. (affinis) and cf. (conformis) species, genera (i.e., Acaulospora spp.), species with invalid names and morphotypes were deleted. All names of AMF species included in the checklist and the names of the authorities were verified and updated according to the website of AMF Phylogeny of A. Schüßler (www.amf-phylogeny.com; accessed in June 2022). The taxonomic classification of the AMF species in the checklist follows Redecker et al. (2013), updated by A. Schüßler (www.amf-phylogeny.com), who collects all the taxonomical data produced by expert AMF taxonomists, and those available in different scientific journals.

Results

The checklist of AMF registered from Oaxaca includes 78 species, distributed in 23 genera, 10 families, and four orders (<u>Table 1</u>). The best-represented families with respect to their species richness were Glomeraceae (26), followed by Gigasporaceae (20) and Acaulosporaceae (15). The families with highest diversity of genera included Glomeraceae and Gigasporaceae (six, respectively), and Diversisporaceae (three). The taxonomic affinity of *Entrophospora infrequens* is unclear and it is indicated as *incertae sedis* (<u>Table 2</u>).

The genera richest in species were *Acaulospora*, *Glomus*, *Sclerocystis*, and *Gigaspora* (Figure 2A); these four genera concentrate 44 % of the species. The AMF highest in species diversity (in all of the plant communities studied) have been reported under the rhizospheres of *Agave angustifolia* (39), *A. karwinskii* (36), and *Zea mays* (33) (Figure 2B). *Funneliformis geosporus* was the most frequent species (100 %), followed by *Acaulospora scrobiculata* (82 %), and *A. spinosa* and *Entrophospora infrequens* (69 % each) (see frequency between parentheses in Table 1).

Table 1. Checklist of the arbuscular mycorrhizal fungi species registered from Oaxaca State, Mexico between 2001 and 2022, and the sampling site where the species has been reported (see abbreviations), and its frequency-of-observation between parentheses. Abbreviations: CF = ferns in cloud forest, MT = mine tailings, PAST = pasture, PP = pine plantations, SAV = savanna, STS= semiarid tropical scrub, SV (TDF) = secondary vegetation (tropical dry forest), TC = tomato and chili cultivars', TDF = tropical dry forest, TRF = tropical rainforest; A. an = *Agave angustifolia*, A. ka = *Agave karwinskii*, A. ma = *Agave marmorata*, A. nu = *Agave nussaviorum*, A. po = *Agave potatorum*, and Z. ma = *Zea mays*. Note: The genus *Claroideoglomus* in the family Claroideoglomeraceae has recently been described as a new order, Entrophosporales, in the Glomeromycota (Błaszkowski *et al.* 2022).

Arbuscular mycorrhizal fungi species

Glomerales

Claroideoglomeraceae

Claroideoglomus claroideum C. Walker & Schuessler; SV (TDF), TDF; A. an, A. ka, A. ma, A. po, Z. ma (44 %)

Claroideoglomus drummondii C. Walker & Schuessler; PAST, PP, SAV, TC, TRF (31 %)

Claroideoglomus etunicatum C. Walker & Schuessler; MT; A. an, A. ka, A. nu, Z. ma (31 %)

Claroideoglomus lamellosum C. Walker & Schuessler; Z. ma (6.3 %)

Glomeraceae

Funneliformis geosporus C. Walker & Schuessler; CF, MT, PAST, PP, SAV, STS, SV (TDF), TC, TDF, TRF; A. an, A. ka, A. ma, A. nu, A. po, Z. ma (100 %)

Funneliformis halonatus Oehl, G.A. Silva & Sieverd.; A. an, A. ka (12.6 %)

Funneliformis mosseae C. Walker & Schuessler; CF, MT, STS, TC; A. an, A. ka, A. ma, A. po, Z. ma (56 %)

Funneliformis verruculosus C. Walker & Schuessler; SV (TDF); A. an (12.6 %)

Glomus ambisporum G.S. Sm. & N.C. Schenck; A. nu (6.3 %)

Glomus glomerulatum Sieverd.; PAST, PP, SAV, TC, TRF (31 %)

Glomus globiferum Błaszk. & Chwat; A. nu (6.3 %)

Glomus macrocarpum Tul. & C. Tul.; A. an, A. ka, A. po (19%)

Glomus microcarpum Tul. & C. Tul.; A. ka, A. ma, A. po (19 %)

Glomus spinuliferum Sieverd. & Oehl; CF, MT, STS; A. an, A. ka, A. ma, A. po (44 %)

Glomus tenebrosum S.M. Berch; SV (TDF), TDF; Z. ma (19 %)

Glomus trufemii B.T. Goto, G.A. Silva & Oehl; PP, SAV, TRF (19 %)

Rhizophagus aggregatus C. Walker; SV (TDF); A. an (12.6 %)

Rhizophagus clarus C. Walker & Schuessler; CF, MT, STS, TC; A. an, A. po, Z. ma (44 %)

Rhizophagus fasciculatus C. Walker & Schuessler; MT, PAST, PP, SAV, TC, TRF; A. an, Z. ma (50 %)

Rhizophagus intraradices C. Walker & Schuessler; TC; A. an, A. ka, A. ma, A. nu, A. po, Z. ma (44 %)

Rhizoglomus microaggregatum Sieverd., G.A. Silva & Oehl; SV (TDF); A. an, Z. ma (19 %)

Sclerocystis liquidambaris C.G. Wu & Z.C. Chen; A. an, A. ka (12.6 %)

Sclerocystis clavispora Trappe; SV (TDF), TDF; A. an, A. ka, A. ma, A. po (38 %)

Sclerocystis dussi Höhn.; SV (TDF); Z. ma (12.6 %)

Sclerocystis rubiformis Gerd. & Trappe; CF, STS; A. an, A. ka, A. nu (31 %)

Sclerocystis sinuosa Gerd. & B.K. Bakshi; SV (TDF), TC; A. an, A. ka, Z. ma (31 %)

Sclerocystis taiwanensis C.G. Wu & Z.C. Chen; SAV (6.3 %)

Septoglomus constrictum Sieverd., G. A. Silva & Oehl; CF, MT, PAST, PP, SAV, TC, TRF; A. an, A. ka, Z. ma (63 %)

Septoglomus viscosum C. Walker, D. Redecker, D. Stille & A. Schüßler; TC; A. po, A. nu (19 %)

Diversisporales

Acaulosporaceae

Acaulospora delicata C. Walker, C.M. Pfeiff. & Bloss; SV (TDF), TDF; A. an, Z. ma (25 %)

Acaulospora denticulata Sieverd. & S. Toro; CF; A. po (12.6 %)

Acaulospora excavata Ingleby & C. Walker; A. an, Z. ma (12.6 %)

Acaulospora foveata Trappe & Janos; PAST, PP, SAV, SV (TDF), TRF; A. nu, Z. ma (44 %)

Acaulospora kentinensis Kaonongbua, J.B. Morton & Bever; MT, TC; Z. ma (19 %)

Acaulospora laevis Gerd. & Trappe; CF, PAST, PP, SAV, SV (TDF), TRF; Z. ma (44 %)

Acaulospora mellea Spain & N.C. Schenck; MT, PAST, PP, SAV, SV (TDF), TDF, TRF; A. an, Z. ma (56 %)

Acaulospora minuta Oehl, Tchabi, Hountondji, Palenz., I.C. Sánchez & G.A. Silva; A. ka (6.3 %)

Acaulospora morrowiae Spain & N.C. Schenck; MT, PAST, PP, SAV, SV (TDF), TC, TRF; A. an, A. ka, A. ma, A. po, Z. ma, (76%)

Acaulospora papillosa C.M.R. Pereira & Oehl; A. an, A. ka (12.6 %)

Acaulospora paulinae Błaszk.; A. ka, A. ma, A. po (19%)

Acaulospora reducta Oehl, B.T. Goto & C.M.R. Pereira; A. ka (6.3 %)

Acaulospora rehmii Sieverd. & S. Toro; A. an, A. ka, A. po, Z. ma (25 %)

Acaulospora scrobiculata Trappe; CF, PAST, PP, SAV, SV (TDF), TC, TDF, TRF; A. an, A. ka, A. ma, A. po, Z. ma (82 %)

Acaulospora spinosa C. Walker & Trappe; CF, MT, PAST, PP, SAV, TRF; A. an, A. ka, A. ma, A. po, Z. ma (69 %)

Diversisporaceae

Diversispora eburnea C. Walker & Schuessler; TC; Z. ma (12.6 %)

Diversispora spurca C. Walker & A. Schuessler; PAST, PP, SAV, TRF; A. an, A. ka (38 %)

Diversispora trimurales C. Walker & Schuessler; A. ka (6.3 %)

Oehlia diaphana Błaszk., Kozłowska & Dalpé; A. nu (6.3 %)

Redeckera fulva C. Walker & Schuessler; STS, SV (TDF), TDF; Z. ma (25 %)

Gigasporaceae

Cetraspora pellucida Oehl, F.A. Souza & Sieverd.; PAST, PP, SAV, STS, SV (TDF), TDF, TRF; A. ka, A. po, Z. ma (63 %)

Dentiscutata cerradensis Sieverd., F.A. Souza & Oehl; A. ka (6.3 %)

Dentiscutata erythropus C. Walker & D. Redecker; SV (TDF) (6.3 %)

Dentiscutata scutata (C. Walker & Dieder.) Sieverd., FA de Souza & Oehl; A. ka (6.3 %)

Dentiscutata reticulata Sieverd., F.A. Souza & Oehl; A. nu (6.3 %)

Gigaspora albida N.C. Schenck & G.S. Sm.; A. an, A. po (12.6 %)

Gigaspora candida Bhattacharjee, Mukerji, J.P. Tewari & Skoropad; A. an, A. ka, Z. ma (19 %)

Gigaspora decipiens I.R. Hall & L.K. Abbott; PAST, SAV, SV (TDF), TDF, TRF; A. an, A. ma, A. po, Z. ma (57 %)

Gigaspora gigantea Gerd. & Trappe; SV (TDF), TDF; A. ka (19 %)

Gigaspora margarita W.N. Becker & I.R. Hall; MT; A. po (12.6 %)

Gigaspora ramisporophora Spain, Sieverd. & N.C. Schenck; A. an (6.3 %)

Racocetra fulgida Oehl, F.A. Souza & Sieverd; A. an, A. ka, A. ma (19 %)
Racocetra gregaria Oehl, F.A. Souza & Sieverd.; PAST, PP, SAV, TRF; A. an (31 %)
Racocetra cromosomica ChimSánch., Varela & Montaño; A. ka (6.3 %)
Racocetra persica Oehl, F.A. Souza & Sieverd.; PP, SAV, TRF; A. ka (25 %)
Racocetra verrucosa Oehl, F.A. Souza & Sieverd.; A. an (6.3 %)
Scutellospora calospora C. Walker & F.E. Sanders; STS; A. ka (12.6 %)
Scutellospora dipurpurescens J.B. Morton & Koske; PAST, PP, SAV, SV (TDF), TC, TRF; A. ma, A. po, Z. ma (57 %)
Scutellospora scutata (C. Walker & Dieder.); A. ka (6.3 %)
Sieverdingia tortuosa Błaszk., Niezgoda & B.T. Goto; A. an (6.3 %)
Pacisporaceae
Pacispora scintillans C. Walker, Vestberg & Schuessler; SV (TDF) (6.3 %)
Archaeosporales
Ambisporaceae
Ambispora appendicula C. Walker; PAST, PP, SAV, TRF; A. an, Z. ma (38 %)
Ambispora gerdemannii C. Walker, Vestberg & Schuessler; MT, SV (TDF); Z. ma (19 %)
Archaeosporaceae
Archaeospora schenckii Walker & Schuessler; PAST, PP, SAV, TRF; A. an (31 %)
Archaeospora undulata (Sieverd.) Sieverd., G.A. Silva, B. Goto & Oehl; CF (6.3 %)
Paraglomerales
Paraglomeraceae
Paraglomus bolivianum Oehl & G.A. Silva; A. an, A. ka (12.6 %)
Paraglomus occultum J.B. Morton & D. Redecker; Z.ma (6.3 %)
Sacculosporaceae
Sacculospora baltica; Oehl, Palenz., I.C. Sánchez, B.T. Goto, G.A. Silva & Sieverd.; A. ka (6.3 %)
Insertae sedis
Entrophospora infrequens R.N. Ames & R.W. Schneid.; CF, MT, STS, SV (TDF), TC, TDF; A. an, A. ka, A. ma, A. po, Z. ma (69 %)

Thirty-four species (of the number of total species) were detected in natural ecosystems (without apparent human intervention, *i.e.*, tropical rainforest, cloud-forest ferns, semi-arid tropical scrub, and tropical dry forest) (see <u>Table 1</u>). Ecosystems with highest species diversity were tropical rainforest (20), followed by cloud-forest ferns and tropical dry forest (12 species, respectively); in contrast, semi-arid tropical scrub demonstrated lowest species diversity (8). In these ecosystems, the genera richest in species were *Acaulospora* (9) and *Glomus* (4). The genera *Dentiscutata, Pacispora, Paraglomus*, and *Rhizoglomus* were absent. *Funneliformis geosporus* (100 %), *A. laevis, A. scrobiculata*, and *E. infrequens* (75 % each, respectively) were the most frequent species (Table 1).

In agroecosystems, 77 species were detected (Table 1). Agroecosystems with the highest species diversity were *A. angustifolia* (39), *A. karwinskii* (36), and *Z. mays* (33); in contrast, *A. nussaviorum* exhibited lowest species diversity (10). A new AMF species, *Racocetra cromosomica*, was recently isolated from the rhizosphere of wild *A. karwinskii*. The genera richest in species were *Acaulospora* (15), *Glomus* (8), and *Gigaspora* (6). *Funneliformis geosporus* (100 %), *A. morrowiae* (91.67 %), and *A. scrobiculata* (83.33 %) were the most frequent species. *Archaeospora undulata* was absent (see Table 1).

Discussion

The checklist of AMF species reported for Oaxaca State (Mexico) reveals high Glomeromycota species richness (<u>Table 1</u>). Ninety percent of AMF families registered for Mexico by Varela *et al.* (2019) and Polo-Marcial *et al.* (2021),



Figure 2. A. Number of arbuscular mycorrhizal fungi species by genera, and B number of arbuscular mycorrhizal fungi species by sampling site in Oaxaca State, Mexico.

and 74 % of families worldwide (Błaszkowski *et al.* 2015, Wijayawardene *et al.* 2020) are represented in the region. More than one half of genera reported for the world and for Mexico (> 64 % and > 80 %, respectively), and nearly one quarter of the species described throughout the world (22 %) and one half (49 %) of AMF species registered for Mexico (Montaño *et al.* 2012, Chimal-Sánchez *et al.* 2018, Varela *et al.* 2019, Polo-Marcial *et al.* 2021, Schüßler 2022) are present in Oaxaca, surpassing the AMF species diversity registered for Chiapas, Tabasco, and Veracruz, which are the Mexican states with the highest fungal diversity reported to date (Varela *et al.* 2008, Rodríguez-Morelos *et al.* 2014, Salgado-García *et al.* 2014, Trejo *et al.* 2016, Bertolini *et al.* 2018, Posada *et al.* 2018). However, this trend may change, as the study and knowledge of AMF species diversity increase in the country, including tropical and subtropical regions and some other states (*i.e.*, the State of Mexico) where AMF species diversity has been poorly explored. This fungal diversity represents increasing efforts for the state of Oaxaca that, up to a few years ago, had been scarcely explored, with only two species recorded (*Acaulospora foveata* and *Sclerocystis clavispora*) at the time of the first inventory of AMF species from Mexico (Varela & Trejo 2001).

Nonetheless, in recent years (last 7 years), there has been an increase in the number of studies on AMF taxonomy and diversity at several ecosystems and agroecosystems of Oaxaca (Figure 1). This is mainly due to a search for alternatives to facilitate the nutrition, growth, and development of plants, as well as to improve their tolerance to environmental stress conditions. Despite the increase observed and efforts made in recent years (Figure 3) to acquire knowledge on AMF species diversity in Oaxaca State, it is still far from well-known and possibly hosts a much higher number than is currently known. This Mexican state presents varied topographic, climatic, edaphic conditions, and high vegetational heterogeneity and vast flora diversity with much endemisms (García-Mendoza *et al.* 2004). The latter could harbor a high and still unknown wealth of AMF species, possibly many of these new species.

Considering the role of plants as umbrella species, hotspots also function as hotspots for other, less well-studied organisms, such as associated soil fungi (Stork & Habel 2014). In this context, the first AMF was recently described as being from Oaxaca, that is, *Racocetra cromosomica* (Chimal-Sánchez *et al.* 2021), isolated from the rhizosphere of *Agave karwinskii*, a microendemic plant. In addition, *Septoglomus mexicanum*, another new AMF, was isolated from the Tehuacán-Cuicatlán Biosphere Reserve, a semi-arid ecosystem shared between the states of Puebla and Oaxaca (Chimal-Sánchez *et al.* 2020). The application of molecular strategies using DNA from the roots or from the rhizospheric soil of selected plants will aid in rapidly increasing knowledge on the AMF community present in Oaxacan systems.



Figure 3. Number of taxonomic studies of arbuscular mycorrhizal fungi diversity conducted every 5 years in Oaxaca State, Mexico.

Glomeraceae, Acaulosporaceae, and Gigasporaceae, with highest AMF species richness in Oaxaca State (Table 2), are the very families that also have the highest numbers of species registered for the entire country (Montaño *et* al. 2012, Varela et al. 2019, Polo-Marcial et al. 2021). According to the richness of AMF recorded in various ecosystems and agroecosystems, Glomeraceae and Acaulosporaceae tend to be the most frequent families (Öpik et al. 2010, Cofré et al. 2019, Varela et al. 2019) and possess a considerable proportion of generalist species, which has been evidenced by their existence under varied and contrasting conditions. In the most recent reviews, to our knowledge, on AMF in Mexico, species from both families prevail (Varela et al. 2019, Polo-Marcial et al. 2021). There is a record of AMF in 19 botanical families, where Cactaceae, Fabaceae and Poaceae have the highest number of analyzed AMF species. No members of Acaulosporaceae, to our knowledge, have been recorded in Asclepiadaceae, Asteraceae, Caricaceae, Chenopodiaceae, Fouqueriaceae, Meliaceae and Selaginaceae. Two reasons can explain these results: the great lack of knowledge on the diversity of AMF associated with particular plant species from various botanical groups, and the lack of Taxonomists studying Glomeromycota. Polo-Marcial et al. (2021) have suggested that one possible explanation for the high species diversity of these families at both the state and national levels is that they are generally more associated with the Neotropical than with the Nearctic region. Acaulospora, Glomus, Sclerocystis, and Gigaspora were the genera with highest specific diversity in Oaxaca State (Figure 2B), as observed in certain other Mexican states (Varela et al. 2008, Rodríguez-Morelos et al. 2014, Salgado-García et al. 2014, Trejo et al. 2016, Bertolini et al. 2018, Posada et al. 2018), and also in Mexico (Montaño et al. 2012, Varela et al. 2019, Polo-Marcial et al. 2021). Acaulospora and Glomus are usually the most frequent and those with greatest AMF diversity in studies on diversity around the world; they are cited as generalist and are capable of existing under environmental and edaphic changeable conditions (Bhardwaj et al. 1997, Carvalho et al. 2003, Escudero & Mendoza 2005). Gigaspora has generally been associated with low-disturbance ecosystems (Alvarez-Lopeztello et al. 2019b) due to its lengthy life cycle and its association with high soil moisture and organic-matter content.

Funneliformis geosporus was the most frequent species and it was present in all vegetation types studied to date (2022), indicating that it could be a generalist species with low environmental specificity, as suggested by Carballar-Hernández *et al.* (2013) and Guadarrama *et al.* (2014). The same situation could be true for *A. scrobiculata*, one of the most cited AMF species in the Mexican territory (Montaño *et al.* 2012), and the second most frequently registered AMF in Oaxaca. In contrast, other important species, such as *Claroideoglomus lamellosum* [recently included in the genus *Entrophospora* (Błaszkowski *et al.* 2022)], *Glomus ambisporum*, *G. globiferum*, *G. macrocarpum*, *Rhizoglomus microaggregatum*, *Sclerocystis liquidambaris*, *S. taiwanensis*, *Acaulospora minuta*, *A. reducta*, *Archaeospora undulata*, *Oehlia diaphana*, *Dentiscutata reticulata*, *Gigaspora candida*, *G. ramisporophora*, *Racocetra fulgida*, *R.*

verrucosa, *Scutellospora calospora*, *Pacispora scintillans*, and *Paraglomus occultum* have been registered only once for Oaxaca State (Table 1). A possible explanation for the low frequency of these species is that no long-term studies (more than 2 years), to our knowledge, have been carried out and only one-half of the studies have considered seasonality (rainy and dry). Therefore, it is recommended that future research consider long-term studies and seasonality. The lack of long-term studies could lead to this AMF species being considered a rare species; for example, *A. minuta* and *A. reducta* are known for Mexico as deriving only from Oaxaca (Chimal-Sánchez *et al.* 2018). If there is no, to our knowledge, further exploration, and number of records, it is valid to consider the species as rare; these are little recorded species, even in the places where they were described (Oehl *et al.* 2011a, Pereira *et al.* 2015). These AMF species were found under the rhizosphere of *A. karwinskii* and *A. angustifolia*; therefore, this rareness may be the result of the lack of registries; the latter prompts continuing to conduct further AMF taxonomic studies in Oaxaca State, in that there remain many ecosystems that have been little explored, such as tropical rainforest, cloud forest, and natural grasslands (Figure 1). In addition, fewer than 1 % of the rhizospheric soils of wild and cultured plant species in Oaxaca have been explored.

Mirzaei & Moradi (2017) identified a high correlation between plant diversity and AMF diversity in semi-arid forests: to greater diversity of plants, greater diversity of AMF and vice versa. This allowed corroborating that a greater plant diversity favors higher spore density and higher root colonization intensity, which contribute to maintaining the inoculum potential in the soil, although these parameters could depend on the hosts involved and the edaphic conditions. This should undoubtedly be considered to improve conservation strategies by directly influencing the maintenance of the systems. It is difficult to estimate how many AMF species could be expected in Oaxaca based on the number of plant species; only about 346 species of Glomeromycota are known on the Earth and there is abundant evidence of their low specificity. Öpik *et al.* (2009) pointed out that generalist plant species share the majority of AMF species, including generalists; these authors propose that the relationships are functional rather than specific between plants and AMF.

Mesoamerica is the center-of-origin of maize and agave. Highest AMF-species richness has been found under the rhizosphere of *Agave karwinskii*, *A. angustifolia*, and *Zea mays* and this unequivocally coincides with that these are the most studied species (at the rhizospheric level) and are those entertaining the greatest significant economic and cultural interest in Oaxaca State. However, in many places, these agave species are not cultivated, and the AMF species diversity that they can harbor under natural conditions is unknown. Another important point that could exert a

Families	Genera (%)	Species (%)
Claroideoglomeraceae	1 (4.35)	4 (5.13)
Glomeraceae	6 (26.09)	26 (33.33)
Acaulosporaceae	1 (4.35)	15 (19.23)
Diversisporaceae	3 (13.04)	5 (6.41)
Gigasporaceae	6 (26.09)	20 (25.64)
Pacisporaceae	1 (4.35)	1 (1.28)
Ambisporaceae	1 (4.35)	2 (2.56)
Archaeosporaceae	1 (4.35)	2 (2.56)
Paraglomeraceae	1 (4.35)	2 (2.56)
Sacculosporaceae	1 (4.35)	1 (1.28)
Incertae sedis	1 (4.35)	1 (1.28)

Table 2. Families, genera, and species of arbuscular mycorrhizal fungi registered in Oaxaca State, Mexico between 2001 and 2022. In parentheses, the proportion of families, genera, and species regarding their total is depicted. Note: The genus *Claroideoglomus* in the family Claroideoglomeraceae has recently been described as a new order, Entrophosporales, in the Glomeromycota (Błaszkowski *et al.* 2022).

strong negative influence on the AMF diversity associated with these cultivated species is the use of chemical fertilizers and pesticides. It has been demonstrated that chemical pesticides reduce abundance of AMF in agricultural soils (Rivera-Becerril *et al.* 2017). Nevertheless, the study of the AMF species diversity of these plant species is far from being completed. There remain many sites at which and environmental conditions under which they are cultivated but have not yet been explored.

The isolation and integration of collections of native AMF comprise an important task in order to preserve these bioresources and to explore their potential use for the production of important plants (*i.e.*, maize and agave), or for other purposes such as the restoration of degraded soils and ecosystems. In this regard, some AMF species that have been isolated and propagated include *Acaulospora mellea*, *Acaulospora scrobiculata*, *Acaulospora spinosa*, *Claroideoglomus drummondii* [recently included in the genus *Entrophospora* (Błaszkowski *et al.* 2022)], *Diversispora spurca*, *Funneliformis geosporus*, and *Septoglomus constrictum*. These AMF species have been inoculated in plant species that can become established in early successional stages typical of humid tropical forests (Álvarez-Lopeztello *et al.* 2021). *Agave* species, able to adapt to nutrient-deficient soils, high temperatures, and high solar radiation, are candidates for the restoration of arid habitats (Cervera Herrera *et al.* 2018). Among the AMF reported in this study, species that respond to isolation and propagation could be used in restoration, particularly those that have shown specific benefits to plants. In Mexico, monospecies inocula have been tested to mycorrhize plants for restoration purposes (Hernández-Cuevas *et al.* 2011, Quiñones-Aguilar *et al.* 2019) or consortia, including species recorded in Oaxaca (Carballar-Hernández *et al.* 2018, Quiñones-Aguilar *et al.* 2019). The recommendation is to work with AMF consortia from environments in ecogeographical regions similar to those to be restored; consortia from stressed environments are particularly attractive.

In conclusion, the findings revealed in this study to date suggest that Oaxaca is the state with the highest AMF species diversity in the country, in that Oaxaca harbors one-half of the species reported for Mexico, reaffirming its place as the most diverse AMF species-rich state. However, this may change, as knowledge and long-term studies on the AMF species diversity increase; different regions have been poorly explored with respect to AMF species diversity. Likewise, our findings also exhibit a lack of studies on AMF diversity; therefore, there is a need to continue exploring the state of Oaxaca, especially in terms of vegetation types such as tropical rainforest, natural grasslands, cloud forest, and temperate forest, plants of economic and cultural interest (maize, tomatoes, and beans), and plants that are endemic that have not been studied (cacti and legumes). The presence of species poorly registered or registered for the first time in Mexico demonstrate the importance of Oaxaca as a reservoir of AMF biodiversity. The number of works on AMF in transformed and non-transformed ecosystems place Oaxaca as one of the best studied states in Mexico.

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Literature cited

- Álvarez-Lopeztello J, Hernández-Cuevas LV, del Castillo RF, Robles C. 2018. Second world record of *Glomus tru-femii* (Glomeromycota: Fungi), an arbuscular mycorrhizal fungus from a Mexican savanna. *Revista Mexicana de Biodiversidad* 89: 298-300. DOI: <u>https://doi.org/10.22201/ib.20078706e.2018.1.2101</u>
- Álvarez-Lopeztello J, Hernández-Cuevas LV, del Castillo RF, Robles C. 2019a. Diversity of arbuscular mycorrhizal fungi associated with *Brachiaria brizantha* pastures in lowlands of Oaxaca, Mexico. *Grassland Science* 65: 197-201. DOI: <u>https://doi.org/10.1111/grs.12224</u>
- Álvarez-Lopeztello J, del Castillo RF, Robles C, Hernández-Cuevas LV. 2019b. Spore diversity of arbuscular my-

corrhizal fungi in human-modified neotropical ecosystems. *Ecological Research* **34**: 394-405. DOI: <u>https://doi.org/10.1111/1440-1703.12004</u>

- Alvarez-Lopeztello J, del Castillo RF, Robles C, Hernández-Cuevas LV. 2021. Arbuscular mycorrhizal fungi improve the growth of pioneer tree species of tropical forests on savanna and tropical rainforest soils under nursery conditions. *Scientia Fungorum* 51: e1296. DOI: <u>https://doi.org/10.33885/sf.2021.51.1296</u>
- Artursson V, Finlay RD, Jansson JK. 2006. Interactions between arbuscular mycorrhizal fungi and bacteria and their potential for stimulating plant growth. *Environmental Microbiology* 8: 1-10. DOI: <u>https://doi.org/10.1111/j.1462-2920.2005.00942.x</u>
- Bautista-Cruz AA, Montaño NM, Camargo-Ricalde SL, Pacheco L. 2014. Hongos micorrizógenos arbusculares y nutrimentos del suelo asociados a cuatro especies de helechos en dos ecosistemas de Oaxaca, México. *Revista Chapingo. Serie Ciencias Forestales y del Ambiente* 20: 199-212.
- Bertolini V, Montaño NM, Chimal-Sánchez E, Varela-Fregoso L, Gómez-Ruiz J, Martínez-Vázquez JM. 2018. Abundancia y riqueza de hongos micorrizógenos arbusculares en cafetales de Soconusco, Chiapas, México. *Revista de Biología Tropical* **66**: 91-105. DOI: <u>https://doi.org/10.15517/rbt.v66i1.27946</u>
- Bhardwaj S, Dudeja SS, Khurana A. 1997. Distribution of vesicular-arbuscular mycorrhizal fungi in the natural ecosystem. *Folia Microbiologica* 42: 589-594. DOI: <u>https://doi.org/10.1007/BF02815471</u>
- Błaszkowski J, Chwat G, Góralska A. 2015. Acaulospora ignota and Claroideoglomus hanlinii, two new species of arbuscular mycorrhizal fungi (Glomeromycota) from Brazil and Cuba. Mycological Progress 14: 1-11. DOI: https://doi.org/10.1007/s11557-015-1042-2
- Błaszkowski J, Sánchez-García M, Niezgoda P, Zubek S, Fernández F, Vila A, Al-Yahya'ei MN, Symanczik S, Milczarski P, Malinowski R, Cabello M, Goto BT, Casieri L, Malicka M, Bierza W, Magurno F. 2022. A new order, Entrophosporales, and three new *Entrophospora* species in Glomeromycota. *Frontiers in Microbiology* 13: 962856. DOI: <u>https://doi.org/10.3389/fmicb.2022.962856</u>
- Carballar-Hernández S. 2009. Variación temporal de la diversidad de hongos de micorriza arbuscular y el potencial micorrízico en especies silvestres de Agave en Oaxaca. MSc Thesis. Instituto Politécnico Nacional.
- Carballar-Hernández S, Hernández-Cuevas LV, Montaño NM, Ferrera-Cerrato R, Alarcón A. 2018. Species composition of native arbuscular mycorrhizal fungal consortia influences growth and nutrition of poblano pepper plants (*Capsicum annuum* L.). *Applied Soil Ecology* **130**: 50-58. DOI: <u>https://doi.org/10.1016/j.apsoil.2018.05.022</u>
- Carballar-Hernández S, Palma-Cruz FJ, Hernández-Cuevas L, Robles C. 2013. Arbuscular mycorrhizal potential and mycorrhizal fungi diversity associated with *Agave potatorum* Zucc. in Oaxaca, Mexico. *Ecological Research* 28: 217-226. DOI: <u>https://doi.org/10.1007/s11284-012-1008-7</u>
- Carvalho LM, Correia PM, Ryel RJ, Martins-Loucão MA. 2003. Spatial variability of arbuscular mycorrhizal fungal spores in two natural plant communities. *Plant and Soil* **251**: 227-236.
- Cervera Herrera JC, Leirana-Alcocer JL, Navarro Alberto JA. 2018. Factores ambientales relacionados con la cobertura de *Agave angustifolia* (Asparagaceae) en el matorral costero de Yucatán, México. *Acta Botanica Mexicana* **124**: 75-84. DOI: <u>https://doi.org/10.21829/abm124.2018.1252</u>
- Chimal-Sánchez E, Reyes-Jaramillo I, Camargo-Ricalde SL, Varela L, Salmerón Castro JY, Montaño NM. 2021. *Racocetra cromosomica* sp. nov. from Oaxaca, Mexico. *Mycotaxon* **136**: 615-626. DOI: <u>https://doi.org/10.5248/136.615</u>
- Chimal-Sánchez E, Reyes-Jaramillo I, Salmerón-Castro JY, Vázquez-Pérez N, Varela-Fregoso L. 2018. Cuatro nuevos registros de hongos micorrizógenos arbusculares (Glomeromycota) asociados con Agave karwinskii y A. angustifolia (Agavaceae) de Oaxaca, México. Acta Botanica Mexicana 125: 173187. DOI: <u>https://doi.org/10.21829/ abm125.2018.1356</u>
- Chimal-Sánchez E, Senés-Guerrero C, Varela L, Montaño NM, García-Sánchez R, Pacheco A, Montaño-Arias SA, Camargo-Ricalde SL. 2020. Septoglomus mexicanum, a new species of arbuscular mycorrhizal fungi from semiarid regions in Mexico. Mycologia 112: 121-132. DOI: <u>https://doi.org/10.1080/00275514.2019.1671147</u>
- Cofré MN, Soteras F, Iglesias MR, Velázquez S, Abarca C, Risio L, Ontivero E, Cabello MN, Domínguez LS, Lugo MA. 2019. Biodiversity of arbuscular mycorrhizal fungi in South America: a review. *In*: Pagano MC, Lugo MA,

eds. Mycorrhizal fungi in South America. Cham: Springer, pp. 49-72. DOI: https://doi.org/10.1007/978-3-030-15228-4_3

- Cruz-Luna DN. 2017. Características morfométricas de hongos formadores de micorrizas asociados a Agave nussaviorum Garcia-Mend. de Oaxaca. BSc Thesis. Universidad Autónoma Benito Juárez de Oaxaca.
- de la Cruz-Ortiz ÁV, Álvarez-Lopeztello J, Robles C, Hernández-Cuevas LV. 2020. Tillage intensity reduces the arbuscular mycorrhizal fungi attributes associated with *Solanum lycopersicum*, in the Tehuantepec Isthmus (Oax-aca), Mexico. *Applied Soil Ecology* **149**: 103519. DOI: <u>https://doi.org/10.1016/j.apsoil.2020.103519</u>
- Escudero V, Mendoza R. 2005. Seasonal variation of arbuscular mycorrhizal fungi in temperate grasslands along a wide hydrologic gradient. *Mycorrhiza* **15**: 291-299. DOI: <u>https://doi.org/10.1007/s00572-004-0332-3</u>
- García-Mendoza AJ, Ordoñez MJ, Briones-Salas M. 2004. *Biodiversidad de Oaxaca*. Mexico City: Instituto de Biología Universidad Nacional Autónoma de México, Fondo Oaxaqueño para la Conservación de la Naturaleza, World Wildlife Fund. ISBN: 970-32-2045-2
- García-Velasco GP. 2016. Diversidad y potencial fitorremediador de hongos de micorriza arbuscular en especies vegetales que colonizan jales mineros. MSc Thesis. Instituto Politécnico Nacional.
- Guadarrama P, Castillo S, Ramos-Zapata JA, Hernández-Cuevas LV, Camargo-Ricalde SL. 2014. Arbuscular mycorrhizal fungal communities in changing environments: the effects of seasonality and anthropogenic disturbance in a seasonal dry forest. *Pedobiologia* 57: 87-95. DOI: <u>https://doi.org/10.1016/j.pedobi.2014.01.002</u>
- Guadarrama-Chávez P, Camargo-Ricalde SL, Hernández-Cuevas L, Castillo-Argüero S. 2007. Los hongos micorrizógenos arbusculares de la región de Nizanda, Oaxaca, México. *Boletín de la Sociedad Botánica de México* 81: 131-137. DOI: <u>https://doi.org/10.17129/botsci.1770</u>
- Hernández-Cuevas L, Guerra-De la Cruz V, Santiago-Martínez G, Cuatlal-Cuahutencos P. 2011. Propagación y micorrización de plantas nativas con potencial para restauración de suelos. *Revista Mexicana de Ciencias Forestales* 2: 87-96. DOI: <u>https://doi.org/10.29298/rmcf.v2i7.566</u>
- Hernández-Morales JL, López-Sánchez C, Robles-Pérez C, Palma-Cruz FJ. 2017. Nuevos reportes de hongos micorrízicos arbusculares en Agave potatorum, en la Mixteca Oaxaqueña. Revista Mexicana de Agroecosistemas 4: 37-47
- Hodge A, Fitter AH. 2010. Substantial nitrogen acquisition by arbuscular mycorrhizal fungi from organic material has implications for N cycling. *Proceedings of the National Academy of Sciences of the USA* **107**: 13754-13759. DOI: <u>https://doi.org/10.1073/pnas.1005874107</u>
- Kivlin SN, Hawkes CV, Treseder KK. 2011. Global diversity and distribution of arbuscular mycorrhizal fungi. Soil Biology and Biochemistry 43: 2294-2303. DOI: <u>https://doi.org/10.1016/j.soilbio.2011.07.012</u>
- López-Guerra I. 2003. Aislamiento e identificación de hongos micorrízicos arbusculares nativos de suelos de los Valles Centrales de Oaxaca. BSc Thesis. Universidad Autónoma Benito Juárez de Oaxaca.
- López-Guerra I. 2006. Hongos de micorriza arbuscular en diferentes sistemas de producción de maguey mezcalero (Agave angustifolia Haw.) en Oaxaca. MSc Thesis. Instituto Politécnico Nacional.
- Méndez-Matias A, Robles C, Hernández-Cuevas LV. 2021. Hongos micorrizógenos arbusculares asociados con el cultivo de maíz en regiones con sequía en Oaxaca. Agrociencia 55: 19-35. DOI: <u>https://doi.org/10.47163/agrociencia.v55i1.2345</u>
- Mirzaei J, Moradi M. 2017. Relationships between flora biodiversity, soil physiochemical properties and arbuscular mycorrhizal fungi (AMF) diversity in a semi-arid forest. *Plant Ecology and Evolution* **150**: 151-159. DOI: <u>https:// doi.org/10.5091/plecevo.2017.1249</u>
- Montaño NM, Alarcón A, Camargo-Ricalde SL, Hernández-Cuevas LV, Álvarez-Sánchez J, González-Chávez MC, Gavito M, Sánchez-Gallén I, Ramos-Zapata J, Guadarrama P, Maldonado-Mendoza IE, Castillo-Argüero S, García-Sánchez R, Trejo D, Ferrera-Cerrato R. 2012. Research on arbuscular mycorrhizae in Mexico: an historical synthesis and future prospects. *Symbiosis* 57: 111-126. DOI: https://doi.org/10.1007/s13199-012-0184-0
- Myers N, Mittermeier RA, Mittermeier CG, da Fonseca GAB, Kent J. 2000. Biodiversity hotspots for conservation priorities. *Nature* **403**: 853-858. DOI: <u>https://doi.org/10.1038/35002501</u>
- Oehl F, Palenzuela J, Sánchez-Castro I, Hountondji F, Tchabi A, Lawouin L, Barea JM, Coyne D, Alves da Silva

G. 2011a. *Acaulospora minuta*, a new arbuscular mycorrhizal fungal species from sub-Saharan savannas of West Africa. *Journal of Applied Botany and Food Quality* **84:** 213-218.

- Oehl F, Sieverding E, Palenzuela J, Ineichen K, da Silva GA. 2011b. Advances in Glomeromycota taxonomy and classification. *IMA Fungus* 2: 191-199. DOI: <u>https://doi.org/10.5598/imafungus.2011.02.02.10</u>
- Öpik M, Davison J, Moora M, Zobel M. 2013. DNA-based detection and identification of Glomeromycota: the virtual taxonomy of environmental sequences. *Botany* **92**: 135-147. DOI: <u>https://doi.org/10.1139/cjb-2013-0110</u>
- Öpik M, Metsis M, Daniell TJ, Zobel M, Moora M. 2009. Large-scale parallel 454 sequencing reveals host ecological group specificity of arbuscular mycorrhizal fungi in a boreonemoral forest. *New Phytologist* 184: 424-437. DOI: <u>https://doi.org/10.1111/j.1469-8137.2009.02920.x</u>
- Öpik M, Vanatoa A, Vanatoa E, Moora M, Davison J, Kalwij JM, Reier Ü, Zobel M. 2010. The online database MaarjAM reveals global and ecosystemic distribution patterns in arbuscular mycorrhizal fungi (Glomeromycota). New Phytologist 188: 223-241. DOI: https://doi.org/10.1111/j.1469-8137.2010.03334.x
- Pereira CMR, Goto BT, Alves da Silva DK, Almeida de Ferreira AC, de Souza FA, Alves da Silva G, Maia LC, Oehl F. 2015. Acaulospora reducta sp. nov. and A. excavata -two glomeromycotan fungi with pitted spores from Brazil. Mycotaxon 130: 983-995. DOI: <u>https://doi.org/10.5248/130.983</u>
- Polo-Marcial MH, Lara-Pérez LA, Goto BT, Margarito-Vista X, Andrade-Torres A. 2021. Glomeromycota in Mexico, a country with very high richness. Sydowia 74: 33-63. DOI: <u>https://doi.org/10.12905/0380.sydowia74-2021-0033</u>
- Posada RH, Sánchez de Prager M, Heredia-Abarca G, Sieverding E. 2018. Effects of soil physical and chemical parameters, and farm management practices on arbuscular mycorrhizal fungi communities and diversities in coffee plantations in Colombia and Mexico. *Agroforestry Systems* 92: 555-574. DOI: <u>https://doi.org/10.1007/s10457-016-0030-0</u>
- Quiñones-Aguilar EE, Hernández Cuevas LV, López Pérez L, Rincón Enríquez G. 2019. Efectividad de hongos micorrízicos arbusculares nativos de rizósfera de *Agave* como promotores de crecimiento de papaya. *Terra Latinoamericana* 37: 163-174. DOI: <u>https://doi.org/10.28940/terra.v37i2.397</u>
- Redecker D, Schüßler A, Stockinger H, Stürmer SL, Morton JB, Walker C. 2013. An evidence-based consensus for the classification of arbuscular mycorrhizal fungi (Glomeromycota). *Mycorrhiza* 23: 515-531. DOI: <u>https://doi.org/10.1007/s00572-013-0486-y</u>
- Reyes-Jaramillo I, Chimal-Sánchez E, Salmerón-Castro JY, Vázquez-Pérez N, Varela-Fregoso L. 2019. Comunidad de hongos micorrizógenos arbusculares (Glomeromycota) asociada con agaves mezcaleros de Oaxaca y su relación con algunas propiedades edáficas. *Revista Mexicana de Biodiversidad* 90: e902777. DOI: <u>https://doi.org/10.22201/ib.20078706e.2019.90.2777</u>
- Rivera-Becerril F, van Tuinen D, Chatagnier O, Rouard N, Béguet J, Kuszala C, Soulas G, Gianinazzi-Pearson V, Martin-Laurent F. 2017. Impact of a pesticide cocktail (fenhexamid, folpel, deltamethrin) on the abundance of Glomeromycota in two agricultural soils. *Science of the Total Environment* 577: 84-93. DOI: <u>https://doi.org/10.1016/j.scitotenv.2016.10.098</u>
- Robles-Martínez ML, Rivera-Becerril F, Hernández-Cuevas LV, Ortega-Larrocea MP, Robles C. 2015. *Spatial and seasonal variations in mycorrhizal potential and diversity of arbuscular mycorrhizal fungi in Agave angustifolia Haw. agroecosystems in Oaxaca, Mexico*. Technical report. Instituto Politécnico Nacional.
- Rodríguez-Morelos VH, Soto-Estrada A, Pérez-Moreno J, Franco-Ramírez A, Díaz-Rivera P. 2014. Arbuscular mycorrhizal fungi associated with the rhizosphere of seedlings and mature trees of *Swietenia macrophylla* (Magnoliophyta: Meliaceae) in Los Tuxtlas, Veracruz, Mexico. *Revista Chilena de Historia Natural* 87: 1-10. DOI: https://doi.org/10.1186/s40693-014-0009-z

Rzedowski J. 1978. Vegetación de México. Mexico City: Limusa. ISBN: 9789681800024

Salgado-García S, Castelán-Estrada M, Jiménez-Jerónimo R, Gómez-Leyva JF, Osorio-Miranda M. 2014. Diversidad de hongos micorrícicos arbusculares en suelos cultivados con caña de azúcar en la región de la Chontalpa, Tabasco. *Revista Mexicana de Micología* **40**: 7-16.

Schüβler A. 2022. *Glomeromycota: species list*. <u>http://www.amf-phylogeny.com/</u> (accessed June 20, 2022).

Smith SE, Read DJ. 2008. Mycorrhizal symbiosis. London: Academic Press and Elsevier. ISBN: 9780123705266

- Smith SE, Smith FA. 2011. Roles of arbuscular mycorrhizas in plant nutrition and growth: new paradigms from cellular to ecosystem scales. *Annual Review of Plant Biology* 62: 227-250. DOI: <u>https://doi.org/10.1146/annurev-arplant-042110-103846</u>
- Stork NE, Habel JC. 2014. Can biodiversity hotspots protect more than tropical forest plants and vertebrates? *Journal of Biogeography* **41**: 421-428. DOI: <u>https://doi.org/10.1111/jbi.12223</u>
- Trejo D, Barois I, Sangabriel-Conde W. 2016. Disturbance and land use effect on functional diversity of the arbuscular mycorrhizal fungi. *Agroforestry Systems* **90** (2): 265-279. DOI: <u>https://doi.org/10.1007/s10457-015-9852-4</u>
- Van der Heijden MG, Klironomos JN, Ursic M, Moutoglis P, Streitwolf-Engel R, Boller T, Wiekmen A, Sanders IR. 1998. Mycorrhizal fungal diversity determines plant biodiversity, ecosystem variability and productivity. *Nature* 396: 69-72. DOI: <u>https://doi.org/10.1038/23932</u>
- Van der Heijden MG, Martin FM, Selosse MA, Sanders IR. 2015. Mycorrhizal ecology and evolution: the past, the present, and the future. *New Phytologist* 205: 1406-1423. DOI: <u>https://doi.org/10.1111/nph.13288</u>
- Varela FL, Estrada-Torres A, Álvarez-Sánchez FJ, Sánchez-Gallén I. 2008. Catálogo ilustrado de hongos micorrizógenos arbusculares de la Reserva de la Biósfera de Los Tuxtlas. Mexico City: Universidad Nacional Autónoma de México. ISBN: 9786072000919
- Varela FL, Hernández-Cuevas LV, Chimal-Sánchez E, Montaño NM. 2019. Diversidad taxonómica de hongos micorrizógenos arbusculares citados de México. In: Álvarez SJ, Rodríguez GP, Alarcón A. eds. Biodiversidad de microorganismos de México. Importancia, aplicación y conservación. Mexico City: Universidad Nacional Autónoma de México. ISBN: 978-607-30-1596-7
- Varela FL, Trejo D. 2001. Los hongos micorrizógenos arbusculares como componentes de la biodiversidad del suelo en México. Acta Zoológica Mexicana (Número Especial) 00: 39-51. DOI: <u>https://doi.org/10.21829/azm.2001.8401845</u>
- Wagg C, Bender SF, Widmer F, van der Heijden MG. 2014. Soil biodiversity and soil community composition determine ecosystem multifunctionality. *Proceedings of the National Academy of Sciences of the USA* 111: 5266-5270. DOI: <u>https://doi.org/10.1073/pnas.1320054111</u>
- Wijayawardene NN, Hyde KD, Al-Ani LKT, Tedersoo L, Haelewaters D, Rajeshkumar KC, Zhao RL, Aptroot A, Leontyev DV, Saxena RK, Tokarev YS, Dai DQ, Letcher PM, Stephenson SL, Ertz D, Lumbsch HT, Kukwa M, Issi IV, Madrid H, Phillips AJL, Selbmann L, Pfliegler WP, Horváth E, Bensch K, Kirk PM, Kolaříková K, Raja HA, Radek R, Papp V, Dima B, Ma J, Malosso E, Takamatsu S, Rambold G, Gannibal PB, Triebel D, Gautam AK, Avasthi S, Suetrong S, Timdal E, Fryar SC, Delgado G, Réblová M, Doilom M, Dolatabadi S, Pawłowska JZ, Humber RA, Kodsueb R, Sánchez-Castro I, Goto BT, Silva DKA, de Souza FA, Oehl F, da Silva GA, Silva IR, Błaszkowski J, Jobim K, Maia LC, Barbosa FR, Fiuza PO, Divakar PK, Shenoy BD, Castañeda-Ruiz RF, Somrithipol S, Lateef AA, Karunarathna SC, Tibpromma S, Mortimer PE, Wanasinghe DN, Phookamsak R, Xu J, Wang Y, Tian F, Alvarado P, Li DW, Kušan I, Matočec N, Mešić A, Tkalčec Z, Maharachchikumbura SSN, Papizadeh M, Heredia G, Wartchow F, Bakhshi M, Boehm E, Youssef N, Hustad VP, Lawrey JD, Santiago AL-CMA, Bezerra JDP, Souza-Motta CM, Firmino AL, Tian Q, Houbraken J, Hongsanan S, Tanaka K, Dissanayake AJ, Monteiro JS, Grossart HP, Suija A, Weerakoon G, Etayo J, Tsurykau A, Vázquez V, Mungai P, Damm U, Li QR, Zhang H, Boonmee S, Lu YZ, Becerra AG, Kendrick B, Brearley FQ, Motiejūnaitė J, Sharma B, Khare R, Gaikwad S, Wijesundara DSA, Tang LZ, He MQ, Flakus A, Rodriguez-Flakus P, Zhurbenko MP, McKenzie EHC, Stadler M, Bhat DJ, Liu JK, Raza M, Jeewon R, Nassonova ES, Prieto M, Jayalal RGU, Erdoğdu M, Yurkov A, Schnittler M, Shchepin ON, Novozhilov YK, Silva-Filho AGS, Gentekaki E, Liu P, Cavender JC, Kang Y, Mohammad S, Zhang LF, Xu RF, Li YM, Dayarathne MC, Ekanayaka AH, Wen TC, Deng CY, Pereira OL, Navathe S, Hawksworth DL, Fan XL, Dissanayake LS, Kuhnert E, Grossart HP, Thines M. 2020. Outline of Fungi and fungi-like taxa. Mycosphere 11: 1060-1456. DOI: https://doi.org/10.5943/mycosphere/11/1/8

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